Global Veterinaria 25 (3): 118-124, 2023 ISSN 1992-6197 © IDOSI Publications, 2023 DOI: 10.5829/idosi.gv.2023.118.124

Morphological Identification of Veterinary Important Mosquitoes in the Selected Study Districts of Afar Region of Eastern Ethiopia

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Abstract: Mosquito survey study was conducted from March to August 2021 to identify veterinary important mosquitoes morphologically in Amibara, Gewane and Asayita districts situated in Afar region of Eastern Ethiopia. Trapping sites were selected based on the ecological significance for introduction and maintenance (mosquito breeding sites) and located within the range of geographical area of 9°11' 41.0712" N - 11°19' 22.4796" N, 40°6' 38.448"- 41°15' 54.0612" E and 358-736 m.a.s.l, altitudinal range. From the total 689 captured mosquitoes after examining their morphological features 369 mosquitoes were identified up to genera level: 204 (29.6%) *Aedes*, 140 (20.3%) *Culex* and 25 (3.6%) *Anopheles*. The remaining 320 mosquitoes identified further at species level and the existence of 11 mosquito species were confirmed in the study area in the following decreasing order of their count: *Culex pipines* and *Mansonia uniformis* which have equal count each 135 (19.6%), *C. antennatus* 28 (4.1%), *Anopheles arabiensis* 10 (1.5%), *Aedes aegypti, A. vexans, A. mcintoshi, C. theileri, An. plumbes,* which also have equal count each 2 (0.3%) *A. cumminsii* and *C. perexiguus* with the least count 1(0.1%). There was significant statistical difference in the catch of mosquitoes between districts and between species (P<0.05). Nowadays mosquito-borne diseases increased due to globalization and the finding of this study might contribute in the selection of effective and sustainable methods for controlling these vectors even though it lack behavioral information of these vectors/mosquitoes which need farther studies.

Key words: Vector-Borne · Mosquito · Aedes · Culex · Mansonia · Anopheles

INTRODUCTION

Mosquito is a small winged insect belongs to the order Diptera and the family Culicidae which is divided into 3 subfamilies (Anophelinae, Culicinae and Toxorhynchitinae) that consists of 3,490 currently recognized species grouped in 41 genera and among these the greatest species diversity occurs in tropical forest. From these sub families Anophelinae and the Culicinae are blood feeders but the third subfamily, the Toxorhynchitinae, do not feed on blood [1, 2].

Adult mosquitoes are small, slender insects of about 3 to 6 mm in length (0.15-0.4 inches). Males have feathery antennae; those with short antennal hairs are females. Mosquitoes use a complex set of cues, long range, midrange and close range to find a host [3]. They generally feed on plant nectar, but females of most species are parasitic and must consume blood to gain nutrients and

proteins needed to produce their eggs and depend on diversity of hosts, ranging from mammals and birds to reptiles, amphibians an even fish [4].

Female mosquito lay between 30 to 300 brown to blackish eggs at one time. *Anopheles* eggs are boat or oval shape and float on the water. *Culex* and some *Mansonia* eggs are also laid on the water, but are deposited in the form of rafts. Eggs of these three species cannot survive desiccation. *Aedes* lay their eggs in damp places just beyond the water line. Some *Aedes* prefer tree holes, clay pots and other containers. *Aedes* eggs can with stand weeks or years of desiccation and can survive cold weather. Hatching is triggered by alternate cycles of flooding and drought and do not hatch all the eggs at the same time. These mosquitoes tend to be timed release pests and never go away without good control measures. Most mosquito larvae must come to the surface to breath. At this time they are most vulnerable and this is the

reason that mosquito control work focuses on the larval stage. Anopheles larvae lie parallel to the water surface and breathe through the holes in their sides called spiracles. Mosquito's importance as pests is insignificant compared with their role as vectors particularly of human diseases such as malaria. Anopheline mosquitoes also transmit malaria parasites to other animals, for example Plasmodium knowlesi and P. cynomolgito to monkeys and P. bergheito to rodents. Culicine mosquitoes transmit malaria parasites such as P. gallinaceum to wild and domesticated birds. Anopheles gambiae is one of the best known, because of its predominant role in the transmission of the most dangerous Plasmodium falciparum [5]. Some species of Anopheles can also serve as the vectors for canine heart worm Dirofilaria immitis, the Filariidae wuchereria bancrofti and Brugia malayi and viruses [6].

Culicinae larvae hang from the surface at angle and breathe through a siphon tube. Many species spend 5-7 days in the larval stage. Mosquitoes can develop anywhere, where there is standing water. The range of habitats is wide. Fresh water, salt water, brackish water, ground pools, wells, cesspools, marshes, containers, tires, tree holes and aquatic vegetation are all areas where mosquitoes can develop [7].

Culex pipiens is primarily interested in birds and poultry, but also attacks humans and other mammals was a vector of St. Louis encephalitis in U.S. epidemics. It is also a vector of the canine heart worm and filariasis in humans. A related species of *Culex* species *Culex tasalis* is a principal vector of western equine encephalitis. Eastern equine encephalitis spread to man and horses by *Aedes* species [8].

Mechanical transmission of a number of poxviruses by biting arthropods is well documented and includes myxoma virus where *Ae. aegypti* has been identified as an important vector [9]. Mosquitoes have also been shown to mechanically transmit *Shope fibroma* virus and fowl pox virus [10, 11]. In the 1959 Kenyan outbreak of lamp skin disease, there were reports of high infestation of *Aedes natronius* and *Culex mirificus* [12].

In addition mosquito-borne arboviral diseases are becoming big health challenge of human and animal worldwide [13]. For example Rift Valley Fever (RVF) is adapted to wide range of vectors, predominantly mosquitoes [14]. The virus has been isolated from more than 53 mosquito species in 8 genera in regions where epizootics occurred [15]. It causes almost 100% mortality rates among young animals and high abortion rates among livestock [16]. The mosquito vectors transmitting RVF can be classified into two major groups, namely primary and secondary vectors [14]. Floodwater mosquitoes of genus Aedes have been considered the primary maintenance host and source of RVFV that initiate disease outbreaks [14, 17, 18]. The genera Culex, Anopheles, Eretmopodites and Mansonia constitute the secondary vectors which take over flooded grounds for breeding, contribute to the amplification of the virus due to their ubiquitous biting patterns, consequently resulting in outbreaks [14, 19]. Aedes ochraceus and Aedes mcintoshi is the most important maintenance vector of RVFV in East and Southern Africa. Similarly, Aedes vexans were found to be responsible for a large outbreak in West Africa and the likely maintenance vector in Saudi Arabia during the emergence of RVFV in 2000 [20]. Further more researchers in the field pointed out the importance of mosquitoes in the transmission of various diseases to animals and to human: for example the overall mosquitoes importance in the transmission of pathogens that causes of arboviral diseases (avian pox, bovine ephemeral fever, dengue fever, Rift Valley fever, West Nile fever), bacterial diseases (Anthrax, Tularemia), helminthic diseases (mosquito-borne filariasis), protozoans (Avian malaria, Human malaria) were indicated in the work of Azari-Hamidian S. and his colleagues[21]. Besides their role in the transmission of diseases Garros C. and his coworkers had also indicated that mosquito bites may cause stress and pain resulting in the reduction of livestock fitness [22].

Identification of mosquito species is one important step to control mosquito borne-diseases. It allows acquiring biological information such as breeding sites, biting and resting habits that differ among mosquito species [23]. Although they have similar morphology, mosquitoes have considerable differences in ecological, epidemiological significance and physiological features, including food preferences [24]. This helps to improve efforts to understand the spatial epidemiology of arboviruses and to predict how these could change in the future and could be used for early warning detection and implementation of control measures are essential.

In Ethiopia entomological study, regarding veterinary important mosquito has not been investigated. Therefore, this study with the following objective aimed to fill this gap. **Objective:** The objective of this study is to identify veterinary important mosquitoes morphologically in the selected parts of Afar regional state at genus and species level.

MATERIALS AND METHODS

Study Area: The study was undertaken in seven selected sites in Zone 3 at Amibara district in Serkamo and Shelako PAs at Gewane district in Galela dura and Entaye hadota PAs in Asayita Zone at Berga dura and Sahale PAs of Afar regional state where the sites were selected based on the ecological significance for introduction and maintenance (mosquito breeding sites). These sites located within the range of geographical area of 9° 11' 41.0712" N - 11°19' 22.4796" N, 40°6' 38.448"- 41°15' 54.0612" E and 358-736 m.a.s.l, altitudinal range.

Karma is the main rainy season in Afar which starts as heavy and stormy rains during early to mid-June, peaking during the month of July and continuing with lighter and lighter showers during the month of August and very limited and declining during early to mid-September. The current survey study was conducted from March to August 2021 in the area after the heavy rain which causes flooding in most parts of the study areas. The main perennial river flowing in the area is Awash but in the current study small rivers like Galela dura, Berga dubura and Sahale were used to position the mosquitoes trap.

People are predominantly involved in small-scale subsistence agriculture production and mainly on livestock husbandry and rear different animals, predominantly cattle, sheep, goat and camels. According to CSA, 2020/21[25] reports the region has 1,959,185 cattle, 4,476,485 sheep and 8,843,082 goats.

Study Population: The study animals for this particular study were Mosquitoes that were collected from the studied three districts.

Study Type: The study type was convenience sampling method to collect Mosquitoes in different selected trap sites of Amibara, Gewane and Asayita districts.

Mosquito Collection and Identification

Mosquitoes Were Collected by Using Two Different Types of Traps: CDC light traps and Modified Onderstepoort UV light traps. CDC light trap baited with sugar-yeast solution where by its fermentation reaction release carbon dioxide which is an important mosquito attractant.

The traps were set in proximity to potential mosquito breeding and feeding sites include indoor and outdoor (near water bodies near animal pen and on field of where there are human and livestock population). The trap placed at 18:00 PM and collected at 6:00 -7:00 AM in succeeding day. The next morning the traps were collected and using collection cup and put in deep freeze (-20°C) for 15 minutes to kill mosquitoes, then each specimen of mosquito inserted in to cryovial tubes which has silica gel to absorb moisture and cotton to protect from breaking. Sorting and identification into genus and species level were made by using dichotomous keys of Walter Reed Bio Systemic Unit (WRBU) Potter[26] and Edwards [27] under the aid of a stereo light microscope.

Data Management and Analysis: All data obtained from the study were entered and managed into Microsoft Excel worksheet and the analysis was made using Chi-square (x^2) test in SPSS version 20.

RESULTS

A total of 689 adult mosquitoes were collected from the three districts of Afar region of Eastern Ethiopia and the highest catch was from Amibara district 453(65.7%) followed by Gewane district 185 (26.9%) and the least catch was that of Asayita district 51 (7.4 %) (Table 1). After examining their morphological features 369 mosquitoes from the total 689 catch were identified up to the following genera level: 204 (29.6%) Aedes, 140 (20.3%) Culex and 25 (3.6%) Anopheles. The remaining 320 mosquitoes identified further at species level and the existence of 11 mosquito species were confirmed in the study area in the following decreasing order of their count: Culex pipines and Mansonia uniformis which have equal count each 135 (19.6%), C. antennatus 28 (4.1%), Anopheles arabiensis 10 (1.5%), Aedes aegypti, A. vexans, A. mcintoshi, C. theileri, An. plumbes, which also have equal count each 2 (0.3%) A. cumminsii and C. perexiguus with the least count 1(0.1%) (Table 1). There was significant statistical difference in the catch of mosquitoes between districts and between species too (P<0.05).

		District			
Mosquito species			Gewane	Asvita	Total
Aedes	Count	126	45	33	204
	% within Mosquito Species	61.80	22 10	16 20	100.00
	% within District	27.80	24 30	64 70	29.60
Culex	Count	107	15	18	140
	% within Mosquito Species	76.40	10.70	12.90	100.00
	% within District	23.60	8.10	35.30	20.30
Anopheles	Count	25	0	0	25
	% within Mosquito Species	100.00	0.00	0.00	100.00
	% within District	5.50	0.00	0.00	3.60
M.uniformis	Count	47	88	0	135
	% within Mosquito Species	34.80	65.20	0.00	100.00
	% within District	10.40	47.60	0.00	19.60
A.aegypti	Count	2	0	0	2
	% within Mosquito Species	100.00	0.00	0.00	100.00
	% within District	0.40	0.00	0.00	0.30
A.cumminsii	Count	1	0	0	1
	% within Mosquito Species	100.00	0.00	0.00	100.00
	% within District	0.20	0.00	0.00	0.10
A.vexans	Count	1	1	0	2
	% within Mosquito Species	50.00	50.00	0.00	100.00
	% within District	0.20	0.50	0.00	0.30
A. mcintoshi	Count	2	0	0	2
	% within Mosquito Species	100.00	0.00	0.00	100.00
	% within District	0.40	0.00	0.00	0.30
Cx. pipines	Count	115	20	0	135
	% within Mosquito Species	85.20	14.80	0.00	100.00
	% within District	25.40	10.80	0.00	19.60
Cx.antennatus	Count	17	11	0	28
	% within Mosquito Species	60.70	39.30	0.00	100.00
	% within District	3.80	5.90	0.00	4.10
Cx. theileri	Count	1	1	0	2
	% within Mosquito Species	50.00	50.00	0.00	100.00
	% within District	0.20	0.50	0.00	0.30
Cx.perexiguus	Count	1	0	0	1
	% within Mosquito Species	100.00	0.00	0.00	100.00
	% within District	0.20	0.00	0.00	0.10
An.arabiensis	Count	8	2	0	10
	% within Mosquito Species	80.00	20.00	0.00	100.00
	% within District	1.80	1.10	0.00	1.50
An.plumbes	Count	0	2	0	2
	% within Mosquito Species	0.00	100.00	0.00	100.00
	% within District	0.00	1.10	0.00	0.30
Total Mosquito	Count	453	185	51	689
	% within Mosquito Species	65.70	26.90	7.40	100.00
	% within District	100.00	100.00	100.00	100.00

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Table 1: Mosquito species identified in three districts at Afar region of Eastern Ethiopia

(X² =201.510 Df =26 and P-Value < 0.05)

DISCUSSION

In comparison with other studies like with the study of Kinyatta and his colleagues [28] who conducted the study in Kenya; neighboring country of Ethiopia and collected 1632 mosquitoes which they identified in to five genera (*Culex, Aedes, Anopheles, Mansonia* and *Ficalbia*), the collected mosquitoes number was found to be lesser and the *Ficalbia* mosquito genera was absent in the current study mosquitoes collection. This might be associated with the limitation of time and spatial coverage of the study areas. But in agreement with the study of Ridha M [29] from the collected mosquito species highest catch were recorded in the mosquito species of *Mansonia* and *Culex*. In spite of its very low abundance in the current mosquito species collections, *A. mcintoshi* have been indicated in Kenya being the predominant mosquito and the most known RVF vector [14, 18]. *Cx. pipines, Cx. theileri* and *Cx. perexiguus* are among the captured species in the current studies whereby their role to transmit many species of avian plasmodium were mentioned in the studies of researchers in different countries [30, 31]. *Cx. Pipines* in Africa and *Cx. perexiguus* in Asia and Europe have been also reported being the principal vectors of West Nile Virus (WNV) [32, 33]. The role of *Cx. Pipines* and *Cx. Perexiguus* in the transmission of RVFV among humans and animals have been mentioned also by Turell and his colleagues [34].

A. aegypti which is captured in this study is mentioned with other vectors being a vector of Lumpy skin disease (LSD) [35].

Flood water *Aedes* species; *A. mcintoshi*, sighted in the work of Rosemary and his co-workers [36] being the primary vector of RVF and in the neighboring country of Kenya the 2006/2007 outbreak of RVF and its transmission was also associated with this mosquito species.

The Epizootics of RVF in East and South Africa and its association with *A. mcintoshi*, *A. cummonsi* and *A.vexans* mosquito vectors have been explained in the study of Meegan and Bailey [37] these mosquito species were also caught in the current study although the virus isolation work was not conducted.

Among other mosquito genera the importance of Mansonia in the transmission of nematode have been indicated in the work of Anderson [38] and in the current study from the captured mosquito species *M.uniformis* has got the highest count like *Cx. Pipines*.

CONCLUSION AND RECOMMENDATION

The current study shows that the study area harbor 9 Culicinae species namely: *A. mcintoshi, A. cummonsi* and *A.vexans, A. aegypti, Cx. pipines, Cx. theileri, Cx. Perexiguus, C. antennatus, M.uniformis* and 2 Anophelinae species ; *Anopheles arabiensis* and *An. Plumbes* because of the diseases they transmit these mosquitoes are the greatest enemies of humans and their livestock.

Nowadays mosquito-borne diseases has also increased due to globalization and the finding of this study might contribute in the selection of the effective and sustainable methods for controlling these vectors. Since this study lack other important behavioral information of these vectors/mosquitoes ; such as their biting rate, host preference, dispersal and the effects of these behaviors on vector mortality and population growth; which are important in understanding the vectorborne disease (VBD) dynamics; which in turn important in the application of the control method. Therefore this study needs to be further strengthened by scaling up its scope to fill its gap.

REFERENCES

- Harbach, R., 2007. The Culicidae (Diptera), a review of taxonomy, classification and Phylogeny Zootatax, 638(1668): 591-638.
- 2. Durden, L. and G. Mullen, 2002. Medical and Veterinary Entomology San Diego, Calif: Academic Press.
- Gillett, J.D., 1972. The Mosquito: Its Life, Activities and Impact on Human Affairs. Doubleday, Newyork, pp: 358.
- Takken, W. and N. Verhulst, 2013. Host Preferences of Blood-Feeding Mosquitoes. Annual Review of Entomology, 433-453. http://doi.org/10.116. WHO (World Health Organization).1995, Vector control for Malaria and other Mosquito- Borne Diseases. WHO Technical Report Series No, 857. Geneva. 93.
- 5. Coetzee, M. and J. Goos, 2013. A new approach to malaria vector control. African Health (16): 18-19.
- WHO (World Health Organization), 1995. Vector control for Malaria and other Mosquito- Borne Diseases. WHO Technical Report Series No, 857. Geneva. 93.
- 7. Quaries, W., 2003. Mosquito Attractants and Traps, Common Sense Pest Control, 19(2)-7.
- Mulhern, T.D., ed, 1975. A training Manual for California Mosquito Control Association. Visalia, C.A.
- Fenner, F., M.F. Day and G.M. Woodroofe, 1952. The mechanism of the transmission of myxomatosis in the European rabbit (Oryctolagus cuniculus) by the mosquito Aedes aegypti. Aust. J. Exp. Biol. Med. Sci., 30: 139-52.
- 10. Kilham, L. and H.T. Dalmat, 1955. Host-virus mosquito relations to Shope fibrom as in cotton tail rabbits. Am. J. Hyg., 61: 45-54.
- Brody, A.L., 1936. The transmission of fowl pox. Cornell University Agric Experimental Station Mem, pp: 165.
- Burdin, M.L. and J. Prydie, 1959. Observations on the first outbreak of lumpy skin disease in Kenya. Bull Epizoot Dis Africa, 7: 21-6.
- Cooper, K.M., D.R. Bastola, R. Gandhi, D. Ghersi, S. Hinrichs, M. Morien and A. Fruhling, 2015. Forecasting the Spread of Mosquito-Borne Disease using Publicly Accessible Data: A Case Study in Chikungunya. In AMIA Annu Symp Proc., pp: 431-440.

- 14. Arum, S.O., C.W. Weldon, B. Orindi, T. Landmann, D.P. Tchouassi, H.D. Affognon and R. Sang, 2015. Distribution and diversity of the vectors of Rift Valley fever along the livestock movement routes in the northeastern and coastal regions of Kenya. Parasit & Vectors, 8(294): 1-9.
- Linthicum, K.J., S.C. Britch and A. Anyamba, 2016. Rift Valley Fever: An Emerging MosquitoBorne Disease. Annual Review of Entomology, 61: 395-415.
- Wilson, W.C., M. Romito, D.C. Jasperson, H. Weingartl, Y.S. Binepal, M.R. Maluleke and J.T. Paweska, 2014. Development of a Rift Valley fever real-time RT-PCR assay that can detect all three genome segments. Journal of Virological Methods, 193: 426-431.
- Himeidan, Y.E., E.J. Kweka, M.M. Mahgoub, E. Amin, E. Rayah and J.O. Ouma, 2014. Recent outbreaks of Rift Valley fever in East Africa and the Middle East. Frontiers in Public Health, 2(169).
- Sang, Rosemary, S. Arum, E. Chepkorir, G. Mosomtai, C. Tigoi, F. Sigei and M. Evander, 2017. Distribution and abundance of key vectors of Rift Valley fever and other arboviruses in two ecologically distinct counties in Kenya. PLOS Neglected Tropical Diseases, 11(2): e0005341.
- 19. FAD, P.R.e.P., 2013. Rift valley fever standard operating procedures: Overview of etiology and ecology. Riverdale, Maryland.
- 20. Pepin, M., M. Bouloy, B.H. Bird, A. Kemp and J. Pawesk, 2010. Rift Valley fever virus (Bunyaviridae: Phlebovirus): an update on pathogenesis, molecular epidemiology, vectors, diagnostics and prevention. Veterinary Research, BioMed Central, 41(61).
- Taira, K., T. Toma, M. Tamashiro and I. Miyagi, 2012. DNA barcoding for identification of mosquitoes (Diptera: Culicidae) from the Ryukyu Archipelago, Japan. Medical Entomology and Zoology, 63(4): 289-306.
- 22. Das, M., M. K.Das and P. Dutta, 2016. Genetic characterization and molecular phylogeny of Aedes albopictus (Skuse) species from Sonitpur district of Assam, India based on COI and ITS1 genes. Journal of Vector Borne Diseases, 53: 240-247.
- CSA, 2020/21. Federal democratic republic of Ethiopia central statistical agency: Agricultural sample survey 2020/21 [2013 E.C.] report on Livestock and Livestock Characteristics (private peasant holdings) (Vol. II).
- 24. Potter M.A., 2016. WRBU: keys to the medically important mosquito species: Identification key to the genera of adult mosquitoes for the world. Adapted from Harbach and Sandlant with the addition of Onirion and Verrallina.

- Edwards, F., 1941. Mosquitoes of the Ethiopian Region. III. Culicine adults and pupae. British Museum (Nat. Hist.), London. The Trustees of the British Museum.
- Azari-Hamidian, S., B. Norouzi and R.E. Harbach, 2019. A detailed review of the mosquitoes (Diptera: Culicidae) of Iran and their medical and veterinary importance. Acta. Trop., 194: 106-122.
- 27. Garros, C., J. Bouyer, W. Takken and R.C. Smallegange, 2018. Control of vector-borne diseases in the livestock industry: new opportunities and challenges. In Pests and vector-borne diseases in the livestock industry. Wageningen Academic Publishers, pp: 337-387.
- Kenyatta Kamau, L., F. Kamani, R. Kamani, M. Kagai, 2011. Determination of vectorial potential of Mansonia species in the transmission of Wuchereria Bancroft in tana-delta district, coast-Kenya. East African Medical Journal, 88: 347
- Ridha, M., N. Rahayu, B. Hairani, D. Perwitasari and H. Kusumaningtyas, 2020. Biodiversity of mosquitoes and Mansonia uniformis as a potential vector of Wuchereria bancrofti in Hulu Sungai Utara District, South Kalimantan, Indonesia, Veterinary World, 13(12): 2815-2821.
- Schoener, E., S.S. Uebleis, J. Butter, M. Nawratil, C. Cuk, E. Flechl, M. Kothmayer, A.G. Obwaller, T. Zechmeister, F. Rubel, K. Lebl, C. Zittra and H.P. Fuehrer, 2017. Avian Plasmodium in Eastern Austrian mosquitoes. Malar J., 16: 389.
- Ferraguti, M., J. Martinez-de la Puente, J. Munoz, D. Roiz, S. Ruiz, R. Soriguer and J. Figuerola, 2013. Avian Plasmodium in Culex and Ochlerotatus mosquitoes from southern Spain: effects of season and host-feeding source on parasite dynamics. PLOS One, 8: 6.
- 32. CDC, 2013. West Nile virus in the United States: Guidelines for surveillance, prevention and control.
- Hubálek, Z. and J. Halouzka, 1999. West Nile fever-a reemerging mosquito-borne viral disease in Europe. Emerg Infect. Dis., 5: 643-650.
- Turell, M.J., S.M. Presely and A.M. Gad, S.E.Cope, D.J. Doham, J.C. Morrill and R.R. Arthur, 1996. Vector Competence of Egyptian Mosquitoes for Rift Valley Fever virus. The American Journal of Tropical Medicine and Hygiene, 54(2): 136-9. pmid: 8619436.
- 35. Tuppurainen, E., T. Alexandrov and D. Beltrán-Alcrudo, 2017. Lumpy skin disease field manual - A manual for veterinarians. FAO Animal Production and Health Manual No. 20. Rome. Food and Agriculture Organization of the United Nations (FAO), pp: 60.

- 36. Rosemary Sang, Samwel Arum, Edith Chepkorir, Gladys Mosomtai, Caroline Tigoi, Faith Sigei and Olivia Wesula Lwande, Tobias Landmann, Hippolyte Affognon, Clas Ahlm, Magnus Evander, 20'17. Distribution and abundance of key vectors of Rift Valley fever and other arboviruses in two ecologically distinct counties in Kenya. https://journals.plos.org/plosntds/article?id=10.137 1/journal.pntd.0005341
- Meegan, J.M. and C.H. Bailey, 1988. Rift Valley fever. In: Monath TP, editor. The arboviruses: epidemiology and ecology. Boca Raton (FL): CRC Press, pp: 51-76.
- Anderson, R.C., 2000. Nematod Parasites of Vertebrates. Their Development and Transmission (2nd Edn.). CABI Publishing, Wallingford.