ISSN 2078-466X

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DOI: 10.5829/idosi.gjbb.2014.9.2.8518

In Silico Profiling of Regulatory Micrornatargets in GJB3 Gene

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Abstract: MicroRNAs are novel class of small non-coding RNAs that target and silence various genes across diverse signalling pathways comprising key physiological networks. Despite an important gene regulator, computational and experimental approaches are still infancy and an overall understanding of the importance of these regulatory transcripts is still far of satisfactory. Currently computer based prediction remains the only source for rapid identification of putative microRNA (miRNA) target. An increasing number of experimentally validated miRNAs targets are now available; utilizing this additional information in search of further targets may help to improve the specificity of bioinformatics based methods for predicting gene target site. An effort has been made in this pilot approach to investigate regulatory miRNA sequence and their location in the candidate gene of hearing impairment (*GJB3* gene) using online bioinformatics tool called miRDB. Using this online database, we identified eleven specific miRNAs (i.e: hsa-miR-466, hsa-miR-3613-3p, hsa-miR-16-1-3p, hsa-miR-144-3p, hsa-miR-561-5p, hsa-miR-326, hsa-miR-2110, hsa-mir-4510, hsa-mir-4419a, hsa-miR-3605-5p and hsa-miR-4738-3p). These targets different regions in the *GJB3* gene. Multiple sequence alignment to investigate whether similarities exists among mature sequences of these selected miRNAs were also performed. This data set will provide concrete bases and will help in experimental validation of these miRNAs.

Key words: Bioinformatics Tools • GJB3 • Microrna • Multiple Alignment • Mirdb

INTRODUCTION

MicroRNAs (miRNAs) are small, endogenous, noncoding RNAs, usually between 18 and 25 nucleotides in length, involved in the regulation of cellular and developmental processes through post-transcriptional gene repression [1-3]. These are expressed either as single transcription unit or as polycistronic transcripts from miRNA clusters, encoded within intronic or intergenic regions of the genome [4, 5]. Primary or pri-miRNAs are formed by polymerase II that drives transcription of miRNAs as inverted repeats embedded in long primary transcripts, which spontaneously fold to form imperfect long hairpins [6-8]. It is then processed into shorter hairpin precursor miRNAs, or pre-miRNAs, in the nucleus bvRNase III enzyme complex of DROSHA (RNASEN)/DGCR8 [9]. Pre miRNAs are transferred into cytoplasm by a trans-nuclear membrane protein called exportin 5 (XPO5) [7, 8]. In cytoplasm, the RNase III enzyme DICER1 cut the pre-miRNA to form

double-stranded miRNA duplex comprising a mature miRNA (guide strand) and a partially complementary passenger, or "star" (*), strand [10,11]. The fully matured miRNA associates with argonaute proteins in the RNA-induced silencing complex (RISC) to direct translational repression by binding to the regions of complementarily in 3'untranslated regions of target mRNAs [12,13].

RISC loading has been shown to be largely asymmetric, with only a single strand of miRNA duplex being incorporated to direct gene silencing [14]. However, some miRNA duplexes encode mature miRNAs on both strands and recent evidence suggests that strand based in miRNA expression may be influenced by tissue-specific processing factors [15]. The altered expression may be due to a variety of mechanisms including transcriptional regulation, amplification, deletion, mutation and epigenetic silencing [16]. Although miRNA signatures were established in tumor cells [17,18], recent studies revealed that the potential capabilities of miRNAs as blood-based biomarkers for cancer and other diseases[19].

Approximately 98% of RNAs in mammalian cells do not code for proteins. The epigenetic factors are associated with hearing loss [20], elevating the possibility that noncoding RNAs, such as miRNAs, might also involve in inner ear development and hearing loss.

In Silico miRNA target identification: Both computational and experimental approaches show that thousands of human genes are regulated by miRNAs [21, 22]. The functional characterization of miRNAs has become one of the most interesting research domains because of their critical roles in gene expression regulation. However, accurate target prediction is one of the major issue faced bymiRNAsbasedresearch is the lack of computational tools. Various types of bioinformatics tools are developed to give insight into the molecular functionalities of microRNA gene regulatory network. One strategy for target prediction is to use machine learning approach but has not been applied to miRNA target prediction to a greater extant. An increasing number of experimentally validated microRNAs targets are now available that utilize this additional information in search of further targets. Which may help to improve the specificity of in silico based methods for target site prediction.

Structure and function of GJB3: The GJB3 gene encoding the gap junction protein connexin 31 (Cx31) was initially mapped to chromosome 1p35- p33 and heterozygous mutations were shown to cause ADNSHL (Autosomal Dominant Non-Syndromic Hearing Loss) [23,24]. GJB3 mutations have been reported to cause ARNSHLand skin disorder callederythro keratodermiavariabilis (MIM 133200). Biallelic GJB3 mutations causing ARNSHL have been reported once in two families in which patients were compound heterozygote for two different GJB3 mutations [25]. Digenic inheritance of non-syndromic deafness caused by mutations in GJB2 and GJB3 has recently been reported. Two different missense mutations (p.N166S and p.A194T) of GJB3 were found in compound heterozygosity with the c.235delC and c.299delAT mutations of GJB2 in three simplex families from China [26].

One gap junction consists of a cluster of closely packed pairs of trans-membrane channels, the connexins, through which materials of low molecular weight diffuse from one cell to a neighbouring cell. Involvement in disease, *GJB3* is a cause of erythrokeratodermiavariabilis (EKV) [MIM:133200]. EKV is a genodermatosis characterized by the appearance of two independent skin lesions: transient figurate erythematous patches and hyperkeratosis that is usually localized but occasionally

occurs in its generalized form. Clinical presentation varies significantly within a family and from one family to another. Palmoplantarkeratoderma is present in around 50% of all cases. Defects in *GJB3* are the cause of deafness autosomal dominant type 2B (DFNA2B) [MIM:612644]. DFNA2 is a form of sensorineural hearing loss. Sensorineural deafness results from damage to the neural receptors of the inner ear, the nerve pathways to the brain, or the area of the brain that receives sound information.

MATERIALS AND METHODS

List of genes associated with hearing loss identified in human were collected from literature and publicly available databases. All relevant publications were identified after searching PubMed (http://www.ncbi. nlm. nih. gov/pubmed) with key phrases, such as gene, genetics, hearing, candidate's genes for hearing, physiology of hearing impairments etc. Among the list of candidate genes, *GJB3* were selected for this study.

We used miRDB for potentialmiRNA target identification in *GJB3*. To briefly explain, miRDB is an online computational tool for miRNA target prediction and functional annotations [5]. All the targets were predicted by a bioinformatics tool MirTarget2, which was developed by analysing thousands of genes impacted by miRNAs with an SVM learning machine. Common features associated with miRNA target binding have been identified and used to predict miRNA targets. miRDB hosts predicted miRNA targets in five species: human, mouse, rat, dog and chicken.

RESULTS AND DISCUSSION

A large number of microRNAs have been identified across a variety of different species. Being as gene regulators, identification of microRNA targets has become an essential step toward understanding these regulatory mechanisms. *In silico* analysis of prediction presently remains the only source for rapid identification of a putative microRNA target because it helps in efficiently allocating experimental resources. *In silico* microRNA target prediction programs are based on specific parameters that can give slightly different results for the same target input. Such limitations can be partially compensated by predicting targets using more than one program. These approaches have been quite successful for a few top ranked results in different diseases' models. In the present study, an attempt was made to predict

target sites in the sequences of selected genes through advance bioinformatics approaches such as miRDB (online tool) were used.

Using this database, 11 potential miRNAs in the sequence of *GJB3* gene were identified, given in Fig. 2. This computer based approach for identification of target site in selected genes will provide base for experimental validation of these novel microRNAs in *GJB3*. If validated experimentally, these miRNAs may be used as novel biomarker for hearing impairment in human.

Salih et al. [27] carried out study on mutation in GJB3 and GJB4 genes involved in Deafness in two Sudanese families using next generation sequencing technique. Similarly a study was carried out in Kenyan and Sudanese children suffering from NSARD (non-syndromic autosomal recessive deafness) due to variants of GJB2 gene. As compare to other areas and other ethnic groups, deafness-associated variants of the coding region of GJB3 were rare in Kenya and Sudan, which clearly demonstrate a causative role of other genetic or epigenetic factors [28]. As compare to these studies our results also suggest that GJB3 may be possibly responsible for hearing impairment.

GJB2 deafness-associated mutations are ethnicity-specific [30] and occur witha frequency of 30 - 50% in most Caucasian populations and 17%in Ghanaians, but they are remarkably rare among Kenyan and Sudanese populations [31]. Similarly the study of Lopez-Bigas *et al.*

[29] pointed out the strong association of E204A with deafness. From these studies [27-29] it can be hypothesized that E204A mutation has pathogenic effect on Connexins' activity which lead to hearing loss.

Trotta et al. [32] analysed the genotypic composition of GJB2 gene and did not witnessed any significant difference between deaf and control group suggesting that GJB2 gene or its genotypic combination with any other gene is not attributing to deafness in Northern Cameroon. Their results were different than those studies conducted at Kenya and Sudan. This also indicate that there are many factors contributing to deafness which vary within different populations such as presence of infections, malnutrition, poverty and poor access to health care. These factors may play a more predominant role than hereditary factors [33].

Details names of the candidate miRNAs, their sequence order, size, seed location, target score, target gene symbol and protein size of the target gene is given for each microRNA (Fig. 3 to Fig. 13).

Multiple alignment of selected miRNA sequences targeting *GJB3*: After identification of list of selected miRNAs (i.e. hsa-miR-466, hsa-miR-3613-3p, hsa-miR-16-1-3p, hsa-miR-144-3p, hsa-miR-561-5p, hsa-miR-326, hsa-miR-2110, hsa-mir-4510, hsa-mir-4419a, hsa-miR-3605-5p and hsa-miR-4738-3p), it was investigated, if there is any similarities among the sequences of these selected miRNAs. Multiple alignment of the mature sequences of

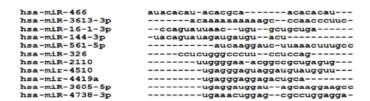


Fig. 1: Represents multiple sequence alignment of 11 selected miRNA genes.

Target Detail	Target Rank	Target Score	miRNA Name	Gene Symbol	Gene Description		
Details			hsa-miR-4510		gap junction protein, beta 3, 31kDa		
Details	2	58	hsa-miR-4419a	GJB3	gap junction protein, beta 3, 31kDa		
Details	3	57	hsa-miR-3605-5p	GJB3	gap junction protein, beta 3, 31kDa		
Details	4	56	hsa-miR-16-1-3p	<u>GJ83</u>	gap junction protein, beta 3, 31kDa		
Details	5	55	hsa-miR-144-3p	GJB3	gap junction protein, beta 3, 31kDa		
Details	6	55	hsa-miR-3613-3p	GJB3	gap junction protein, beta 3, 31kDa		
Details	7	52	hsa-miR-2110	GJB3	gap junction protein, beta 3, 31kDa		
Details	8	51	hsa-miR-561-5p	GJB3	gap junction protein, beta 3, 31kDa		
Details	9	51	hsa-miR-4738-3p	GJB3	gap junction protein, beta 3, 31kDa		
<u>Details</u>	10	51	hsa-miR-466	GJB3	gap junction protein, beta 3, 31kDa		
Details	11	51	hsa-miR-326	GJB3	gap junction protein, beta 3, 31kDa		

Fig. 2: List of 11 miRNA predicated by online tool miRDB in the sequence of *GJB3* gene.

MicroRNA and Target Gene Description: miRNA Name hea-miR-4510 miRNA Sequence UGAGGGAGUAGGAUGUAUGGUU **Target Score Seed Location** 466, 630 65 **NCBI Gene ID** GenBank Accession NM 001005752 2707 3' UTR Length Gene Symbol GJB3 806 Gene Description gap junction protein, beta 3, 31kDa 3' UTR Sequence 1 CCACAGGGCA GGGGTGGGGC AACATGCGGG CTGCCAATGG GACATGCAGG GCGGTGTGGC AGGTGGAGAG GTCCTACAGG GGCTGAGTGA CCCCACTCTG AGTTCACTAA GTTATGCAAC 121 TITCGTTTTG GCAGATATTT TITGACACTG GGAACTGGGC TGTCTAGCCG GGTATAGGTA 181 ACCCACAGGC CCAGTGCCAG CCCTCAAAGG ACATAGACTT TGAAACAAGC GAATTAACTA 241 TCTACGCTGC CTGCAAGGGG CCACTTAGGG CACTGCTAGC AGGGCTTCAA CCAGGAAGGG 301 ATCAACCCAG GAAGGGATGA TCAGGAGAGG CTTCCCTGAG GACATAATGT GTAAGAGAGG 361 TGAGAAGTGC TCCCAAGCAG ACACAACAGC AGCACAGAGG TCTGGAGGCC ACACAAAAAG 421 TGATGCTCGC CCTGGGCTAG CCTCAGCAGA CCTAAGGCAT CTCTACTCCC TCCAGAGGAG 481 CCGCCCAGAT TCCTGCAGTG GAGAGGAGGT CTTCCAGCAG CAGCAGGTCT GGAGGGCTGA 541 GAATGAACCT GACTAGAGGT TCTGGAGATA CCCAGAGGTC CCCCAGGTCA TCACTTGGCT 601 CAGTGGAAGC CCTCTTTCCC CAAATCCTAC TCCCTCAGCC TCAGGCAGTG GTGCTCCCAT 661 CTTCCTCCCC ACAACTGTGC TCAGGCTGGT GCCAGGCCTTT CAGACCCTGC TCCCAGGGAC 721 TTGGGTGGAT GCGCTGATAG AACATCCTCA AGACAGTTTC CTTGAAATCA ATAAATACTG 781 TGTTTTATAC AAAAAAAAAA AAAAAA

Fig. 3: Illustrate the miRNA sequence (has-miR-4510), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

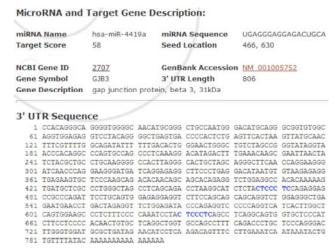


Fig. 4: Illustrate the miRNA sequence (has-miR-4419a), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

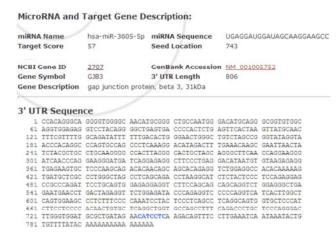


Fig. 5: Illustrate the miRNA sequence (has-miR-3605-5p), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

MicroRNA and Target Gene Description: hsa-miR-16-1-3p miRNA Sequence miRNA Name CCAGUAUUAACUGUGCUGCUGA **Previous Name** hsa-miR-16-1* 56 Seed Location Target Score NCBI Gene ID 2707 GenBank Accession NM 001005752 Gene Symbol 3' UTR Length Gene Description gap junction protein, beta 3, 31kDa 3' UTR Sequence 1 CCACAGGGCA GGGGTGGGGC RACATGCGGG CTGCCAATGG GACATGCAGG GCGGTGTGGC 61 AGGTGGAGAG GTCCTACAGG GGCTGAGTGA CCCCACTCTG AGTTCACTAA GTTATGCAAC 121 TTTGGTTTTG GCAGATATTT TTTGACACTG GGAACTGGGC TGTCTAGCGC GGTATAGGTA 121 ACCCACAGGC CCAGTGCCAG CCCTCAAAGG ACATAGACTT TGAAACAAGC GAATTAACTA 241 TCTACGCTGC CTGCAAAGGG CCACTTAGGG CACTGCAGG AGGGCTTCAA CCAGGAAGGG 301 ATCAACCCAG GAAGGGATGA TCAGGAGAGG 361 IGAGGAGTEC TOCCAAGGAG ACACAACAGC AGCACAGAGG TOTGGAGGCC ACACAAAAAA 421 IGATGCTCGC CCTGGGCTAG CCTCAGCAGA CCTAAGGCAT CTCTACTCCC TCCAGAGGAG 481 CCGCCCAGAT TCCTGCAGTG GAGAGGAGGT CTTCCAGCAG CAGCAGGTCT GGAGGGCTGA 541 GAATGAACCT GACTAGAGGT TOTGGAGATA CCCAGAGGTC CCCCAGGTCA TCACTTGGCT 601 CAGTGGAAGC CCTCTTTCCC CAAATCCTAC TCCCTCAGCC TCAGGCAGTG GTGCTCCCAT 661 CTTCCTCCCC ACAACTGTGC TCAGGCTGGT GCCAGCCTTT CAGACCCTGC TCCCAGGGAC 721 TIGGGIGGAT GCGCIGATAG AACATCCICA AGACAGITIC CIIGAAATCA ATAAATACTG 781 IGITITATAC AAAAAAAAA AAAAAA

Fig. 6: Illustrate the miRNA sequence (has-miR-16-1-3p), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

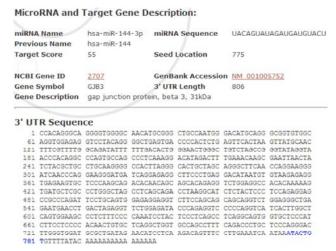


Fig. 7: Illustrate themiRNA sequence (has-miR-144-3p), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

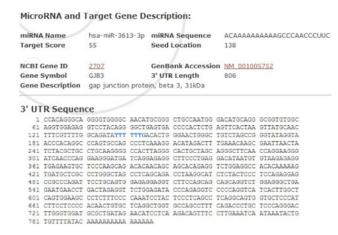


Fig. 8: Illustrate the miRNA sequence (has-miR-3613-3p), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

MicroRNA and Target Gene Description: hsa-miR-2110 miRNA Sequence UUGGGGAAACGGCCGCUGAGUG Target Score 52 **Seed Location** 616 NCBI Gene ID GenBank Accession NM 001005752 Gene Symbol GJB3 3' UTR Length 806 Gene Description gap junction protein, beta 3, 31kDa 3' UTR Sequence STEGGGC AACATGCGGG CTGCCAATGG GACATGCAGG GCG 61 AGGTGGAGAG GTCCTACAGG GGCTGAGTGA CCCCACTCTG AGTTCACTAA GTTATGCAAC 121 TTTCGTTTTG GCAGATATTT TTTGACACTG GGAACTGGGC TGTCTAGCCG GGTATAGGTA 121 TITGGTTITG GCAGATATTI TITGACACTG GGAACTGGGC TGTCTAGCCG GGTATAGGTA 181 ACCACCAGGC CCAGTGCCAG CCCTCAAAGG ACATAGACTT TGAAACAAGC GAALTAACTA 241 TCTACGCTGC CTGCAAGGGG CCACTAAGGG CATGCTAGC AGGGCTTCAA CCAGGAAGGG 301 ATCAACCCCAG GAAGGGA TCACGAAGGG CTTCCCTGAG GACATAATGT GTAAGGAGGG 361 TGAGAAGTGC TCCCAAGCAG ACACACAGAG CACCAGAGGT CTGTGGAGGCC ACACAAAAAG 421 TGATGCTGCC CCTGGGCTAG CCTCAGCAGA CCTAAGGGAT CTCTACTCCC TCCAGGGGG 481 CCGCCCAGAT TCCTGCAGTG GAGAGGAGGT CTCCAGCAGGTC GGAGGGCTG 481 CGACCAGAT CGACCAGTG TCGAGGAGT CCCCAGGTCA TCACTTGGCT 601 CASTGGAAGC CCTCTTTCC CAAATCCTAC TCCCTCAGCC TCAGGCCTGT GTGCTCCCAT 661 CTTCCTCCCC ACAACTGTGC TCAGGCTGGT GCAGCCTGT CCCCAGGGCC 721 TTGGGTGGAT GCCCTGATAG AACATCCTCA AGACAGTTC CTTGAAATCA ATAAATACTG 781 TGGTTTATAC AAAAAAAAAA AAAAAAA 781 TGTTTTATAC AAAAAAAAA AAAAAA

Fig. 9: Illustrate the miRNA sequence (has-miR-2110), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

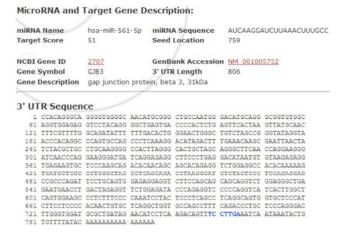


Fig. 10: Illustrate the miRNA sequence (has-miR-561-5p), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

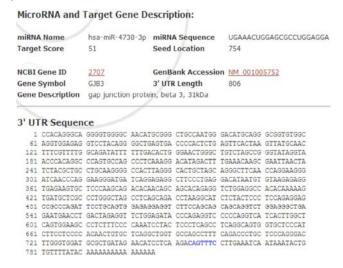


Fig. 11: Illustrate the miRNA sequence (has-miR-4738-3p), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

Micro	RNA and	Target Ge	ne Descrip	tion:			
miRNA Name Target Score		hsa-miR-466	miRNA Sequence Seed Location		AUACACAL	AUACACAUACACGCAACACACA	
		51			347	347	
NCBI Gene ID Gene Symbol Gene Description		2707	GenB	GenBank Accession		NM 001005752 806	
		GJB3	3' UTR Length		806		
		gap junction protein, beta 3, 31kDa					
3' U1	R Sequen	ce					
1	CCACAGGGCA	GGGGTGGGGC	AACATGCGGG	CTGCCAATGG	GACATGCAGG	GCGGTGTGGC	
61	AGGTGGAGAG	GTCCTACAGG	GGCTGAGTGA	CCCCACTCTG	AGTTCACTAA	GTTATGCAAC	
121	TTTCGTTTTG	GCAGATATTT	TTTGACACTG	GGAACTGGGC	TGTCTAGCCG	GGTATAGGTA	
181	ACCCACAGGC	CCAGTGCCAG	CCCTCAAAGG	ACATAGACTT	TGAAACAAGC	GAATTAACTA	
241	TCTACGCTGC	CTGCAAGGGG	CCACTTAGGG	CACTGCTAGC	AGGGCTTCAA	CCAGGAAGGG	
301	ATCAACCCAG	GAAGGGATGA	TCAGGAGAGG	CTTCCCTGAG	GACATAATOT	GTAAGAGAGG	
361	TGAGAAGTGC	TCCCAAGCAG	ACACAACAGC	AGCACAGAGG	TCTGGAGGCC	ACACAAAAAG	
421	TGATGCTCGC	CCTGGGCTAG	CCTCAGCAGA	CCTAAGGCAT	CTCTACTCCC	TCCAGAGGAG	
481	CCGCCCAGAT	TCCTGCAGTG	GAGAGGAGGT	CTTCCAGCAG	CAGCAGGTCT	GGAGGGCTGA	
541	GAATGAACCT	GACTAGAGGT	TCTGGAGATA	CCCAGAGGTC	CCCCAGGTCA	TCACTTGGCT	
601	CAGTGGAAGC	CCTCTTTCCC	CAAATCCTAC	TCCCTCAGCC	TCAGGCAGTG	GTGCTCCCAT	
661	CTTCCTCCCC	ACAACTGTGC	TCAGGCTGGT	GCCAGCCTTT	CAGACCCTGC	TCCCAGGGAC	
721	TTGGGTGGAT	GCGCTGATAG	AACATCCTCA	AGACAGTTTC	CTTGAAATCA	ATAAATACTG	

Fig. 12: Illustrate themiRNA sequence (has-miR-466), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

miRNA Name		hsa-miR-326	miRNA Sequence		CCUCUGGGCCCUUCCUCCA		
Target Score		51	Seed Location		571		
NCBI	Gene ID	2707	GenB	ank Accessio	on NM 00100	5752	
Gene Symbol		GJB3 3' UTF		R Length	806		
Gene Description		gap junction protein, beta 3, 31kDa					
	- 6	ATOMIC .					
3' UT	R Sequen	ce					
1	CCACAGGGCA	GGGGTGGGGC	AACATGCGGG	CTGCCAATGG	GACATGCAGG	GCGGTGTGGC	
61	AGGTGGAGAG	GTCCTACAGG	GGCTGAGTGA	CCCCACTCTG	AGTTCACTAA	GTTATGCAAC	
121	TTTCGTTTTG	GCAGATATTT	TTTGACACTG	GGAACTGGGC	TGTCTAGCCG	GGTATAGGTA	
181	ACCCACAGGC	CCAGTGCCAG	CCCTCAAAGG	ACATAGACTT	TGAAACAAGC	GAATTAACTA	
241	TCTACGCTGC	CTGCAAGGGG	CCACTTAGGG	CACTGCTAGC	AGGGCTTCAA	CCAGGAAGGG	
301	ATCAACCCAG	GAAGGGATGA	TCAGGAGAGG	CTTCCCTGAG	GACATAATGT	GTAAGAGAGG	
361	TGAGAAGTGC	TCCCAAGCAG	ACACAACAGC	AGCACAGAGG	TCTGGAGGCC	ACACAAAAAG	
421	TGATGCTCGC	CCTGGGCTAG	CCTCAGCAGA	CCTAAGGCAT	CTCTACTCCC	TCCAGAGGAG	
481	CCGCCCAGAT	TCCTGCAGTG	GAGAGGAGGT	CTTCCAGCAG	CAGCAGGTCT	GGAGGGCTGA	
541	GAATGAACCT	GACTAGAGGT	TCTGGAGATA	CCCAGAGGTC	CCCCAGGTCA	TCACTTGGCT	
601	CAGTGGAAGC	CCTCTTTCCC	CARATCCTAC	TCCCTCAGCC	TCAGGCAGTG	GIGCICCCAT	
661	CTTCCTCCCC	ACAACTGTGC	TCAGGCTGGT	GCCAGCCTTT	CAGACCCTGC	TCCCAGGGAC	
721	TTGGGTGGAT	GCGCTGATAG	AACATCCTCA	AGACAGTTIC	CTTGAAATCA	ATAAATACTG	
-	TGTTTTATAC	AAAAAAAAA	AAAAAA				

Fig. 13: Illustrate themiRNA sequence (has-miR-326), its seed sequence and it location in the sequence of target gene *GJB3* predicated by online tool miRDB.

these miRNAs were performed using Clustal Omega, which is maintained by EMBL-EBI (European Molecular Biology Laboratory- The European Bioinformatics Institute, i.e: part of the EMBL).

In our study the multiple alignment results showed that there is high level of sequences' variation among these selected miRNA except hsa-mir-4510, hsa-mir-4419a and hsa-miR-3605-5p where there is some sequence similarities, as shown in Fig. 1. As compare to the study of Mishra and Chandrasekharan [34], they used CLUSTAL-W alignment tool for alignment of all miRNA precursors in miRBase for selected species of hexapoda. They analysed conservation among *Apismellifera*, *Bombyxmori* and *Anopheles gambiae*. They concluded

that about 82% mature sequences fall under conserved region while 13% are outside the conserved region by 1-2 nucleotides.

CONCLUSION

It was concluded that there are 11 target sites (seed sequences) for miRNA on *GJB3* gene. These are non-identical. Every gene containsmiRNA target sites that can be arranged using different bio informatics tools. This might help molecular biologists to diagnose genetic disorders such asunko gene expression rate, siRNA b ased drug designing and easy and short analysis of a genetically blemishedpedigree for a specific

loci. In future amiRNA based DNA micro array technology can be developed for mutation's identification.

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