

Abattoir Survey on Prevalence of Bovine Fasciolosis in Guduru and Abay Chomaan Districts

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Abstract: A cross sectional study was conducted from April, 2014 to December, 2014 with the aim of determining the prevalence of bovine *fasciolosis* and its risk factors in Abaychoman and Guduru municipal abattoir. A post mortem examination was used as diagnostic tool for this research to detect adult liver fluke found in the bile ducts of slaughtered animals. Parasites were identified into species level by using standard parasitological techniques. Following post-mortem examination, a total of 384 indigenous cattle slaughtered during the study period were considered and 125(32.6%) were found to be positive for one or both of the *Fasciola* species. Out of this positive samples; *F.gigantica* was prevalent (47.2%) when compared to *F.hepatica* (39.2%) or mixed infections (13.6%). In this study, different variables such as body condition, origin, sex and age were considered. Accordingly, the prevalence of the parasites in different body conditions was found to be: 60% (135) in poor, 23.5% (170) in medium and 5.1% (79) in good body conditioned animals and was statistical significant ($P=0.000$). The infection rates in different origins of animals were also analyzed and a prevalence of 28.5% (179) and 36% (205) was recorded in Abaychomen and Guduru respectively. Based on the sexes of animal slaughtered, the prevalence rate of bovine *fasciolosis* in male was 34.3% (216) and in female 30.4% (168). The risk factors such as origin of animals and sex were not statistically significant. The prevalence of fasciolosis in different age groups of animals slaughter in both abattoirs was compared and found to be 15.3% (131) in young and 41.5% (253) in adult animals and it was statistically significant ($P=0.00$). Univariate logistic regression analysis of ages of animals confirmed that the probability of old animals to be infected by fasciolosis was 3.9 times higher when compared to young animals ($OR=3.9$; $CI= 2.29$; 6.74). Finally, the abattoir based prevalence recorded in this study area suggests that a detailed epidemiological study as well as assessment of the overall economic loss incurred due to fasciolosis is required to implement systematic disease prevention and control methods in the study area.

Key words: Abattoir • Bovine • Fasciolosis • Prevalence • Abaychomen • Guduru

INTRODUCTION

Livestock are an important component of nearly all farming systems in Ethiopia and provide draught power, milk, meat, manure, hides, skins and other products (Funk *et al.*, 2012). Currently, the population of livestock found in Ethiopia is estimated to be 53.4 million cattle, 25.5 million sheep and 22.78 million goats (CSA 2011). Though, Ethiopia has substantial livestock resources, its level of productivity is low due to constraints of disease. Out of

these diseases; Fasciolosis is a major disease which imposes direct and indirect economic impact on livestock production, particularly of sheep and cattle (keyyu *et al.*, 2005; Menkir *et al.*, 2007)[1-20].

The two most important species of this genus are *Fasciola hepatica* and *Fasciola gigantica* and are commonly known as liver fluke (Urquhart *et al.*, 1996). Fasciolosis caused by *Fasciola hepatica* and *Fasciola gigantica* is one of the most prevalent helminthes infections of ruminants in different parts of the world. It

causes significant morbidity and mortality. Both the high land (*Fasciola hepatica*) and the low land (*Fasciola gigantica*) types of live flukes cause severe losses in many parts of Ethiopia where suitable ecological conditions for the growth and multiplication of intermediate host (snails) are found. Areas (site) with seasonally flooded pastures grazing lakeshore, slowly flowing water ways and banks of rivers are among the conducive environment for breeding of snails vectors of fasciolosis (Abebe *et al.*, 2007). The epidemiology of fasciolosis is dependent on the ecology of snail's intermediate host. There are many different species of *Lymnaea* but now generally agreed that two snail types are involved in the transmission of flukes, although there are species variant in different countries (Miodra and Lan McIntyre, 1991). In case of *Fasciola hepatica* the most know intermediate host is *Lymnaea truncatula* of mud snail which prefers moist temperature conditions (15°C-22°C) though some variants found in tropics have adapted to higher temperature and can be breed and survive at 26°C with sufficient moisture during the season, *Lymnaea truncatula* is capable of aestivation for at least a year in dry mud (Miodrag and Lan McIntyre, 1996). The intermediate host for *F. gigantica* is *L. natalensis*. These are aquatic snail which prefers tropical or subtropical condition and thrive in well oxygenated non polluted water. There are however adaptable to an amphibious environment and can aestivate during day weather (Miodrag and Lan McIntyre, 1996)[21-40].

Bovine fasciolosis is an economically important parasite disease of cattle caused by Fasciolidae family, trematodes of the genus *Fasciola*. The parasite imposes direct and indirect economic impact on livestock production and productivity; particularly of sheep and cattle (Keyyu *et al.*, 2005; Menkir *et al.*, 2007). Infected cattle can exhibit poor weight gain and dairy cattle have lower milk yield and possibly metabolic diseases (Mason, 2004). For example, Kithuka *et al.* (2002) reported up to 0.26 million USD annual losses attributable to fasciolosis-associated liver condemnations in cattle slaughtered in Kenya. Another study conducted by Keyyu *et al.* (2006) reported up to 100% liver condemnation rates in some slaughter slabs in Iringa region in Tanzania due to liver flukes in cattle. Although a number of studies have been undertaken with regard to abattoir based prevalence and evaluation of the economic loss due to fasciolosis in different parts of Ethiopia (Tadele and Worku, 2007; Jibat *et al.*, 2008; Fufa *et al.*, 2009; Gebretsadik *et al.*, 2009).

Apart from its veterinary and economic importance throughout the world, fasciolosis has recently been shown to be a re-emerging and widespread zoonosis affecting many people (Esteban *et al.*, 2003). In spite of its higher incidence and economic importance of the parasite in different areas of the country, no research has been conducted in study area. Therefore, the objectives of this study were:

- To determine the prevalence of bovine fasciolosis and its associated risk factors in cattle slaughter in Guduru and Abay choman municipal Abattoir,
- To identify the fluke's species in local area.

MATERIALS AND METHODS

Study Area: The study was conducted in Guduru and Abay choman woreda municipal abattoirs. Guduru is located in Oromia regional state, Horo Guduru Wollega zone west of Addis Ababa, 288km far. The elevation of the area ranges from 1316-2430 meters above sea level. The area receives a mean annual rainfall of about 1100-1600mm. The annual mean temperature is 25°C. The natural grasslands in the areas are traditionally used as grazing fields for livestock. The main grazing land in the area includes swampy area, forest margin, water lodged lands, mountain sides. The climatic condition of the area comprises 21% Kola and 79% Weynadegas. Abay choman werada is located in Oromia regional state, Horo Guduru Wollega zone west of Addis Ababa at a distance of 295km. The elevation ranges from 1316-2430 mean sea level. This area receives a mean annual rainfall of about 1300mm. The annual mean temperature is 26°C. The weather condition shares 60% Bereha and 40% Weynadega. The farming system of both sites are mixed type where crop production and livestock rearing area done side by side. In study area majority of the animal kept by rural farmers are cattle and sheep and also some goats and equines. District covers areas of 820km with livestock population of Bovine 224, 192, Ovine 11,213, Caprine 12,666, Donkey 7,424, Horse 3,221, Mule 720 & Poultry 184,191.

Study Animals: The study animals were cattle of all age and sexes slaughtered in Guduru and Abay choman woreda municipal abattoir. It was difficult to precisely indicate the geographical origin of all animals slaughtered.

Sampling Method and Sample Size Determination: A systematic random sampling technique was used as sampling strategy to collect all the necessary data from abattoir survey of the study animals. The sample size required for this study was determined based on the expected prevalence (50%) of bovine fasciolosis and the 5% desired absolute precision and 95% CI according to Thrusfield (2005).

$$n = \frac{1.96^2 \times P_{exp} (1 - P_{exp})}{d^2}$$

where:

n= is the required sample size

P_{exp}= is expected prevalence

d= is the level of precision (5%).

Accordingly, 384 animals were supposed to be sampled from both abattoirs.

Study Design and Sampling Methodology: A cross-sectional study was conducted from April 2014 to December 2014 with special attention to the livers of the slaughtered animals.

Post Mortem Inspection and Fasciola Species Identification: Post mortem examination of liver and associated bile duct was carefully performed by visualization and palpation of the entire organ followed by transverse incision of the organ across thin left lobe in order to confirm the case (Urquhart, 1996). Species identification of the recovered *Fasciola* was also conducted based on morphological features of the agents and classified in to *Fasciola hepatica*, *Fasciola gigantica* and unidentified or immature forms of liver fluke (Urquhart, 1996).

Data Analysis: All data were coded and entered to Microsoft excel and a preliminary analysis was done on it. STATA-12 (2005) was used to analyze the data. Univariate logistic regression analysis was conducted to see the association between the risk factors and the

occurrence of the infection. Confidence interval and p-value was used to notice the presence of association and Odds Ratios was employed to assess the strength and direction of this association. The outcome variables considered were the adult *Fasciola* species detected during routine postmortem inspection. Descriptive statistics were carried out to summarize the prevalence of each parasite.

RESULTS

Out of the 384 indigenous cattle slaughtered at both Guduru and Abaychomen municipal abattoirs and examined for fasciolosis, 125 animals (liver) were found to be positive for one or both of the species of *Fasciola*. Hence, the overall abattoir based prevalence was 32.6% (95% confidence interval of (0.279-0.375)). Of the 125 livers found to contain fluke infection during post mortem inspection, 49 (39.2%) harbored *F. hepatica*, 59 (47.2%) *F. gigantica*, 17 (13.6 %) had mixed infections (Table 1).

In this study, different variables such as body condition, origin, sex and age were considered. Accordingly, the prevalence of the parasites in different body conditions was found to be: 60% (135) in poor, 23.5% (170) in medium and 5.1% (79) in good body conditioned animals and the infection rate in different origins of animals were Abaychomen 28.5% (179) and Guduru 36% (205). Based on the sexes of animal slaughtered, the prevalence rate of bovine fasciolosis in male was 34.3% (216) and in female 30.4% (168). All the risk factors (origin of animals and sex), there were not statistically significant difference among them. The prevalence fasciolosis in different age groups of animals slaughter in both abattoirs was compared and found to be 15.3% (131) in young and 41.5% (253) in adult animals and it was statistically significant. This was performed using univariate logistic regression analysis and was found to be statistically significant (P=0.000). In addition to that, as animals age increased, the probability of infected by fasciolosis was 3.9 times higher when compared to young animals (OR=3.9; CI= 2.29; 6.74) (Table 2)[41-60].

Table 1: Prevalence of *Fasciola* species in cattle slaughtered at Guduru and Abaychomen municipal abattoirs (n=384)

<i>Fasciola</i> species	No of infected liver	Prevalence (%)
<i>Fasciola hepatica</i>	49	39.2
<i>Fasciola gigantica</i>	59	47.2
Mixed	17	13.6
Total	125	100

Table 2: Association between prevalence and risk factors based on post mortem findings at Guduru and Abaychomen municipal abattoirs

Variables	Positive	Negative	Total	Prevalence (%)	95% CI	P-value
Body condition						
Poor	81	5	135	60	0.51-0.68	0.000
Medium	40	130	170	23.5	0.17-0.30	
Good	4	75	79	5.1	0.01-0.12	
Origin						
Abaychomen	51	128	179	28.5	0.22-0.35	0.123
Guduru	74	131	205	36	0.29-0.43	
Sex						
Male	74	142	216	34.3	0.27-0.41	0.807
Female	51	117	168	30.4	0.23-0.37	
Age						
Young	20	111	131	15.3	0.09-0.22	0.000
Adults	105	148	253	41.5	0.35-0.47	

DISCUSSION

Bovine fasciolosis exists in almost all region of Ethiopia. However, the prevalence, epidemiology and *Fasciola* species prevailing in a given locality varied. This is mainly attributed to the variation in the climate and ecological condition such as altitude, rainfall and temperature and livestock management system (Graber, 1978). In the present study, the overall prevalence was found to be 32.6%. This result was in line with the findings of Mihreteab *et al.* (2010) which was reported to be 32.3%, however, it was significantly higher than the prevalence of bovine fasciolosis reported by Fufa *et al.* (2009) at Welaita Sodo (12.7%), at Soddo abattoir (4.9%), by Gebretsadik *et al.* (2009) at Mekelle (24.32%) and by Swai and Ulicky (2009) at Hawi, Tanzania (14.05%). This might be attributed to the variation in agro-ecological conditions which favors for the survival of both the parasite and the intermediate host. On the other hand, the prevalence of bovine fasciolosis in the present study was lower when compared with the previous reports in different parts of Ethiopia (Tadele and Worku, 2007) at Jimma (46.58%), in South Gondar (83.08%) and (Dagne,1994) in Debre Berhan (80%). The variation in climato-ecological conditions such as altitude, rainfall, temperature, livestock management system and suitability of the environment for survival and distribution of the parasite as well as the intermediate host might have played their own role in such differences. One of the most important factors that influence the occurrence of fasciolosis in a certain area is availability of suitable snail habitat (Urquhart *et al.*, 1996). In addition, optimal base temperature to the levels of 10 °C and 16°C are necessary for snail vectors of *Fasciola hepatica* and *Fasciola*

gigantica, respectively. These thermal requirements are also needed for the development of *Fasciola* with in the intermediate host. The ideal moisture conditions for snail breeding and development of larval stages within the snails are provided when rainfall exceeds transpiration and field saturation is attained. Such conditions are also essential for the development of fluke eggs, miracidiae searching for snails and dispersal of cercariae (Urquhart *et al.*, 1996).

In the present study, species identification revealed that *Fasciola gigantica* was more prevalent (47.2%) than *F. hepatica* (39.2%) and mixed infections (13.6 %). The higher prevalence of *Fasciola gigantica* might be associated with the existence of favorable ecological biotopes for the intermediate host *L. natalensis* and most of the animals may come from low land areas which are the preferable agro-ecology for the intermediate host, *L. natalensis* (Abebe *et al.*, 2007). Unlike the present study, Gebretsadik *et al.* (2009) reported that 56.42% of cattle were infected with *Fasciola hepatica* and 9.17% with *Fasciola gigantica*. However, in another study, Fufa *et al.* (2009) stated that the most common liver fluke species affecting cattle at Welaita Sodo were *Fasciola gigantica*. Yilma and Malone (1998) indicated that *Fasciola gigantica* in Ethiopia is found at altitudes below 1800 meters above sea level. While *Fasciola hepatica* is found at altitude of 1200-2560 meters above sea level. Mixed infections by both species can be encountered at 1200-1800 meters above sea level. According to Yilma and Malone (1998), such discrepancy is attributed mainly to the variation in climatic and ecological conditions such as altitude, rainfall and temperature as well as livestock management system.

There was a statistically significant association between body conditions of the animals and the prevalence of *Fasciola* infection ($P < 0.05$). In support of this finding, a study conducted in Mekelle (Yohannes, 2008) indicated that the association between the prevalence of fasciolosis and body condition of the animals was also statistically significant. Obviously, this could be due to the fact that animals with poor body condition are usually less resistant and are consequently susceptible to infectious diseases. Similar results were also reported by Abebe *et al.* (2011).

In the other way round, the presence of high prevalence of fasciolosis in animals with poor body condition may be due to the effect of the parasite in the animal as *Fasciola* species are blood and tissue fluid suckers and even damage the parenchyma of the liver (immature *Fasciola*) and causes bleeding while the adult parasites are in the bile ducts, which ultimately deplete protein from the host (Urquhart *et al.*, 1996; Marquardt *et al.*, 2000) which leads to poor body condition. Furthermore, cholangitis and liver cirrhosis might be caused by chronic fasciolosis and could reduce bile flow to the duodenum and hence reduced lipid emulsification, digestion and absorption of fatty acid and lipid soluble vitamins. It may also associate with the rationale that animals with poor body condition may have poor immunity and hence may be susceptible to any infectious diseases.

The infection rate fasciolosis in Abaychomen was 28.5% (179) and Guduru 36% (205). The presence of relatively high prevalence in Guduru 36% but absence of statistical variation in the prevalence of fasciolosis between the two origins of animals may be due to the similar in agroecology of the two study areas.

Regarding sexes based prevalence of bovine fasciolosis in slaughtered animal, was found to be relatively higher in males (34.3%) than in females (30.4%). This could be associated to the fact that male animals travel long distance for draught power and harvesting crops, this may lead to lowering immunity.

In the present study, out of all the variables considered (Body condition, origin of animals, sex and age); only body condition and age of animals were found to be statistically significant. Hence, as age increased, the prevalence of the disease was found to be increased, i.e., prevalence was found to be 41.5% in adult and 15.3% in young animals slaughter in both abattoirs. In addition to that, as animals age increased, the probability of infected by fasciolosis was 3.9 times higher when compared to

young animals (OR=3.9; CI= 2.29; 6.74). This may associated with the fact that as age increased, the probability of exposure old animals to parasite (s) will be increased [61-75].

CONCLUSION

Fasciolosis is a major disease which imposes direct and indirect economic impact on livestock production, particularly of sheep and cattle in Ethiopia. Some of the economic losses in the cattle industry induced by fasciolosis are: mortality, liver condemnation, reduced production (meat, milk) and expenditures of anthelmintic for treatment, prevention and control. The present study was conducted at Abaychomen and Guduru municipal abattoir to determine the prevalence of fasciolosis and its associated risk factors. Out of the 384 indigenous cattle slaughtered and examined at both Guduru and Abaychomen municipal abattoirs, 125 (32.6%) of animals and their liver were found to be positive for one or both of the species of *Fasciola*. Of the 125 livers which were found to harbor liver flukes, 47.2% (59) were *F. gigantica* and 39.2% (49) were *F. hepatica* and the rest were mixed infections. In this study, different variables were considered, however, only body conditions and age were found to be statistically significant. According the present study, old animals were 3.9 times more prone to fasciolosis than young and animals slaughtered in Guduru municipal abattoir had a relatively higher prevalence (36%) than in Abaychomen (28.5%). Furthermore, male animals were susceptible for fasciolosis than females. Based on the above conclusions; the following recommendations are forwarded:

- ▶ The finding of present study strongly suggests for the need for appropriate and feasible control measures in study area,
- ▶ Community based control programs such as drainage of swampy area and fencing of watering points should be adopted in the area,
- ▶ If possible regular de-worming of animals before and after rainy season is warranted,
- ▶ Supplementation of important nutrient feed in dry season is important to avoid stress conditions that affect the host resistance and susceptibility to parasitic diseases,
- ▶ Further detailed epidemiological study as well as assessment of the overall economic loss incurred due to fasciolosis should be performed in the study area in order to expand and implement disease investigation and control strategy.

REFERENCES

1. Abebe, G., G. Taddese, Y. Jobre, D. Pedon and M. Asrat, 2007. Evaluation of the bspatial distribution of *Fasciola hepatica* and *Fasciola gigantica* using geographica information system (GIS) in the Nile River Basin, Ethiopia. *Ethipian Veterinary Journal*, 11(2): 41-57.
2. Al Habbib, W.M.S. and S.S. Al Zako, 1981. The effect of different temperatures on development of intra-molluscan stages of *F.gigantica*, *J. Ther. Biology*, 6: 373-377.
3. Alemayew, R., W. Tefera, D. Seble, M. Nebyou, A. Dinka and A. Fufa, 2012. Bovine Fasciolosis: Coprological, Abattoir survey and Financial loss due to liver condemnation in Bishoftu municipal Abattoir, Central Ethiopia, *J. Vet. Med. Anim. Health*, pp: 24-26.
4. Alula, P., K. Addisu and W. Amanuel, 2013. Prevalence and Economic significance of Bovine fasciolosis in Nekemte municipal Abattoir, *J. Vet. Med. Anim. Health*, 5(8): 202-205.
5. Andrews, SI., 1999. The life cycle of *Fasciola hepatica*. In: *Fasciolosis* (edited by Dalton, J P.). Dublin City University. CAB International Publishing, UK, pp: 1-3.
6. Bahru, G. and M. Ephrem, 1979. Preliminary survey of bovine fasciolosis. *Res. Agri. Science*, 1: 50-127.
7. Beyazn, C., 1995. Preliminary study on bovine fasciolosis in Eastern Gojjam Region. DVM thesis. Faculty of veterinary medicine, Addis Ababa University. Dabre Zeit, Ethiopia.
8. Bowman, D., 1999. *Parasitological for veterinarians* seventh edition. Ithaca, New York, pp: 109-116.
9. Bowman, D.D., E.R. Lynn, L.M. Eberhard and A. Alcaraz, 2003. *Georgis' Parasitology for Veterinarians*. Eighth edition. USA, W. B. Saunders Company, pp: 144-220.
10. Central statistical Authority (CSA) 2011. Agriculture sample survey 2010-2011. Report on Livestock and livestock characteristics, Statistical Bulleth No. 505. Addis Ababa, Ethiopia, pp: 11.
11. Dagne, M., 1994. Survey on prevalence and economic significance of bovine fasciolosis at Debre Berhan region. DVM thesis, Faculty of veterinary medicine Addis Ababa University. Dabre Zeit, Ethiopia.
12. Dechasa, T., W. Antenah and F. Dechasa, 2012. Prevalence, gross pathological lesions and economic losses of bovine fasciolosis at Jimma municipal abattoir, Ethiopia, *J. Vet. Med. Anim. Health.*, 4(1): 6-11.
13. Devendra, C. and B. Marca, 1983. *Goat production in tropics: Common Wealth Agriculture Bureaux*. Published by Unwin Limited, old working, Surrey, pp: 90-92.
14. Elmer, R. Noble and Glenna A. Noble, 1982. *Parasitology. The Biology of Animal Parasites*. Fifth edition, *Lea and Febigen*, Philadelphia, pp: 172.
15. Elmer, R. Noble and Glenna, A. Noble, 1982. *Parasitology. The Biology of Animal Parasites*. Fifth edition. Lea and Febigen, Philadelphia, pp: 172.
16. Ephrem. B., M. Wassie and G. Abadi, 2011. Prevalence and economic losses of Bovine Fasciolosis in Dessie municipal abattoir, south wollo zone, Ethiopia. *Erop. J. Bio. Sci.*, 4(2): 53-59.
17. Equar, Y., A. Gashaw, M. Girmay and G. Pa, 2012. Prevalence of bovine fasciolosis, amplitude of liver Condemnation and its economic impact in Mekelle municipal abattoir, *Int. J. livestock Res*, 2(2): 196-205.
18. Esteban, J., C. Gonzalez, F. Curtale, C. Mun oz-Antoli, M. Valero, M. Bargaes, M. El sayed, A. El Wakeel, Y. Abdel-Wahab, A. Montresor, D. Engels, L. savioli and S. Mas-coma, 2003. Hyperendemic fasciolosis associated with schistosmais in Villages in the Nile delt of Egypt, *Am Jour of Trop Med Hyg.*, 69: 429-437.
19. Fairweather, I., T. Threadgold and B. Hanna, 1999. Development of *Fasciola hepatica* in the mammalian host. In: *Fasciolosis* (edited by Dalton, J P.). Dublin City University. CAB International Publishing, UK, pp: 98-100.
20. Fekadu, R., 1988. Ruminant fasciolosis: Studies on the clinical occurrence, coprology, morphology and abattoir survey in Debre Brehan and surrounding area, M.S.C. Thesis F.V.M, A.A.U.
21. Fufa, A., A. Loma, M. Bekele and R. Alemayew, 2009. Bovine Fasciolosis: Coprological, Abattoir survey and its economic impact due to liver condemnation at Soddo municipal Abattoir southern Ethiopia. *Trop. Animal Health prod*.
22. Funk, C., J. Rowland, G. Eilerts, E. Kebebe, N. Biru, L. White and G. Galu, 2012. A climate trend analysis of Ethiopia. *Climate change adaptation series*. U.S.Geological survey. Ramine early warning system Net-work informing (FEWSNET). Fact sheet, pp: 3053.

23. Gebrecherkos and Berihun Afera, 2012. Prevalence of bovine fasciolosis and its economic significance of at Adigrat, Tigray, Ethiopia. *Trop. Ani. Health and prod.*, 26(8).
24. Gebrestadik, B., B. Kassahun and T. Gebrehiwot, 2010. Prevalence and economic significance of fasciolosis in cattle in Mekele area of Ethiopia, *Trop. Ani. Health and prod.*, 41(7).
25. Gebru, L., 2008. Epidemiology and economic importance of fasciolosis of domestic ruminants in selected sites of Tigray regional state, northern Ethiopia. MSc Thesis, Faculty of Veterinary Medicine, Addis Ababa University, Debre Zeit, Ethiopia, pp: 13.
26. Georgi, J., 1985. *Parasitology for Veterinarians*. Fourth edition. UK Saunders Company, pp: 70-85.
27. Grabber, M. and P. Daynes, 1974. Molluscs vectors and trematodes of human's animal in Ethiopia, *Revue d'élevage Médecine Vétérinaires Days Tropicaux*, 27: 307-322.
28. Graber, M., 1975. Helminths and helminthosis of domestic and wild animals in Ethiopia. *Rev. Med. Vet. Parasitol*, 1: 13-95.
29. Hanson, J. and B. Perry, 1994. *The Epidemiology, Diagnosis and Control of Helminth Parasites of Ruminants*. A Hand Book. Food and Agricultural Organization of the United Nations, Rome, Italy, pp: 72-89.
30. Hendrix, C.M., 1998. *Diagnostic Veterinary Parasitology*. Second edition. USA: MOSBY, Inc., pp: 108-136.
31. Herenda, D., P. Chambers, A. Ettriqui, P. Seneviratna and J. Dalsilvat, 2000. *Manual on meat inspection for developing countries*, food and Agricultural organization of the united Nation (FAO), Rome, Italy.
32. Kaufman, J., 1996. *parasitic infection of Domestic Animals*, A diagnostic manual, Berlin Germany, CIBA- Geigy, pp: 6-8.
33. Keyyu, J., J. Monrad, N. KyVsgaard and A. Kassukuu, 2005. Epidemiology of *fasciola gigantica* and Amphistomes in cattle on traditional, small scale dairy and large scale dairy farms in the southern highlands of Tanzania. *Trop. Ani. Health Prod.*, 37: 303-314.
34. Khallaayoune, K. and M. El-Hari, 1991. Variations Saisonnières de infection par *Fasciola hepatica* Chez le chevere dans La region du Hauz (Maoc). *Research Veterinarian*, 22: 219-225.
35. Lora, R., 2001. *Veterinary Parasitology*, by Butterworth-Heinemann., A member of the Read Elsevier Group, pp: 268-277.
36. Machanicka, B., 2000. Fasciolosis. *Acta, Parasitologia*, 45: 123-124.
37. Malone, J. and J. Yilma, 1998. Predicting outbreaks of fasciolosis: from Ollerenshow to Satellites. In: *Fasciolosis* (edited by Dalton, J.P.). Dublin City University. CAB International Publishing, pp: 1-3.
38. Mas-Coma, S., M.D. Bargues and J.G. Estenban, 1999. Human fasciolosis. In: *Fasciolosis* (edited by Dalton, J. P.). Dublin City University. CAB International Publishing, UK, 411-428.
39. Mas-coma, S., M. Bargues and M. Valero, 2005. Fasciolosis and othe plant-born trematode zoonosis, *Int. J. Parasitol*, 35: 1255-1278.
40. Mason, C., 2004. Fasciolosis associated with metabolic disease in a dairy herd and its effects on health and Productivity, *Cattle Practice*, 12: 7-13.
41. Menkier, M., A. Ugglu and P. waller, 2007. Prevalence and seasonal incidence of nematode parasites and fluke infections of sheep and goats in eastern Ethiopia. *Trop. Ani. Health Prod.*, 39(7): 521-531.
42. Merck, N., 1998. *Veterinary Manual*, 8th ed. Volume I, USA with House Station, pp: 197-9.
43. Michael, A., 2004. infection prevalence of ovine fasciolosis in irrigation shems along the upper Awash River Basic and effects of strategic Anthelminitic Treatment in selected up stream Areas, MSc thesis Addis Ababa University, pp: 1-20.
44. Mihreteab, B., T. Haftom and G. Yehenew, 2010. Bovine fasciolosis: Prevalence and its economic loss due to liver condemnation at Adwa municipal Abattoir, North Ethiopia. *EJAST*, 1(1): 39-47.
45. Miodra, R. and D. Lan McIntyre, 1996. *Disease of cattle in the Tropics*. London, pp: 531-534.
46. Mira, S. and S. Ralph, 1989. *Manual for Tropical Veterinary Parasitology*. UK, CAB International Publishing, UK, pp: 71.
47. Mulugeta, H.S., J. Getachew, M. Tafesse, W. Getachew, G. Kinfe and Y. Teshome, 1989. The significance of helminthe parasites in livestock production. Paper Presented at the Third Livestock Improvement Conference., Addis Ababa, Ethiopia, pp: 82.
48. Nicholson, M.J. and M.H. Butterworth, 1986. A Guide to condition scoring of zebu cattle. *ILCA. Lpp-DFID*, pp: 1-29.
49. Njau, B.C. and R.G. Scholltens, 1991. The roll of traditionally harvested hay in the transmission of ovine fasciolosis in Ethiopian highlands., *Vet. Res. Com.*, 15: 369-372.

50. Njau, B., O. Kasali, R. scholtens and N. Akale work, 1989. The influence of watering practice on the transmission of *Fasciola* among sheep in Ethiopian highlands., *Vet. Res. Commu.*, 3(1): 67-74.
51. Nuradis, I., W. Pawlos and T. Tadele, 2010. Prevalence of Bovine fasciolosis and economic importance due to liver condemnation at kombolcha Industrial Abattoir, Ethiopia, *The internet J. Vet. Med.*, 8(2).
52. Ogurninade, A. and B. oguminade, 1980. Economic importance of fasciolosis in Nigeria. *Trop. ani. Health Prod*, 12: 155-160.
53. Payne, W.J.A., 1990. *An Introduction to Animal Husbandry in the Tropics*. Fourth edition. ELBS, UK, Longman, British, pp: 238-258.
54. Phiri, A., I. Phiri, C. Sikasunge and J. Monrad, 2005. Prevalence of fasciolosis in Zambian cattle observed at selected abattoir with emphasis on age, sex and origin, *J. Vet. Med. B*, 52: 414-416.
55. Radostits, D.M., D.C. Blood and C.C. Gray, 1994. *Veterinary Medicine text book of the diseases of cattle, sheep, Goat, Pig and horse*. 8th Ed. ELBS and Bailleire Tindall.
56. Radostits, O.M., C.C. Gay, D.C. Blood and K.W. Hinchcliff, 2000. *Veterinary medicine. A text book of diseases of cattle, sheep, pigs, goats and horses*. 9th edn. USA, W.B. Saunders Company Ltd, pp: 1339-1352.
57. Radostitis, O., D. Blood and J. Henderson, 2007. *A text book of the disease of cattle, sheep, pips, goats and horses*. 10th ed of veterinary medicine ELBS Vallier India., pp: 250-340.
58. Rahmeto, A., A. Fufa, B.Mulugeta, M. Solomon, M. Bekele and R. Alemayehu, 2010. Fasciolosis: prevalence, financial losses due to liver condemnation and evaluation of a simple sedimentation diagnostic technique in cattle slaughtered at Hawasa municipal abattoir, southern Ethiopia. *Ethiopia Vet. J.*, 14(1): 39-51.
59. Ramajo, V., A. Oleaga, P. Casnueva, G. Hillyer and A. Muro, 2001. Vaccination of sheep against *Fasciola hepatica* with homologous fatty acid binding proteins. *Vet. Parasitol*, 97: 35-46.
60. Roberts, J.A., E. Estuningsih and T.W. Spithill, 1997. Resistance of Indonesian Thin Tail sheep against *Fasciola hepatica* and *F. gigantica*, *J. Vet. Parasitol*, 68: 69-7.
61. Robinson, M. and J. Dalton, 2009. Zoonotic helminth infections with particular emphasis of fasciolosis and other trematodies. *philosophal transactions of the Royal society of London, Series B, Biological Science*, 364: 2763-2776.
62. Rokni, M., 2002. Diagnosis of human fasciolosis in the Gilan province of northern Iran: application of cathepsin L-ELISA. *Diagnostic Microbiology and Infectious Disease*, 44: 175-179.
63. Schoenian, S., 2003. *Parasite of Sheep and Goats*. Western Maryland Research and Education Center. Area Agent and Extension, pp: 1-3.
64. Shiferaw, M., T. Ephrem and B. Feyisa, 2011. Prevalence of bovine *fasciolosis* and its economic significance in and around Assela, *The Int. J. Vet.* 11(3)
65. Soulsby, E., 1982. *Helminthes, Arthropods and protozoa of Domesticated Animals*. seventh Edition. Braviere Tindall, London, UK., pp: 40-52.
66. Soulsby, E., 1986. *Helminth Arthropad and protozoa of Domestic animals*. 7th ed. Baillere. Tindal, London, UK., 334: 40-52.
67. Spithill, T. and J. Dalton, 1998. progress in development of liver fluke vaccines. *Parasitology Today*, 14(6): 224-228.
68. Spithill, T.W., P.M. Smoker and D.B. Copeman, 1999. *Fasciola gigantica: epidemiology, control, Immunology and molecular biology*, In: Dalton, J.P. (ED). *Fasciolosis*. CAB International Publishing Walling Ford, pp: 465-525.
69. Swell, M.M., 1966. The pathogenesis of fasciolosis. *J. Vet. Res.* In: *Seasonal dynamics of F. gigantica* burden in grazing Timahdit Sheep Morroco. (Edited by Dakkak, 1991). In. *J. Parasitol*, 21: 307-314.
70. Tadele, T. and T. worku, 2007. The prevalence and economic significance of Bovine fasciolosis at Jimma Abattoir, Ethiopia, *Int. J. Vet. Med.*, 2: 1-7.
71. Theodoridis, Y., J.L. Duncan and J.M. MacLean, 1991. Pathophysiological studies on *Dicrocoelium dendriticum* infection in sheep, *J. Vet. Parasitol*, 39: 61-66.
72. Thrusfield, M., 2005. *Vet. Epidemiology*. Second edition, University of Edinburgh, Black well science, pp: 180-188.
73. Torgerson, P. and J. Claxton, 1999. Epidemiology of fasciolosis. In: *Fasciolosis* (edited by Dalton, J. P.). Dublin City University. CAB International Publishing, UK, pp: 113-145.
74. Urquhart, G., J. Armour, J. Duncan, A. Dunn and F. Jennings, 1996. *Veterinary parasitology*. 2ed. Black well science UK, pp: 103-113.
75. Zheng, H.J., C.W. Zheng-Hou Tao and W.F. Pessens, 1990. Comparison of dot ELISA with Sandwich ELISA for the detection of circulating antigens inpatient with Bancroftiasis., *J. Trop. Med. Hyg*, 42: 546-549.