Middle-East Journal of Scientific Research 13 (8): 998-1009, 2013 ISSN 1990-9233 © IDOSI Publications, 2013 DOI: 10.5829/idosi.mejsr.2013.13.8.3531

## Physiological and Agro-Morphological Response to Drought Stress

Mostafa Ahmadizadeh

Young Researchers Club, Jiroft Branch, Islamic Azad University, Jiroft, Iran

**Abstract:** Although, drought stress has been well documented as an effective parameter in decreasing crop production; developing and releasing new varieties which are adaptable to water deficit conditions can be a constructive program to overcome unsuitable environmental conditions. A good understanding of factors limiting yield now provides us with an opportunity to identify and then select for physiological traits, which increase drought tolerance and yield under rainfed conditions. Applying different physiological and agro-morphological tests to appreciate drought tolerance in plant leads to faster selection methods. Therefore, these characters can be used as an indirect selection criterion for screening drought tolerance plant materials, this strategy will lead to new cultivars with high yield potential and high yield stability that in turn will result in superior performance in dry environments.

Key words: Morphological traits • Chlorophyll fluorescence • Oxidative stress • Germination • Electrolyte leakage

## INTRODUCTION

Drought is raising threat of world. Most of the countries of the world are facing the problem of drought. The insufficiency of water is the principle environmental stress and to enter heavy damage in many part of the world for agricultural products [1-5]. Among the environmental stresses, drought stress is one of the most adverse factors for plant growth and productivity [6, 7]. Drought stress can reduce grain yield, have estimated the average yield loss of 17 to 70% in grain yield due to drought stress [8]. Drought is a complex physicalchemical process, in which many biological macro molecules and small molecules are involved, such as nucleic acids, proteins, carbohydrates, lipids, hormones, ions, free radicals, mineral elements [9-14]. The ability of a cultivar to produce high yield over a wide range of environmental condition is very important. Response of plants to water stress depends on several factors, such as developmental stage, intensity and duration of stress and cultivar genetics [15, 4]. The plant response is complex because it reflects over space and time the integration of stress effects and responses at all underlying levels of organization [16]. Improving drought resistance is, therefore, a major objective in plant breeding programs for rainfed agriculture in these regions [17-19]. Knowledge of genetic behavior and type of gene action controlling target traits is a basic principle for designing an appropriate breeding procedure for the purpose of genetic improvement. Hence, the success of any selection or hybridization breeding program for developing droughttolerant varieties depends on precise estimates of genetic variation components for traits [20, 21]. It inhibits the photosynthesis of plants, causes changes in chlorophyll contents and components and damage to the photosynthetic apparatus [22]. In addition, it inhibits the photochemical activities and decreases the activities of enzymes in the Calvin Cycle in photosynthesis [17]. Conventional plant breeding attempts have changed over to use physiological selection criteria since they are time consuming and rely on present genetic variability. Tolerance to abiotic stresses is very complex, due to the intricate of interactions between stress factors and various molecular, biochemical and physiological phenomena affecting plant growth and development [16, 23].

Morphological and agronomic traits have a special role to determine the importance of each trait on increasing yield, as well as to use those traits at the breeding programs, which at least lead to improving yield and introducing commercial varieties under end seasonal drought stress condition [24]. Morphological characters

Corresponding Author: Mostafa Ahmadizadeh, Young Researchers Club, Jiroft Branch, Islamic Azad University, Jiroft, Iran. Cell: +989194457655. include root length, spike number, grain number per spike, 1000 grain weight, awn length [25, 15, 4]. Wajid et al. [26] reported that wheat crop produces highest grain yield by applying irrigation at all definable growth stages. Because irrigation is an expensive input, farmer, agronomist, economist and engineer need to know the response of yield to irrigation. Furthermore, Jahfari [27] and Rafique [28] reported that yield and yield components are significantly increased within different wheat cultivars. Garavandi and Kahrizi [29] by evaluating 20 bread wheat genotypes reports that genotypes has higher genetic diversities for grain yield, spike number per square meter, number of seed per spike, spike density and awn length in comparison with other traits. Development of cultivars with high yield is the main goal in water limited environments, but success has been modest due to the varying nature of drought and the complexity of genetic control of plant responses [30]. In plants, a better understanding of the morpho-anatomical and physiological basis of changes in water stress resistance could be used to select or create new varieties of crops to obtain a better productivity under water stress conditions [31]. The reactions of plants to water stress differ significantly at various organizational levels depending upon intensity and duration of stress as well as plant species and its stage of growth [32]. Understanding plant responses to drought is of great importance and also a fundamental part for making the crops stress tolerant [33].

The purpose of this research was to study drought stress effect on some physiological and agromorphological, so that responses of these traits to drought stress can be evaluated in resistance to drought stress.

**Oxidative Stress and Antioxidant Defense Systems:** When plants are subjected to various abiotic stresses, some reactive oxygen species (ROS) such as super-oxide (O2<sup>-</sup>), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl radicals (OH) and singlet oxygen are produced. These ROSs may initiate destructive oxidative processes such as lipid peroxidation, chlorophyll bleaching, protein oxidation and damage to nucleic acids [34, 17]. The antioxidant defenses appear to provide crucial protection against oxidative damages in cellular membranes and organelles in plants grown under un-favorable conditions [35]. Active oxygen species were considered to be important damaging factors in plants which exposed to stressful environmental conditions such as drought [36-38]. The antioxidant defense system in the plant cell includes both enzymatic (antioxidants), such as superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX) [37, 39, 38] and non-enzymatic antioxidants including  $\beta$ -carotenes, ascorbic acid (AA) [40],  $\alpha$ -tocopherol ( $\alpha$ -toc) [41], reduced glutathione (GSH) [42]. Carotenes form a key part of the plant antioxidant defense system, but they are very susceptible to oxidative destruction. B-carotene, present in the chloroplasts of all green plants is exclusively bound to the core complexes of PSI and PSII. Protection against damaging effects of ROS at this site is essential for chloroplast functioning. Here  $\beta$ -carotene, in addition to function as an accessory pigment, acts as an effective antioxidant and plays a unique role in protecting photochemical processes and sustaining them [43, 23]. A major protective role of  $\beta$ carotene in photosynthetic tissue may be through direct quenching of triplet chlorophyll, which prevents the generation of singlet oxygen and protects from oxidative damage [44]. To keep the levels of active oxygen species under control, plants have non-enzymatic and enzymatic antioxidant systems to protect cells from oxidative damage [17]. Superoxide dismutases (SODs), a group of metalloenzymes, are considered as the first defense against ROS, being responsible for the dismutation of O<sup>2-</sup> to H<sub>2</sub>O<sub>2</sub> and O<sup>2</sup>. CAT, APX, POD are enzymes that catalyze the conversion of  $H_2O_2$  to water and O2 [17, 38]. Transformation of many plant genera for useful traits, such as oxidant-resistance is now routine [45]. Recently the involvement of H<sub>2</sub>O<sub>2</sub> and SOD in regeneration of plants has also been proposed [46]. Hydrogen peroxide is commonly taken as an indicator of oxidative stress, because it is induced by activated oxygen species (AOS) and also influencing the level of lipid per oxidation [35]. However, H<sub>2</sub>O<sub>2</sub> is also toxic to cells and has to be further detoxified by CAT and/or peroxidase (POD) to water and oxygen [37, 38].

Acclimation of plants to drought is considered to promote antioxidants defense systems to face the increased levels of activated oxygen species (AOS), which in turn, cause membrane damage by lipid peroxidation and indicated by malondialdehyde (MDA) content, which is one of the main parameters for evaluating membrane oxidation extent and are toxic for cells [47, 38]. The decline in CAT activity is regarded as a general response to many stresses [48, 49]. Ahmadizadeh *et al.* [38] with suited on 37 durum wheat landraces from Iran and Azerbaijan republic reported that the activity of SOD and CAT decreased in susceptible landraces, where as in resistant landraces SOD and CAT remained unchanged and in some cases they showed an increase under stress condition. In these genotypes (resistant) increasing SOD and CAT accompanied with ID decrease in the membrane. There are many reports in the literature that underline the intimate relationship between enhanced or constitutive antioxidant enzyme activities and increased resistance to environmental stresses in several plant species, such as rice [48], foxtail millet [50], tomato [51], sugar beet [52], oilseed rape [17], wheat [53, 54, 38] and barley [55].

Drought Stress and PSII Activity: Photosystem II (PSII) is highly sensitive to environmental limiting factors and PSII reaction center and its chemical reaction being adversely affected by drought stress [56, 57]. Photosynthetic carbon reduction and carbon oxidation cycles are the main electron sink for PSII activity during mild drought [58]. It was shown that PSII functioning and its regulation were not quantitatively changed during desiccation. The CO2 molar fraction in the chloroplasts declines as stomata close in drying leaves. Havaux [59] has investigated the impact of various environmental stresses (drought, heat, strong light) applied separately or in combination on the PSII activity. The existence of a marked antagonism between physicochemical stresses (e.g. between water deficit and HT) was established, with a water deficit enhancing the resistance of PSII to constraints as heat, strong light. Similar results were obtained on bean plants [60]. Noctor et al. [61] provided quantitative estimation of the relative contributions of the chloroplast electron transport chain and the glycolate oxidase load placed on the photosynthetic leaf cell.

Chlorophyll Content: Drought stress produced changes in the ratio of chlorophyll 'a' and 'b' and carotenoids [44]. Chlorophyll content is positively associated with photosynthetic rate which increases biomass production and grain vield. Significant relationships between chlorophyll content and yield and yield components facilitate selection of high yielding genotypes [62]. Photosystem II (PSII) is highly sensitive to environmental inhibiting factors and water stress will damage its reaction centers severely. The chemical reaction of PSII is also affected strictly by water stress [63, 57]. Chlorophyll concentration has been known as an index for evaluation of source, therefore decrease of this can be consideration as a non stomata limiting factor in the drought stress conditions. There are reports about decrease of chlorophyll content in drought stress conditions [64]. Also, it is reported that chlorophyll content of resistant

and sensitive cultivars to drought and thermal stress reduced. But resistant cultivar to drought and thermal stress conditions had high chlorophyll content [65]. Some study has demonstrated that chlorophyll content is positively correlated with photosynthetic rate [66]. Increasing the chlorophyll content in crops may be an effective way to increase biomass production and grain yield [67, 68]. It has reported under drought stress rate of chlorophyll a to b has increased on wheat [69]. Decrease of chlorophyll content and water potential of soil has represented in plants such as sunflower [70] and Tobacco [71]. A reduction in chlorophyll content was reported in drought stressed cotton [72] and Catharanthus roseus [32]. The chlorophyll content decreased to a significant level at higher water deficits in sunflower plants [73] and in Vaccinium myrtillus [74]. Other reports have represented that drought stress did not have effect on chlorophyll concentration [75]. Pastori and Trippi [71] expressed that resistant genotypes of wheat and corn had higher chlorophyll content than sensitive genotypes under the oxidative stress. Ashraf et al. [69] also reported that drought stress will reduce concentration of chlorophyll b more than chlorophyll a.

Chlorophyll Fluorescence: The use of chlorophyll fluorescence from intact, attached leaves proved to be a reliable, nonintrusive method for monitoring photosynthetic events and for judging the physiological status of the plant [76]. Fluorescence induction patterns and derived indices have been used as empirical diagnostic tools in stress physiology [77]. Thus, PSII fluorescence can be regarded as a biosensing device for stress detection in plants. One of the most important parameters in rapid fluorescence kinetics is variable Fluorescence (Fv), i.e., the difference between maximal and minimal fluorescence (Fm-F0). The variable to maximum fluorescence ratio (Fv/Fm) is an indicative of potential or maximum quantum yield of PSII [78]. It is an important parameter of the physiological state of the photosynthetic apparatus. The declining slope of Fv/Fm is a good indicator to evaluate photo-inhibition of plants exposed to environmental stresses such as drought and heat, accompanied by high irradiance [57]. A promising approach is the use of chlorophyll fluorescence, a technique that can provide large amounts of data with a minimum of expertise and time and without injury to the plants. Chlorophyll fluorescence works on the principle that photosynthesis is one of the core functions in the physiology of plants. The functional state of photosynthesis has been considered an ideal physiological activity to monitor the health and vitality of plants [79]. Chlorophyll fluorescence techniques are often used to detect environmental, chemical and biological stress in plant tissue [80]. According to Paknejad et al. [56] drought stress reduces the variable (Fv) and initial (F0) fluorescence parameters and quantum yield (Fv/Fm). Under dry conditions chlorophyll fluorescence was considered as a useful tool for screening and breeding of wheat cultivars [81]. Vazan [82] reported that drought stress reduces variable fluorescence (FV), initiative fluorescence (F0) and quantum yield (FV, FM).

Relative Water Content (RWC): Drought stress affects water status in plants. Relative water content is useful means for determining the physiological water status of plants [7]. Relative water content is the indicators of degree of drought stress. RWC of leaves is higher in the initial stages of leaf development and declines as the dry matter accumulates and leaf matures. Obviously, stressed plants have lower RWC than non-stressed plants. RWC of non-stressed plants range from 85 to 90%, while in drought stressed plants; it may be as low as30% [83]. In studies that performed on 4 cultivars of bread wheat, RWC reduced to 43 percent (from 88% to 45%) by moisture stress [84]. Mationn et al. [85] represented a similar report as regards a drop in the amount of RWC in tolerant and sensitive cultivars of barley. Significant differences in leaf water potential and RWC were recorded among the tolerant and intolerant cultivars of wheat; results were consistent with Subrahmanyam et al. [86] and Tas and Tas [87]. Therefore osmotic regulation will help to cell development and plant growth in water stress. It is defined that decrease of relative water content close stomata and also after blocking of stomata will reduce photosynthesis rate. It is reported that high relative water content is a resistant mechanism to drought and that high relative water content is the result of more osmotic regulation or less elasticity of tissue cell wall [65]. Overall decrease in RWC under drought stress was highly significant in all the cultivars used, in accordance with Allahmoradi et al. [88] in Mungbean, Mohammadkhani and Heidari [89] in maize, Moaveni [14] and Farshadfar et al. [2] in wheat. While, Liu et al. [90] reported a gradual decrease in RWC after application of PEG treatment as water stress and Gonzalez et al. [91] has also recorded significant decrease in Leaf Ø and RWC in barley under drought stress. Shamsi [65] reported that with an increase in the Intensity of drought stress on wheat cultivars, there was a decrease in relative water content.

Cell Membrane Stability: Cell membranes are one of the first targets of many plant stresses [38] and it is generally accepted that the maintenance of their integrity and stability under water stress conditions is a major component of drought tolerance in plants [92]. These modifications occur mainly in drought sensitive plants and lead to a loss of semi permeable properties of the cell membrane, which is the main reason of metabolic damages developed in water stress plants. Therefore the integrity and stability of cell membrane in water deficit conditions can be considered a possible adaptive value indicative of stress resistance. Cell membrane stability may be determined through estimation of the extent of cell membrane damage in desiccated of leaf fragment in vitro whit a polyethylene glycol solution (PEG) and subsequent measurement of electrolyte leakage into aqueous medium [93]. The degree of cell membrane injury induced by water stress may be easily estimated through measurements of electrolyte leakage from the cells [92, 38]. These tests determine the degree of cell membrane damage caused by stress based on electrolyte leakage from the cells. The technique is relatively simple, repeatable and rapid and requires inexpensive equipment, can be used on plant material from a variety of cultural systems and it is suitable for the analysis of large numbers of samples [77, 94]. However, despite its many advantages, electrolyte leakage was found to be markedly influenced by various experimental parameters, especially washing time of collected samples before PEG exposure [95, 96], intensity and duration of the PEG treatment [97] and duration of the rehydration period [98].

Saneoka *et al.* [99] and Azizi-e-Chakherchaman *et al.* [100] in Lentil studied the relationship between plasma membrane stability (obtained from EC measurement) and grain yield in stress and non stress conditions. They reported that plasma membrane stability in genotypes under stress was significantly lower than genotypes under non stress conditions. The cell membrane stability has been exclusively used as selection criterion for different abiotic stresses including drought and high temperature in wheat [101], rice [102] and sorghum [103]. The test to detect the integrity of cell membrane is called cell membrane stability (CMS) and was used to characterize drought resistance in plants [104, 92, 94, 57, 38].

Germination and Recovery Germination: Seedling emergence is one of the stages of growth that is sensitive to water deficit. Therefore, seeds germination, are prerequisites for the success of stand establishment of crop plants. Under semiarid regions, low moisture is limiting factor during germination. The rate and degree of seedling establishment are extremely important factors in determination of both yield and time of maturity [105, 106]. Crop establishment depends on an interaction between seedbed environment and seed quality [106]. It is critical to understand the seed germination ability of droughttolerant forage species under drought stress and their recovery response when removed from drought condition. This information will help in the successful establishment of pastures in dry land. Stress tolerance of plants varies among species and their ecotypes [107]. Recovery germination of seeds in fresh-water after they were exposed to saline conditions has been investigated [108] to determine if seeds can remain viable after being exposed to hypersaline conditions [109]. One of the prerequisites to successful breeding for drought tolerance is availability of reliable methods for screening of desirable genotypes. Classical breeding may be complemented with laboratory method which is created models for simulation of water deficiency and drought conditions. In this respect, one of the most popular approach is to use high molecular weight osmotic substances, like polyethylene glycol (PEG), added to the medium for seed germination or plant/cell development [105, 106, 110-114]. Coefficient of velocity of germination (CVG) indices evaluates drought stress tolerance. Genotype with height coefficient of velocity of germination (CVG) is in stress condition. There are significant differences between laboratory data and drought stress tolerance, such as growth seedling, root length, root/shoot [115] and coefficient of velocity of germination (CVG) [106].

Germination in solution with high osmotic potential is one of the most important laboratory methods suggested for screening drought tolerance of crop plants. Good laboratory tests for screening genotypes have to show significant correlation with drought resistance [116]. Genotypic ability for high root to shoot length ratio contribute to drought tolerance. The efficiency of soil water uptake is by the root system therefore it is a key factor in determining the rate of transpiration and tolerance to drought. Water uptake by the root is a complex parameter that depends on root structure, root anatomy and the pattern by which different parts of the root contribute to overall water transport [117]. Germination in polyethylene glycol (PEG), measurements of root length or rooting depth and the survival or growth of seedlings which is subjected to osmotic, have been

suggested for drought screening [106]. An increased root growth due to water stress was reported in sunflower [118] and Catharanthus roseus [32]. Takele [119] found to have differential responses of genotypes to variable soil moisture deficits for their specific seedling shoot and root lengths. Grzesiak *et al.* [120] noticed varietal differences in seedling growth and coleoptile length affected by drought simulated by a water solution of mannitol of chemical water potential of -0.3 and -0.6 MPa. The characters germination percentage and root to shoot length ratio showed considerable variability under stress conditions.

Effects of Drought Stress on Morphological Characteristics: It has been established that drought stress is a very important limiting factor at the initial phase of plant growth and establishment. It affects both elongation and expansion growth [23]. Morphological characters such as number of tillers, grain per spike number, fertile tillers number per plant, 1000 grain weight, peduncle length, awn length, plant height, spike length, kernel number per spike, grain weight per spike and etc. affect the wheat tolerance to the moisture shortage in the soil [18, 121, 4, 5, 122, 123]. Study of yield contributing components in respect of their genetic mechanism is very important for improvement in grain yield. Information regarding interrelationships between quantitatively inherited plant traits and their direct and indirect effects on grain yield is of great importance for success in selections to be conducted in breeding programs [124]. The yield components like grain number and grain size were decreased under pre-anthesis drought stress treatment in wheat [125]. In some other studies on maize, drought stress greatly reduced the grain yield, which was dependent on the level of defoliation due to water stress during early reproductive growth [126]. Water stress reduces seed yield in soybean usually as a result of fewer pods and seeds per unit area [127]. Heydari et al. [128] to study genetic diversity of different traits in 157 lines of double haploid bread wheat, indicated that their understudy lines have higher genetic diversity for last internode length, number of fertile spike per area unit, plant height, number of grain and grain yield per spike in comparison with other traits like grain volume weight, days to maturity, days to heading and days to anthesis. Bahari and Sabzi [129] Studying morphological traits with grain yield of durum and aestivum genotypes showed harvest index, no. grain in m2 and No. of grain spike traits most have a role in increasing yield.

Ahmadizadeh et al. [122] with suited on 37 durum wheat landraces from Iran and Azerbaijan Republic reported that under drought stress conditions there were positive significant correlations between the yield and the fertile tillers number per plant, spike length, awn length and number of grains per spike. Garavandi and Kahrizi [29] by evaluation of 20 bread wheat genotypes reported that genotypes have higher genetic diversities for grain yield, spike number per square meter, number of seed per spike, spike density and awn length in comparison with other traits. Saleh [130] also obtained the similar results. Asaduzzaman et al. [131] also believe that moisture stress reduces grain yield of mungbean and maximum negative effects of drought obtained with once irrigation during growth season. Rafiei Shirvan and Asgharipur [132] also obtained the similar results. According to Ashraf and Foolad [133] glycine betaine and proline by applying osmotic adjustment, reduce the negative effects of stress in the incidence of drought conditions.

## CONCLUSIONS

Drought tolerance consists of ability of crop to growth and production under water deficit conditions. A long term drought stress effects on plant metabolic reactions associates with, plant growth stage, water storage capacity of soil and physiological aspects of plant. Achieving a genetic increase in yield under these environments has been recognized to be a difficult challenge for plant breeders while progress in yield grain has been much higher in favorable environments. Present results showed that plants in drought stress make changes in some of their physiological and biochemical features. Also, the results of this research represented, that drought stress causes low grain yield and in drought stress conditions that the cultivars that have more relative water content (RWC), chlorophyll content, cell membrane stability and activity antioxidant enzymes are more tolerance to drought stress.

## REFERENCES

- Khan, A.S., S. Ul-Allah and S. Sadique, 2010. Genetic variability and correlation among seedling traits of Wheat (*Triticum aestivum*) under water stress. Inter. J. Agric. Biol., 2: 247-250.
- Farshadfar, E., V. Rasoli, J.A. Teixeira da Silva and M. Farshadfar, 2011. Inheritance of drought tolerance indicators in bread wheat (*Triticum aestivum* L.) using a diallel technique. Australian J. Crop Sci., 5(7): 870-878.

- Amjad Ali, M., K. Jabran, S.I. Awan, A. Abbas, M. Ehsanullah Zalkiffal and A. Tuba, 2011. Morpho-physiological diversity and its implications for improving drought tolerance in grain sorghum at different growth stages. Aust. J. Crop Sci., 5(3): 308-317.
- Ahmadizadeh, M., A. Nori, H. Shahbazi and M. Habibpour, 2011. Effects of drought stress on some agronomic and morphological traits of durum wheat (*Triticum durum* Desf.) landraces under greenhouse condition. African J. Biotechnol., 10(64): 14097-14107.
- Ahmadizadeh, M., H. Shahbazi, M. Valizadeh and M. Zaefizadeh, 2011. Genetic diversity of durum wheat landraces using multivariate analysis under normal irrigation and drought stress conditions. African J. Agric. Res., 6(10): 2294-2302.
- Reddy, A.R., K.V. Chiatanya and M. Vivekanandan, 2004. Drought-induced responses of photosynthesis and antioxidant metabolism in higher plants. J. Plant Physiol., 161: 1189-1202.
- Makbul, S., N. Saruhan-guler, N. Durmus and S. Guven, 2011. Changes in anatomical and physiological parameters of soybean under drought stress. Turk. J. Bot., 35: 369-377.
- Nouri-Ganbalani, A., G. Nouri-Ganbalani and D. Hassanpanah, 2009. Effects of drought stress condition on the yield and yield components of advanced wheat genotypes in Ardabil, Iran. J. Food, Agric. Environ., 7(3&4): 228-234.
- Apel, K. and H. Hirt, 2004. Reactive oxygen species: metabolism, oxidative stress and signal transduction, Annu. Rev. Plant Biol., 55: 373-399.
- Chaves, M.M., J.M. Aroco and J. Pereira, 2003. Understandin plant responses to drought from genes to the whole plan Funct, Plant Biol., 30: 239-264.
- 11. Chen, Z. and D.R. Gallie, 2004. The ascorbic acid redox state controls guard cell signaling and stomatal movement, Plant Cell, 16: 1143-1162.
- 12. Munns, R., 2002. Comparative physiology of salt and water stress, Plant Cell Environ., 25(2): 239-252.
- Zhu, J.K., 2003. Regulation of ion hormeostasis under salt stress, Curr. Opin Plant. Biol., 6(5): 441-445.
- 14. Moaveni, P., 2011. Effect of water deficit stress on some physiological traits of wheat (*Triticum aestivum*). Agric. Sci. Res. J., 1(1): 64-68.
- Eskandari, H. and K. Kazemi, 2010. Response of different bread wheat (*Triticum aestivum* L.) genotypes to post-anthesis water deficit. Not. Sci. Biol., 2(4): 49-52.

- Yordanov, I., V. Velikova and T. Tsonev, 2003. Plant responses to drought and stress tolerance, Bulg. J. Plant Physiol., Special Issue, 187-206.
- Abedi, T. and H. Pakniyat, 2010. Antioxidant enzyme changes in response to drought stress in ten cultivars of oilseed rape (*Brassica napus* L.). Czech J. Genet. Plant Breed., 46(1): 27-34.
- Dadbakhsh, A. A. Yazdansepas and M. Ahmadizadeh, 2011. Study drought stress on yield of wheat (*Triticum aestivum* L.) genotypes by drought tolerance indices. Advances in Environ. Biol., 5(7): 1804-1810.
- Ahmadizadeh, M., M. Valizadeh, H. Shahbazi and M. Zaefizadeh, 2011. Performance of durum wheat landraces under contrasting conditions of drought stress. World Applied Sciences Journal, 13(5): 1022-1028.
- Mohammadi, A.A., G. Saeidi and A. Arzani, 2010. Genetic analysis of some agronomic traits in flax (*Linum usitatissimum* L.). Aust. J. Crop Sci., 4(5): 343-352.
- Nouri, A., A. Etminan, J.A. Teixeira Da Silva and R. Mohammadi, 2011. Assessment of yield, yield related traits and drought tolerance of durum wheat genotypes (*Triticum turjidum* var. *durum* Desf.). Aust. J. Crop Sci., 5(1): 8-16.
- 22. Nayyar, H. and D. Gupta, 2006. Differential sensitivity of C3 and C4 plants to water deficit stress: Association with oxidative stress and antioxidants. Environ. and Exp. Bot., 58: 106-113.
- Jaleel, C.A., P. Manivannan, A. Wahid, M. Farooq, H.J. Al-Juburi, R. Somasundaram and R. Panneerselvam, 2009. Drought stress in plants: A review on morphological characteristics and pigments composition. International J. Agric. and Biol., 11(1): 100-105.
- Mollasadeghi, V., A.A. Imani, R. Shahryari and M. Khayatnezhad, 2011. Classifying bread wheat genotypes by multivariable statistical analysis to achieve high yield under after anthesis drought. Middle-East J. Sci. Res., 7(2): 217-220.
- Plaut, Z., B.J. Butow, C.S. Blumenthal and C.W. Wrigley, 2004. Transport of dry matter into developing wheat kernels. Field Crops Res., 96: 185-198.
- Wajid, A., A. Hussain, M. Maqsood, A. Ahmad and M. Awais, 2002. Influence of sowing date and irrigation levels on growth and grain yield of wheat. Pak. J. Agri. Sci., 39(1): 22-24.

- Jahfari, H.A., 2004. Modeling the growth, radiation use efficiency and yield of new wheat cultivars under varying nitrogen rats. M.Sc. Thesis, Dept. Agron. Univ. Agri. Faisalabad, Pakistan.
- Rafique, M., 2004. Effect of different levels of irrigation on growth, water use efficiency and yield of different wheat cultivars. Ph.D. Thesis Department of Agronomy, University of Agriculture Faisalabad, Pakistan.
- Garavandi, M. and D. Kahrizi, 2010. Evaluation of genetic diversity of bread wheat genotypes for phonologic and morphologic traits. The 11<sup>th</sup> Crop Science and Plant Breeding Congress Iran, pp: 537-541.
- Mirbahar, A.A., G.S. Markhand, A.R. Mahar, S.A. Abro and N.A. Kanhar, 2009. Effect of water stress on yield and yield components wheat (*Triticum aestivum* L.) varieties. Pak. J. Bot., 41(3): 1303-1310.
- Martinez, J.P., H. Silva, J.F. Ledent and M. Pinto, 2007. Effect of drought stress on the osmotic adjustment, cell wall elasticity and cell volume of six cultivars of common beans (*Phaseolus vulgaris* L.). European J. Agron., 26: 30-38.
- Jaleel, C.A., R. Gopi, B. Sankar, M. Gomathinayagam and R. Panneerselvam, 2008. Differential responses in water use efficiency in two varieties of *Catharanthus roseus* under drought stress. Comp. Rend. Biol., 331: 42-47.
- Zhao, C.X., L.Y. Guo, C.A. Jaleel, H.B. Shao and H.B. Yang, 2008. Prospects for dissecting plantadaptive molecular mechanisms to improve wheat cultivars in drought environments. Comp. Rend. Biol., 331: 579-586.
- Terzi, R. and A. Kadioglu, 2006. Drought stress tolerance and the antioxidant enzyme system in *ctenanthe setosa*. Acta Biologica Cracoviensia Series Botanica, 48(2): 89-96.
- 35. AL-Ghamdi, A.A., 2009. Evaluation of oxidative stress tolerance in two wheat (*Triticum aestivum*) cultivars in response to drought. International J. Agric. and Biol., 11: 7-12.
- Upadhyaya, H., M.H. Khan and S.K. Panda, 2007. Hydrogen peroxide induces oxidative stress in detached leaves of *Oryza Sativa* L. Gen. Appl. Plant Physiol., 33(1-2): 83-95.
- Sai Kachout, S., A. Ben Mansoura, J.C. Leclerc, R. Mechergui, M.N. Rejeb and Z. Ouerghi, 2009. Effects of heavy metals on antioxidant activities of Atriplex hortensis and A. rosea. JFood, Agric. and Environ., 7(3&4): 938-945.

- Ahmadizadeh, M., M. Valizadeh, M. Zaefizadeh and H. Shahbazi, 2011. Antioxidative protection and electrolyte leakage in durum wheat under drought stress condition. J. Appl. Sci. Res., 7(3): 236-246.
- Malik, A.A., W.G. Li, L.N. Lou, J.H. Weng and J.F. Chen, 2010. Biochemical/ physiological characterization and evaluation of in vitro salt tolerance in cucumber. African J. Biotechnol., 9(22): 3284-3292.
- 40. Pignocchi, C. and C.H. Foyer, 2003. Apoplastic ascorbate metabolism and its role in the regulation of cell signaling. Curr. Opin. Plant Biol., 6: 379-389.
- Muller, M., I. Hernandez, L. Alegre and S. Munne-Bosch, 2006. Enhanced a-tocopherol quinine levels and xanthophyll cycle de-epoxidation in rosemary plants exposed to water deficit during a Mediterranean winter. J. Plant Physiol., 163: 601-606.
- Xu, P.L., Y.K. Guo, J.G. Bai, L. Shang and X.J. Wang, 2008. Effects of long-term chilling on ultrastructure and antioxidant activity in leaves of two cucumber cultivars under low light. Physiologia Plantarum, 132: 467-478.
- Havaux, M., 1998. Carotenoids as membrane stabilizers in chloroplasts. Trends Plant Sci., 3: 147-151.
- Farooq, M., A. Wahid, N. Kobayashi, D. Fujita and S.M.A. Basra, 2009. Plant drought stress: effects, mechanisms and management. Agron. Sustain. Dev., 29: 185-212.
- Tertivanidis, K., C. Goudoula, C. Vasilikiotis, E. Hassiotou, R. Perl-Treves and A. Tsaftaris, 2004. Superoxide dismutase transgenes in sugarbeets confer resistance to oxidative agents and the fungus C. beticola. Transgenic Res., 13(3): 225-233.
- 46. Zheng, Q., J. Bao, L.K. Liang and X. Xiao, 2005. Effects of antioxidants on the plant regeneration and GUS expressive frequency of peanut (Arachis hypogaea) explants by Agrobacterium tumefaciens. Plant Cell Tissue and Organ Culture, 81(1): 83-90.
- Shao, H.B., Z.S. Liang and M.A. Shao, 2005. Changes of some anti-oxidative enzymes under soil water deficits among 10 wheat genotypes at maturation stage. Colloids Surf. B: Biointerfaces, 45: 7-13.
- Guo, Z., W. Ou, S. Lu and Q. Zhong, 2006. Differential responses of antioxidative system to chilling and drought in four rice cultivars differing in sensitivity. Plant Physiology and Biochemistry, 44: 828-836.

- Gunes, A., D. Pilbeam, A. Inal and S. Coban, 2008. Influence of silicon on sunflower cultivars under drought stress, I: Growth, antioxidant mechanisms and lipid peroxidation. Commun. Soil Science and Plant Nutrition, 39: 1885-1903.
- Sreenivasulu, N., B. Grimm, U. Wobus and W. Weschke, 2000. Differential response of antioxidant compounds to salinity stress in salttolerant and salt-sensitive seedlings of foxtail millet (*Setaria italica*). Physiologia Plantarum, 109: 435-442.
- 51. Mittova, V., M. Volokita, M. Guy and M. Tal, 2000. Activities of SOD and the ascorbate-glutathione cycle enzymes in subcellular compartments in leaves and roots of the cultivated tomato and its wild salttolerant relative *Lycopersicon pennellii*. Physiologia Plantarum, 110: 42-51.
- 52. Bor, M., F. Ozdemir and I. Turkan, 2003. The effect of salt stress on lipid peroxidation and antioxidants in leaves of sugar beet Beta vulgaris L and wild beet Beta maritima L. Plant Sci., 164: 77-84.
- 53. Zaefyzadeh, M., R.A. Quliyev, S.M. Babayeva and M.A. Abbasov, 2009. The effect of the interaction between genotypes and drought stress on the superoxide dismutase and chlorophyll content in durum wheat landraces. Turk. J. Biol., 33: 1-7.
- Shahbazi, H., M. Taeb, M.R. Bihamta and F. Darvish, 2010. Inheritance of antioxidant activity of bread wheat under terminal drought stress. American-Eurasian J. Agric. and Environ. Sci., 8(6): 680-684.
- 55. Acar, O., I. Turkan and F.O. Zdemir, 2001. Superoxide dismutase and peroxidase activities in drought sensitive and resistant barley (*Hordeum vulgare* L.) varieties. Acta Physiologiae Plantarum, 3: 351-356.
- Paknejad, F., M. Nasri, H.M. Tohidi-Moghadam, H. Zahedi and M. Jami-Alahmadi, 2007. Effects of drought stress on chlorophyll fluorescence parameters, chlorophyll content and grain yield of wheat cultivars. J. Biological Sci., 7(6): 841-847.
- 57. Nori, A., M. Ahmadizadeh, H. Shahbazi and S. Aharizad, 2011. Evaluation of physiological responses of durum wheat landraces (*Triticum Durum*) to terminal drought stress. Advances in Environ. Biol., 5(7): 1947-1954.
- Cornic, G. and C. Fresneau, 2002. Photosynthetic carbon reduction and carbon oxidation cycles are the main electron sinks for photosystem 2 activity during a mild drought. Annals Bot., 89: 887-894.

- Havaux, M., 1992. Stress tolerance of photosystem II in vivo. Antagonistic effects of water, heat and photoinhibition stresses. Plant Physiol., 100: 424-432.
- Yordanov, I., V. Velikova and T. Tsonev, 1999. Influence of drought, high temperature and carbamide cytokinin 4-PU-30 on photo synthetic activity of plants. 1. Changes in chlorophyll fluorescence quenching. Photosynthetica, 37: 447-457.
- Noctor, G., S. Veljovic-Jovanovic, S. Driscoll, L. Novitskaya and C.H. Foyer, 2002. Drought and oxidative load in the leaves of C3 plants: a predominant role of photorespiration? Annals Bot., 89: 841-850.
- Pandey, R.M. and R. Singh, 2010. Genetic studies for biochemical and quantitative characters in grain amaranth (*Amaranthus hypochondriacus* L.). Plant Omics J., 3(4): 129-134.
- Paknejad, F., M. Mirakhori, M. Jami Al-Ahmadi, M.R. Tookalo, A.R. Pazoki and P. Nazeri, 2009. Physiological response of soybean (*Glycine max*) to foliar application of methanol under different soil moistures. American J. Agric. and Biological Sci., 4(4): 311-318.
- Kuroda, M., T. Qzawa and H. Imagawa, 1990. Changes in chloroplast peroxidase activities in relation to chlorophyll loss in barley leaf segments. Physiologia Plantarum, 80: 555-560.
- Shamsi, K., 2010. The effects of drought stress on yield, relative water content, proline, soluble carbohydrates and chlorophyll of bread wheat cultivars. J. Animal and Plant Sci., 8(3): 1051-1060.
- Thomas, J., A. Jeffrey, C. Atsuko and M.K. David, 2005. Reguling the proton budget of higher plant photosynthesis. Proc. Natl. Acade. Sci. USA., 102(27): 9709-9713.
- Wang, F.H., G.X. Wang, Y. Lix, J.L. Huvang and J.K. Zheng, 2008. Heredity, physiology and mapping of chlorophyll content gene in rice (*Oryza sativa* L). J. Plant Physiol., 165: 324-330.
- Habibi, F., GH. Normahamadi, H. Heidary Sharif Abad, A. Eivazi and E. Majidi Heravan, 2011. Effect of cold stress on cell membrane stability, chlorophyll a and b contain and proline accumulation in wheat (*Triticum aiestivum* L.) variety. African J. Agric. Res., 6(27): 5854-5859.
- Ashraf, M.Y., A.R. Azmi, A.H. Khan and S.A. Ala, 1994. Effect of water stress on total phenols, Peroxidase activity and chlorophyll content in wheat. Acta Physiologiae Plantarum, 16(3): 185-191.

- Sgherri, C.L.M., C. Pizino and F. Navari- Izzo, 1993. Chemical changes and O2 Production in thylakoid membrances under water stress. Plant Physiol., 87(2): 211-216.
- 71. Pastori, G.M. and V.S. trippi, 1992. Oxidative stress induces high rate of glutathione reductase synthesis in a drought resistant maize strain. Plant Cell Physiology, 33: 957-961.
- 72. Massacci, A., S.M. Nabiev, L. Pietrosanti, S.K. Nematov, T.N. Chernikova, K. Thor and J. Leipner, 2008. Response of the photosynthetic apparatus of cotton (*Gossypium hirsutum*) to the onset of drought stress under field conditions studied by gas-exchange analysis and chlorophyll fluorescence imaging. Plant Physiol. Biochem., 46: 189-195.
- Kiani, S.P., P. Maury, A. Sarrafi and P. Grieu, 2008. QTL analysis chlorophyll fluorescence parameters in sunflower (*Helianth annuus* L.) under well-watered and water-stressed conditions. Pla. Sci., 175: 565-573.
- Tahkokorpi, M., K. Taulavuori, K. Laine and E. Taulavuori, 2007. After effects of drought-related winter stress in previous and current year stems of *Vaccinium myrtillus* L. Environ. Exp. Bot., 61: 85-93.
- 75. Kulshreshtha, S., D.P. Mishra and R.K. Gupta, 1987. Changes in content of chlorophyll, proteins and lipids in whole chloroplast and chloroplast membrane fractions at different leaf water potentials in drought resistant and sensitive genotypes of wheat. Photosynthetica., 21(1): 65-70.
- Rizza, F., D. Pagani, A.M. Stanca and L. Cattivelli, 2001. Use of chlorophyll fluorescence to evaluate the cold acclimation and freezing tolerance of winter and spring oats, S. Afr. J. Bot., 120: 389-396.
- Kocheva, K., P. Lambrevb, G. Georgiev, V. Goltsev and M. Karabaliev, 2004. Evaluation of chlorophyll fluorescence and membrane injury in the leaves of barley cultivars under osmotic stress. Bioelectrochemistry, 63: 121-124.
- Behra, R.K., P.C. Mishra and N.K. Choudhury, 2002. High irradiance and water stress induce alterations in pigment composition and chloroplast activities of primary wheat leaves. J. Plant Physiol., 159: 967-973.
- Percival, G.C. and C.N. Sheriffs, 2002. Identification of drought-tolerant woody perennials using chlorophyll fluorescence. J. Arboriculture, 28(5): 215-223.
- 80. Ramin, A.A., R.K. Prange, J.M. DeLong and P.A. Harrison, 2008. Evaluation of relationship between moisture loss in grapes and chlorophyll fluorescence measured as F0 (F- $\alpha$ ) reading. J. Agric. Sci. Technol., 10: 471-479.

- Flagella, Z., D. Pastore, R.G. Campanile and N. Di-Fonzo, 1995. The quantum yield or photosynthetic electron transport evaluated by chlorophyll fluorescence as an indicator of drought tolerance in durum wheat. J. Agric. Sci. Cambridge, 125: 325-329.
- Vazan, S., 2002. Effects of chlorophyll parameters and photosynthesis efficiency in difference beet. Assay Ph.D Islamic azad university science and research Tehran-Branch, pp: 285.
- Reddy, T.Y., V.R. Reddy and V. Anbumozhi, 2003. Physiological responses of groundnut (*Arachis hypogea* L.) to drought stress and its amelioration: a critical review. Plant Growth Regulation, 41: 75-88.
- Siddique, M.R.B., A. Hamid and M.S. Islam, 2000. Drought stress effects on water relations of wheat. Bot. Bull. Acad. Sin., 41: 35-39.
- Mationn, M.A., J.H. Brown and H. Ferguon, 1989. Leaf water potential, relative water content and diffusive resistance as screening techniques for drought resistance in barley. Agron. J., 81: 100-105.
- 86. Subrahmanyam, D., N. Subash, A. Haris and A.K. Sikka, 2006. Influence of water stress on leaf photosynthetic characteristics in wheat cultivars differing in their susceptibility to drought. Photosynthetica, 44(1): 125-129.
- Tas, S. and B. Tas, 2007. Some physiological responses of drought stress in wheat genotypes with different ploidity in Turkiye. World J. Agri. Sci., 3(2): 178-183.
- Allahmoradi, P., M. Ghobadi, S. Taherabadi and S. Taherabadi, 2011. Physiological aspects of mungbean (*Vigna radiata* L.) in response to drought stress. International Conference on Food Engineering and Biotechnol., 9: 272-275.
- Mohammadkhani, N. and R. Heidari, 2008. Droughtinduced accumulation of soluble sugars and proline in two maize varieties. W. Appl. Sci. J., 3(3): 448-453.
- Liu, W.J., S. Yuan, N.H. Zhang, T. Lei, H. Duan, H.G. Liang and H.H. Lin, 2006. Effect of water stress on photosystem 2 in two wheat cultivars. Biological Plantarum, 50(4): 597-602.
- Gonzalez, A., I. Martín and L. Ayerbe, 2008. Yield and osmotic adjustment capacity of barley under terminal water stress conditions. J. Agron. Crop Sci., 194(2): 81-91.
- 92. Collado, M.B., M.J. Arturi, M.B. Aulicino and M.C. Molina, 2010. Identification of salt tolerance in seedling of maize (*Zea mays* L.) with the cell membrane stability trait. International Research J. Plant Sci., 1(5): 126-132.

- Seidler-Lozykowska, K., H. Bandurska and J. Bocianowski, 2010. Evaluation of cell membrane injury in caraway (*carum carvi* L.) genotypes in water deficit conditions. Acta Societatis Botanicorum Poloniae., 79(2): 95-99.
- 94. Habibpor, M., M. Valizadeh, H. Shahbazi and M. Ahmadizadeh, 2011. Study of drought tolerance with cell membrane stability testing and relation with the drought tolerance indices in genotypes of wheat (*Triticum Aestivum* L.). W. Appl. Sci. J., 13(7): 1654-1660.
- 95. Blum, A. and A. Ebercon, 1981. Cell membrane stability as a measure of drought and heat tolerance in wheat. Crop Sci., 21: 43-47.
- 96. Premachandra, G.S. and T. Shimada, 1987. The measurement of cellmembrane stability using polyethylene glycol as a drought tolerance test in wheat. Jpn. J. Crop Sci., 56: 92-98.
- 97. Vasquez-Tello, A., Y. Zuily-Fodil, A.T. Pham Thi and J.B. Viera Da Silva, 1990. Electrolyte and Pi leakages and soluble sugar content as physiological tests for screening resistance to water stress in Phaseolus and Vigna species. J. Exp. Bot., 41: 827-832.
- Bandurska, H., A. Stroinski and M. Zielezinska, 1997. Effects of water deficit stress on membrane properties, lipid peroxidation and hydrogen peroxide metabolism in the leaves of barley genotypes. Acta Soc. Bot. Pol., 66: 177-183.
- Saneoka, H., R.E.A. Moghaieb, G.S. Premachandra and K. Fujita, 2004. Nitrogen nutrition and water stress effects on cell membrane stability and leaf water relations in agrostis palustris huds. Environ. Expermin. Bot., 52: 131-138.
- 100. Azizi-e-Chakherchaman, S.H., H. Mostafaei, A. yari, M. Hassanzadeh, S.H. Jamaati-e-Somarin and R. Easazadeh, 2009. Study of relationships of leaf relative water content, cell membrane stability and duration of growth period with grain yield of lentil under rain-fed and irrigated conditions. Res. J. Biological Sci., 4(7): 842-847.
- 101. Blum, A., N. Klueva and H.T. Nguyen, 2001. Wheat cellular thermotolerance is related to yield under stress. Euphytica., 117: 117-123.
- 102. Tripathy, J.N., J. Zhang, S. Robin, T.T. Nguyen and H.T. Nguyen, 2000. QTLs for cell-membrane stability mapped in rice (*Oryza Sativa* L.) under drought stress. Theor. App. Genet., 100: 1197-1202.

- 103. Premachandra, G.S., H. Saneoka, K. Fujita and S. Ogata, 1992. Leaf water relations, osmotic adjustment, cell membrane stability, epicuticular wax load and growth as affected by increasing water deficits in sorghum. J. Exp. Bot., 43: 1569-1576.
- 104. Arvin, M.J. and D.J. Donnelly, 2008. Screening potato cultivars and wild species to abiotic stresses using an electrolyte leakage bioassay. J. Agric. Sci. Technol., 10: 33-42.
- 105. Garg, G., 2010. Response in germination and seedling growth in Phaseolus mungo under salt and drought stress. J. Environ. Biol., 31: 261-264.
- 106. Ahmadizadeh, M., M. Valizadeh, M. Zaefizadeh and H. Shahbazi, 2011. Evaluation of interaction between genotype and environments in term of germination and seedling growth in durum wheat landraces. Advances in Environ. Biol., 5(4): 551-558.
- 107. Shen, Y.Y., Y. Li and S.H.G. Yan, 2003. Effects of salinity on germination of six salt-tolerant forage species and their recovery from saline conditions. New Zealand J. Agric. Res., 46(3): 263-269.
- Macke, A. and I.A. Ungar, 1971. The effect of salinity on germination and early growth of *Puccinellia nuttalliana*. Canadian J. Botany, 49: 515-520.
- 109. Khan, M.A. and I.A. Ungar, 1999. Effect of salinity on seed germination of *Triglochin Maritima under* various temperature regimes. Great Basin Naturalist, 59(2): 144-150.
- 110. Turkan, I., M. Bor, F. Zdemir and H. Koca, 2005. Differential responses of lipid peroxidation and antioxidants in the leaves of drought-tolerant P. acutifolius and drought-sensitive P. vulgaris subjected to polyethylene glycol mediated water stress, Plant Sci., 168: 223-231.
- 111. Gholamin, R., M. Khayatnezhad, S. jamaati-e-Somarin and R. Zabihi-e-Mahmoodabad, 2010. Effects of polyethylene glycol and NaCl stress on two cultivars of wheat (*Triticum durum*) at germination and early seedling stages. American-Eurasian J. Agric. And Environ. Sci., 9(1): 86-90.
- 112. Alaei, M., M. Zaefizadeh, M. Khayatnezhad, Z. Alaei and Y. Alaei, 2010. Evaluation of germination properties of different durum wheat genotypes under osmotic stress. Middle-East J. Scientific Res., 6(6): 642-646.
- 113. Al-Karaki, G.N., A. Al-Ajmi and Y. Othman, 2007. Seed germination and early root growth of three barley cultivars as affected by temperature and water stress. American-Eurasian J. Agric. and Environ. Sci., 2(2): 112-117.

- 114. Homayoun, H., M. Sam-Daliri and P. Mehrabi, 2011. Study of PEG stress effects on wheat (*Triticum aestivum* L.) cultivar at germination stage. Middle-East J. Scientific Res., 9(1): 71-74.
- 115. Dhanda, S.S., G.S. Sethi and R.K. Behl, 2004. Indices of drought tolerance in wheat genotypes at early stages of plant growth. J. Agron. Crop Sci., 190: 6-12.
- 116. Guoth, A., I. Tari, A. Galle1, J. Csiszar, L. Cseuz and L. Erdei, 2008. Changes in photosynthetic performance and ABA levels under osmotic stress in drought tolerant and sensitive wheat genotypes. Acta Biologica Szegediensis, 52(1): 91-92.
- 117. Ambika Rajendran, R., A.R. Muthiah, A. Manickam, P. Shanmugasundaram and A. John Joel, 2011. Indices of drought tolerance in sorghum (*Sorghum bicolor* L. Moench) genotypes at early stages of plant growth. Research J. Agric. and Biological Sci., 7(1): 42-46.
- Tahir, M.H.N., M. Imran and M.K. Hussain, 2002. Evaluation of sunflower (*Helianthus annuus* L.) inbred lines for drought tolerance. Int. J. Agric. Biol., 3: 398-400.
- 119. Takele, A., 2000. A greenhouse experiment was conducted to understand the seedling germination behavior and growth of sorghum genotypes. Acta Agron. Hungarica, 48(1): 95-102.
- 120. Grzesiak, S., W. Filek, G. Skrudlik and B. Niziol, 1996. Screening for drought tolerance: Evaluation of seed germination and seedling growth for drought resistance in legume plants. J. Agron. Crop Sci., 177(4): 245-252.
- 121. Habibpor, M., M. Valizadeh, H. Shahbazi and M. Ahmadizadeh, 2011. Genetic diversity and correlation among agronomic and morphological traits in wheat genotypes (*Triticum Aestivum* L.) under influence of drought. Advances in Environ. Biol., 5(7): 1941-1946.
- 122. Ahmadizadeh, M., M. Valizadeh, H. Shahbazi, M. Zaefizadeh and M. Habibpor, 2011. Morphological diversity and interrelationships traits in durum wheat landraces under normal irrigation and drought stress conditions. Advances in Environ. Biol., 5(7): 1934-1940.
- 123. Khayatnezhad, M., M. Zaefizadeh, R. Gholamin, S. jamaati-e- Somarin and R. Zabihi-e-Mahmoodabad, 2010. Study of morphological traits of wheat cultivars through factor analysis. American-Eurasian J. Agric. and Environ. Sci., 9(5): 460-464.

- 124. Abinasa, M., A. Ayana and G. Bultosa, 2011. Genetic variability, heritability and trait associations in durum wheat (*Triticum turgidum* L. var. *durum*) genotypes. African J. Agric. Res., 6(17): 3972-3979.
- 125. Edward, D. and D. Wright, 2008. The effects of winter water-logging and summer drought on the growth and yield of winter wheat (*Triticum aestivum* L.) European J. Agron., 28: 234-244.
- 126. Kamara, A.Y., A. Menkir, B. Badu-Apraku and O. Ibikunle, 2003. The influence of drought stress on growth, yield and yield components of selected maize genotypes. J. Agric. Sci., 141: 43-50.
- 127. Specht, J.E., K. Chase, M. Macrander, G.L. Graef, J. Chung, J.P. Markwell, M. Germann, J.H. Orf and K.G. Lark, 2001. Soybean response to water. A QTL analysis of drought tolerance. Crop Sci., 41: 493-509.
- 128. Heydari, B., Gh. Saeedi, B.A. Seyyed Tabatabaei and K. Soenaga, 2006. Evaluation of genetic diversity and estimation of heritability of some quantity traits in double haploid lines of wheat. Iranian J. Agric. Sci., 37(2): 347-356.

- 129. Bahari, M. and H. Sabzi, 2004. Correlation of pheno-morphological traits with grain yield durum wheat genotypes. Eighth Iranian Congress of Agronomy and Plant Breeding, September, Faculty of Agricultural Sciences, University of Guilan, pp: 20.
- 130. Saleh, A.H., 2011. Perfomance, correlation and path coefficient analysis for grain yield and its related traits in diallel crosses of bread wheat under normal irrigation and drought conditions. World J. Agric. Sci., 7(3): 270-279.
- Asaduzzaman, F. K., J. Ullah and M. Hasanuzzaman, 2008. Response of mungbean (*Vigna radiata* L.) to nitrogen and irrigation management. American-Eurasian J. Scientific Res., 3(1): 40-43.
- Rafiei Shirvan, M. and M.R. Asgharipur, 2009. Yield reaction and morphological characteristics of some mungbean genotypes to drought stress. J. Modern Agriculture Knowledge, 5(15): 67-76.
- 133. Ashraf, M. and M.R. Foolad, 2007. Roles of glycine betaine and proline in improving plant abiotic stress resistance. Environ. and Exp. Bot., 59: 206-216.