

Assessment of Antimicrobial Efficacy of Silver Nanoparticles Versus Calcium Hydroxide as Intra Canal Medication (*In vivo* Study)

¹Soha A Abdou, ²Medhat A. Kataia, ³Magdy M.M. Ali,
⁴Reham El Sayed Hassan and ⁵Sahar I. Negm

¹Department of Endodontics, Research Institute of Ophthalmology, Giza, Egypt

²Department of Endodontics, Faculty of Oral and Dental Medicine Cairo University, Cairo, Egypt

³Department of Endodontics, Faculty of Dental Medicine El Minia University, El Minia, Egypt

⁴Department of Medical Microbiology and Immunology, Research Institute of Ophthalmology, Giza, Egypt

Abstract: The aim of the present study was to prepare and characterize silver nanoparticles AgNPs paste as intra canal medication and compare its antimicrobial activity against calcium hydroxide Ca(OH)₂ paste. Twenty patients with necrotic pulp tissues were divided into 2 groups according to the type of medicament used. Two microbiological samples were taken, one after access cavity preparation and the second after removal of intra canal medication which was inserted for one week after mechanical preparation. The colonies were identified by culture characteristics, microscopic examination, biochemical reactions and polymerase chain reaction (PCR). Colony forming units CFUs were counted and data were analyzed. There was a statistically significant decrease in CFUs between before and after treatment with Ca(OH)₂ group and 0.03 mg/ml AgNPs group with 17.95 % and 42.86 % percentage reduction respectively. But in AgNPs group, there was no statistically significant decrease in number of CFUs for *Candida* between before and after treatment. AgNPs paste showed good antibacterial effect and less antifungal effect.

Key words: Intracanal medication • Calcium hydroxide • Silver nanoparticles • Necrotic pulp

INTRODUCTION

Microorganisms are the main causative factors in the development of necrosis of the dental pulp and the formation of periapical lesions. Although we try to clean and irrigate the root canal system properly, some bacteria still remain entrapped in the dentinal tubules. It has been shown that, if the canal is not dressed with a disinfectant between visits, microorganisms will multiply rapidly within days to near the original numbers [1].

Several studies demonstrated that, some of microorganisms were resistant to the most commonly used intra canal medication calcium hydroxide Ca (OH)₂ [2]. Recent studies have focused on using nanoparticles to disinfect root canals. Silver is an effective antimicrobial agent with low toxicity and broad spectrum activity Among the various options available.

AgNPs which are one of the most commercialized nanomaterials. They are widely applied as biocides for

their strong antimicrobial activity [3]. It seems valuable to evaluate the antimicrobial efficacy of AgNPs as intra canal medication and compare it with the commonly used intra canal medication, Ca (OH)₂.

MATERIALS AND METHODS

Preparation of Silver Nanoparticles: Silver nanoparticles (AgNPs) have been prepared by chemical reduction method as reported by Turkevich *et al.* [4] and, Lee and Meisel [5]. A solution of silver nitrate (AgNO₃) has been used as Ag⁺ ions precursor. The color of the solution slowly turned into grayish yellow, indicating the reduction of the Ag⁺ ions to Ag nanoparticles.

Patient Selection and Classification: Twenty patients medically free with necrotic pulp tissues of single rooted teeth with no periapical lesion, their age ranges from 20 to 50 years of both sexes were selected. Patients were

randomly divided into two groups according to type of medicament used. The medicaments used were Ca(OH)₂ paste and 0.03 mg/ml AgNPs paste.

Endodontic Treatment Was Done in Two Visits.

The First Visit: After approval of the ethics committee of Minia University, access cavities were prepared using sharp round bur mounted on a high-speed hand piece. The first microbiological samples were taken using H-files size 30. The files were inserted into a sterile Wasserman tubes containing 1 ml of sterile brain heart infusion (BHI) broth. The tubes were incubated at 37°C for one day before culturing. Working length was determined by periapical radiographs using bisecting angle technique. Mechanical preparations were completed using REVO-S * rotary system mounted in endodontic motor **. Irrigation was done using normal saline at all procedure steps. AgNPs was placed inside the canals using sterile plastic syringe. The access cavities were closed using sterile cotton pellet and temporary filling.

The Second Visit: After seven days, Intra canal medicaments were removed by irrigation with 10ml of sterile normal saline. The second microbiological samples were taken and cultured. After confirmation of the length of the master cone by periapical radiographic X ray; obturation was done. The obturation was done using Revo-S gutta percha points.

* MICRO MEGA.

** Motor X smart E-Cube, Korea.

Antimicrobial Effect Evaluation: After incubation of microbiological samples, BHI broth was spread on different types of agar plates. After culturing, the blood and Sabouraud agar plates were incubated aerobically at 37° C for two days. The Wilkins agar plates were incubated anaerobically at 37°C for five days. The colonies were identified by Culture characteristics [6], Microscopic examination, Biochemical reactions (Catalase test) [7] and Polymerase chain reaction (PCR).

PCR: DNA extraction was done to the samples according to Qiagen DNA extraction protocol for bacteria (DNeasy blood and tissue handbook 2016) followed by PCR amplification with specific primers using hot start master mix method [8]. Detection of amplified products was done using Agarose gel electrophoresis [9] and Real time curve analysis [10].

Counting of the Types of Microorganisms: After incubation, serial dilution was done to 1/100000 for all specimens. Colonies of each microorganism were counted from the plates and the colony forming units CFUs /ml were calculated according to the following equation: - CFUs / ml = number of colonies × dilution × volume of infected BHI broth [11].

Follow up and Prognosis: Follow up was done for all patients after 1month and 3 months by clinical examination and radiographic evaluation.

Statistical Analysis: Data were presented as mean, standard deviation (SD), frequency and percentage. Dependent t-test used to compare between before and after intra canal medication for Log¹⁰ CFU/ml. Wilcoxon signed rank test was used for detection of each microorganism.

RESULTS

Nanoparticles Characterization Results

Determination of Size and Shape of Silver Nanoparticles AgNPs: The particle size and shape were determined using Transmission electron microscope (TEM) which was performed on JEOL JEM-2100 at an accelerating voltage of 200 kV. The average size of the particles was 50 ± 5 nano microns (nm) and the particles were spherical in shape.

Determination of Optical Properties of AgNPs: The optical properties of the prepared nanoparticles were characterized using UV-Vis absorption spectra which were obtained on an Ocean Optics USB2000+VIS-NIR Fiber optics spectrophotometer. Optical Properties (Abs.): λ_{max} = 401 nm.

In vivo Antimicrobial Effect of the Tested Intra Canal Medication:

By Colony Forming Units CFUs: In Ca(OH)₂ group: There was a statistically significant decrease in number of colony forming units CFUs between before and after treatment with Ca(OH)₂ with 17.95 % percentage reduction. In cocci, bacilli and *candida*; there was a statistically significant decrease in number of CFUs between before and after treatment with 23.29, 21.13 and 23.90 % percentage reduction, respectively (Table 1, Fig. 1).

Table 1: Mean and standard deviation (SD) for colony forming units (10^{10} CFUs/ml) for different microorganisms

		Time				% of reduction	p-value
		Pre-treatment		Post-treatment			
		Mean	SD	Mean	SD		
Cocci	Group I (Ca(OH) ₂)	7.3	0.95	5.6	2.41	-23.29%	0.025*
	Group II (AgNPs)	6.8	1.4	1.3	1.77	-80.88%	≤0.001*
Bacilli	Group I (Ca(OH) ₂)	7.1	1.29	5.6	1.58	-21.13%	0.003*
	Group II (AgNPs)	7.1	1.45	3.2	1.4	-54.93%	≤0.001*
Candida	Group I (Ca(OH) ₂)	6.57	1.9	5	2.1	-23.90%	0.026*
	Group II (AgNPs)	5.5	1.38	3.29	1.25	-40.18%	0.195 NS
Total	Group I (Ca(OH) ₂)	7.8	1.23	6.4	1.51	-17.95%	0.01*
	Group II (AgNPs)	7	1.89	4	1.25	-42.86%	≤0.001*

*=significant, NS= Non-Significant

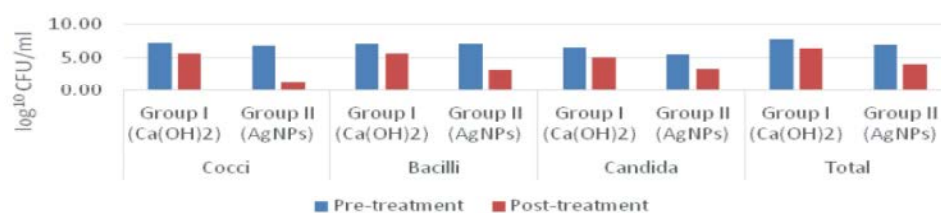


Fig. 1: Bar chart showing the mean colony forming units (10^{10} CFUs/ml) for different microorganisms



Fig. 2: Detection of the sequence of the primers of *Actinomyces* at 100 nm to 600 nm

In 0.03 mg/ml AgNPs: There was a statistically significant decrease in number of colony forming units CFUs between before and after treatment with 0.03 mg/ml AgNPs with 42.86 % percentage reduction. In cocci and bacilli; there was a statistically significant decrease in number of CFUs between before and after treatment with 80.88 and 54.93 % percentage reduction, respectively. But in *Candida*, there was no statistically significant decrease in number of CFUs between before and after treatment with 40.18 % percentage reduction (Table 1, Fig. 1).

By PCR:

Effect of Intra Canal Medication Type on Identified Microorganisms: In both intra canal medication groups, there was no statistically significant difference in types of microorganisms before and after treatment.

DISCUSSION

A new silver nanotechnology has become favourable due to its effects against biofilms [12]. The

AgNPs interact with multiple targets in the microbial cell, such as cell membrane, enzymes and plasmids, simultaneously providing the bacteria least capacity to gain resistance [13]. The size of AgNPs was 50 nm as the size of the particle was related to the antimicrobial activity; the smaller particles give more bactericidal effects than larger particles [14- 16]. Concentration of AgNPs used in this study was 0.03 mg/ml. As reported by Xinping [3] this concentration had the most effective antimicrobial properties with least toxicity. Ca(OH)₂ was chosen as the base line because of its prevalent clinical application [17, 18]. Despite the limitations of Ca(OH)₂, however its antimicrobial action as intra canal medication is well documented [18, 19].

All microbiological steps which were done in this study were done under laminar air flow to prevent contamination [3]. Polymerase chain reaction (PCR) was done for accurate confirmation of the types of microorganisms [20, 21]. The medicaments were removed using saline before bacterial sampling because it has no any antimicrobial effect so it does not affect the results of the study [22, 23]. Counting the colony forming units (CFUs) was performed because it allows the calculation of the quantity of microorganisms within the root canals [24].

The results of the present study showed that the percentage reduction of numbers of microorganisms before and after treatment with 0.03 mg/ml AgNPs paste group was significantly higher than Ca(OH)₂ paste group. This may be attributed to high affinity of silver to negatively charged molecules within the bacterial cells which inactivate critical functions of bacterial cells and prevent bacterial growth and biofilm formation. Silver nanoparticles are effective against a broad range of pathogens, which form biofilm, such as *E. coli*, *Strep. pneumoniae*, *S. aureus* and *Actinomyces niger* [25]. The antimicrobial efficacy of Ca(OH)₂ is related to hydroxyl ion release in an aqueous environment, thick mixtures paste of Ca(OH)₂ may not be ideal as intra canal dressings [26]. Also the possibility of neutralizing the high alkalinity of Ca(OH)₂ by the dentin and biofilm matrix of bacteria, which lead to reduced antibacterial effect [27, 28]. This result is in accordance with the results of Javidi [29] who stated that Ca(OH)₂ alone was unable to eliminate *E. faecalis* sufficiently even after 1 week. However, combining Ca(OH)₂ with silver nanoparticles effectively eliminated *E. faecalis* after only one day. Also Sadeghi [30] stated that nanosilver solution had antimicrobial properties against *Actinomyces viscosus* and *Streptococcus sanguinis*. Zhang [31] concluded that AgNPs alone or with Ca(OH)₂ had an obvious inhibitory

effect on the biofilm of *E. faecalis*. The result of this study is in contrast with the results of Mozayeni [32] who concluded that Nanosilver gel was not effective against *E. Faecalis*. This may be attributed to the different synthesis procedure of nanosilver particles and the fact that the added gel may have inhibited the release of nanoparticle ions.

But for *Candida*, there was no statistically significant decrease in number of CFUs between before and after treatment in AgNPs paste group. This result is in accordance with the results of Mozayeni [33] who concluded that antifungal efficacy of Ca(OH)₂ and 2% CHX gels was significantly higher than that of nanosilver gel in all tested samples. And with the results of Ballal [34] who concluded that Ca(OH)₂ showed higher efficacy at the first 24 hours against *Candida albicans*. The result of this study is in contrast with the results of Baker [14] who stated that 8 mg/cm² AgNPs had antimicrobial activity against *Candida albicans*. This may be due to using different concentration of AgNPs.

There was no statistically significant difference in types of microorganisms before and after treatment except for *Enterococcus* which appeared after treatment significantly in 0.03 mg/ml AgNPs group. This may be attributed to the detection of a bacterial species after treatment by molecular methods may result from the detection of DNA molecules still present in the specimen, which does not mean cell viability [35].

CONCLUSIONS

- The antimicrobial effect of 0.03mg/ml AgNPs paste was better than Ca(OH)₂ paste.
- AgNPs paste seems to be a valuable option in the battle against different types of bacteria.

REFERENCES

1. Kishen, A., 2010. Advanced therapeutic options for endodontic biofilms. Endod. Topics, 22: 99-123.
2. Davis, J.M., J. Maki and J.K. Babcall, 2007. An *in vitro* comparison of the antimicrobial effects of various endodontic medicaments on enterococcus faecalis. J. Endod., 33: 567-569.
3. Xinping, L.L. Shengli, Z. Miaotao, Z. Wenlong and L. Chuanghong, 2011. Evaluations of antibacterial activity and cytotoxicity on Ag nanoparticles. Rare Metal Materials and Engineering, 40: 0209-0214.
4. Turkevich, J., P.C. Stevenson and J. Hiller, 1951. Discuss. Faraday Soc., pp: 11-55.

5. Lee, P.C. and D. Meisel, 1982. Adsorption and Surface – Enhanced Raman of Dyes on Silver and Gold Sols. *Journal of Physical Chemistry*, 86: 3391-3395.
6. Søgaard, M., M. Nørgaard and H. Schønheyder, 2007. First notification of positive blood cultures: high accuracy of the Gram stain report. *J. Clin. Microbiol.*, 45: 1113-1117.
7. Putnam C.D., A.S. Arvai, Y. Bourne and J.A. Tainer, 2000. Active and inhibited human catalase structures: ligand and NADPH binding and catalytic mechanism. *J. Molecular Biol.*, 296: 295-309.
8. Primose, S.B., R.M. Twyman and R.W. Old, 2001. *Principles of Gene Manipulation*. Sixth edition.
9. Lee, P.Y., J. Costumbrado, C. Hsu and W.H. Kim, 2012. Agarose Gel Electrophoresis for the Separation of DNA Fragments. *Journal of Visualized Experiments*, 62: 1-6.
10. Nadkarni, M.A., F.E. Martin, N.A. Jacques and N. Hunter, 2002. Determination of bacterial load by real-time PCR using a broad-range (universal) probe and primers set. *Microbiology*, 148: 257-266.
11. Sinha, N., S. Patil, P.K. Dodwad, A.C. Patil and B. Singh, 2013. Evaluation of antimicrobial efficacy of calcium hydroxide paste, chlorhexidine gel and a combination of both as intra canal medicament: An in vivo comparative study. *J. Conserv. Dent.*, 16: 65-70.
12. Fidalgo, T.K., R. Barcelos, M.B. Portela, R.M. Soares, R. Gleiser and F.C. Silva Filho, 2010. Inhibitory activity of root canal irrigants against *Candida albicans*, *Enterococcus faecalis* and *Staphylococcus aureus*. *Braz. Oral Res.*, 24: 406-412.
13. Rai, M.K., S.D. Deshmukh, A.P. Ingle and A.K. Gade, 2012. Silver nanoparticles: the powerful nanoweapon against multidrug-resistant bacteria. *J. Appl. Microbiol.*, 112: 841-852.
14. Baker, C., A. Pradhan, L. Pakstis, D.J. Pochan and S.I. Shah, 2005. Synthesis and antibacterial properties of silver nanoparticles. *J. Nanosci Nanotechnol.*, 5: 244-249.
15. Panáček, A., L. Kvítek and R. Prucek, 2006. Silver colloid nanoparticles: synthesis, characterization and their antibacterial activity. *J. Phys. Chem. B.*, 110: 16248-16253.
16. Monteiro, D.R., L.F. Gorup, A.S. Takamiya and A.C. Ruvollo-Filho, E.R. De Camargo and D.B. Barbosa, 2009. The growing importance of materials that prevent microbial adhesion: antimicrobial effect of medical devices containing silver. *Int J. Antimicrob Agents*, 34: 103-110.
17. Ørstavik, D. and M. Haapasalo, 1990. Disinfection by endodontic irrigants and dressings of experimentally infected dentinal tubules. *Endod Dent Traumatol.*, 6: 142-149.
18. Heling, I., D. Steinberg, S. Kenig, Gavrilovich, M.N. Seal and M. Friedman, 1992. Efficacy of a sustained-release device containing chlorhexidine and Ca(OH)₂ in preventing secondary infection of dentinal tubules. *Int Endod J.*, 25: 20-24.
19. Siqueira, J.F. and M. De Uzeda, 1997. Intracanal medicaments: evaluation of the antibacterial effects of chlorhexidine, metronidazole and calcium hydroxide associated with three vehicles. *J. Endod.*, 23: 167-169.
20. Siqueira, J.F., T. Guimaraes-Pinto and I.N. Rocas, 2007. Effects of chemomechanical preparation with 2.5% sodium hypochlorite and intracanal medication with calcium hydroxide on cultivable bacteria in infected root canals. *J. Endod.*, 33: 800-805.
21. Rôças, I.N. and J.F. Siqueira, 2010. Identification of bacteria enduring endodontic treatment procedures by a combined reverse transcriptase-polymerase chain reaction and reverse-capture checkerboard approach. *J. Endod.*, 36: 45-52.
22. Schäfer, E. and K. Bössmann, 2005. Antimicrobial efficacy of chlorhexidine and two calcium hydroxide formulations against *Enterococcus faecalis*. *J. Endod.*, 31: 53-56.
23. Sharifian, M.R., N. Shokouhinejad, M. Aligholi, M. Emaneini, K. Arash and H. Assadian, 2008. *In vitro* comparison of the effectiveness of chlorhexidine and two calcium hydroxide formulation on *Enterococcus faecalis*. *Iranian End J.*, 3: 50-56.
24. Cwikla, S.J., M. Bélanger, S. Giguère, A. Progulske-Fox and F.J. Vertucci, 2005. Dentinal tubule disinfection using three calcium hydroxide formulations. *J Endod.*, 31: 50-52.
25. Bhardwaj, S.B., M. Mehta and K. Gauba, 2009. Nanotechnology: role in dental biofilms. *Indian J. Dent Res.*, 20: 511-513.
26. Behnen, M.J., L.A. West, F.R. Liewehr, T.B. Buxton, J.C. McPherson, 2001. Antimicrobial activity of several calcium hydroxide preparations in root canal dentin. *J. Endod.*, 27: 765-767.
27. Evans, M., J.K. Davies, G. Sundqvist and D. Figdor, 2002. Mechanisms involved in the resistance of *Enterococcus faecalis* to calcium hydroxide. *Int Endod J.*, 35: 221-228.

28. Stuart, C.H., S.A. Schwartz, T.J. Beeson and C.B. Owatz, 2006. *Enterococcus faecalis*: its role in root canal treatment failure and current concepts in retreatment. *J. Endod*, 32: 93-98.
29. Javidi, M., F. Afkhami, M. Zarei, K. Ghazvini and O. Rajabi, 2013. Efficacy of a combined nanoparticulate/calcium hydroxide root canal medication on elimination of *Enterococcus faecalis*. *Aust Endod J*, pp: 1-5.
30. Sadeghi, R., P. Owila, M.B. Rezvani and F. Taleg-hani, 2011. An *in-vitro* comparison between anti-microbial activity of nanosilver and chlorhexidine against *Streptococcus sanguinis* and *Actinomyces viscosus*. *J. Islam Dent Assoc*, 23: 225-231.
31. Zhang, F.H., M. Li, Z.J. Wei and B. Zhao, 2016. The effect of a combined nanoparticulate/calcium hydroxide medication on the biofilm of *Enterococcus faecalis* in starvation phase. *Shanghai Kou Qiang Yi Xue.*, 25: 11-15.
32. Mozayeni, M.A., A. Haeri, O. Dianat and A.R. Jafari, 2014. Antimicrobial Effects of Four Intra canal Medicaments on *Enterococcus faecalis*: An *in vitro* Study. *Iranian Endodontic Journal*, 9: 195-198.
33. Mozayeni, M.A., A. Hadian, P. Bakhshaei and O. Dianat, 2015. Comparison of antifungal activity of 2% chlorhexidine, calcium hydroxide and nanosilver gels against *Candida Albicans*. *Journal of Dentistry, Tehran University of Medical Sciences*, 12: 109-117.
34. Ballal, V., M. Kundabala, S. Acharya and M. Ballal, 2007. Antimicrobial action of calcium hydroxide, chlorhexidine and their combination on endodontic pathogens. *Aust Dent J.*, 52: 118-121.
35. De Souza, C.A.S., R.P. Teles, R. Souto, M.A.E. Chaves and A.P.V. Colombo, 2005. Endodontic therapy associated with calcium hydroxide as an intracanal dressing: microbiologic evaluation by the checkerboard DNA-DNA hybridization technique. *J. Endod*, 31: 79-83.