

Prevalence of Fasciolosis in Cattle Slaughtered at Dambi Dollo Town Municipal Abattoir; Kellem Wollega Zone, Ethiopia

¹Tesfaye Itefa, Yoseph Alemu, Ebisa Regasa, Amanuel Alemu and ²Milkiyas Kena

¹Kellem Wollega Zone Agriculture Office, Expert of Livestock Health, Ethiopia

²Gawo Kebe District Agriculture Office, Livestock Health Experts, Ethiopia

Abstract: The study was carried out from January to December 2021 with the main objectives of to determine the prevalence of fasciolosis infections in cattle, slaughtered at DambiDollo Municipal Abattoir. The slaughtered animals were daily inspected for liver fasciolosis throughout the year of 2021. Macroscopic fasciolosis was detected from a total of 4424 basing on animals species, sex, season and *Fasciola* species. In addition to this, fecal samples from 100 female cattle were collected for microscopic examination. The total prevalence rate of *Fasciola* species infection occurs in the study area were about 1364/4424 (30.83%) from the total cattle slaughtered carcasses. On sex based case, prevalence of fascioliasis was 8/24 (33.33%) and 1356/4400 (30.82%) for females and males cattle carcasses, respectively. The study revealed that the significance of season in finding that highest fasciolosis infection was recorded during winter and autumn. It constitutes a major cause of economic losses at study area and threat public health.

Key words: *Fasciola gigantica* • *Fasciola hepatica* • Liver Fluke • Slaughterhouse • Snails

INTRODUCTION

Slaughterhouses provide an excellent meat inspection place, where many zoonotic diseases observed but meat poor handling in or out the abattoir can leading to both economic losses and a lot of public health hazardous [1, 2]. Fasciolosis considered the top of all the domestic ruminants' parasitic zoonotic worldwide infection that is endemic in a tropical area and KellemWollega, DambiDollo [3-5].

The genus *Fasciola* "liver fluke" is belonging to trematodehelminthes which containing two main species; *Fasciola gigantica* and *Fasciola hepatica* is very common observed in the liver of cattle and other ruminants [6-8]. Fasciolosis reduces animal productivity, weight gain and the production of meat and milk. In addition, it causes moderate icterus, metabolic disorders and secondary infections due to decrease immunity by chronic fasciolosis and liver condemnation during postmortem inspection in slaughterhouses while the acute fasciolosis may lead to mortalities [9-11]. Human fasciolosis infection occurs accidentally after ingestion of eggs/larvae while ruminant ingestion of forage containing metacercarial cyst [12].

Ingested parasite lives in hepatic parenchyma or in bile duct, which causing liver hemorrhagic black tunnels [13]. Diagnosis is depending on the history of snail habitats or fasciolosis on the farm, symptoms, postmortem examinations, feces and blood examination for *Fasciola* eggs [14].

There is no enough information on the ruminants' fasciolosis in the study area, DambiDollo town. Therefore, this study was designed with the aim of determining the prevalence of fasciolosis infections in cattle slaughtered in DambiDollo town slaughterhouse.

MATERIALS AND METHODS

The Study Area: A cross-sectional study was conducted in DambiDollo Municipal abattoir to detect the prevalence of the fasciolosis (liver flukes) from the slaughtered cattle. DambiDollo town is the capital city of KellemWollega Zone Administration. It is a part of the Oromia region, which is located to the west of the Addis Ababa/Finfinne between 8°32'N latitude and 34°48'E longitude with an elevation between 1701 and 1827 meters above sea level. DambiDollo town is located 652 km to the west of Addis Ababa/Finfinne. DambiDollo Municipal

abattoir slaughtered about 4424 cattle animals during 2021. According to the Ethiopian legislations of meat inspection, slaughtering of female cattle never been allowed before all teeth are changed (over 5 years) while they approved for slaughtering after about 2 years.

Samples Collection: A total of 4424 (4400 bulls and 24 cows) local breed cattle slaughtered at DambiDollo Municipal abattoir were inspected for the presence of liver fluke/fasciolosis allover 2021 which was efficiently inspected by naked eye and palpation for the presence of gross lesion and the worms, then further examinations done at Laboratory level. All data samples collected were transported in an icebox to the laboratory of Type B Veterinary Clinic of the town for further examinations within 24 hrs.

Samples Preparation for Postmortem Inspection: Liver and gall bladder postmortem inspection by making multiple cuts and sub cuts about 1 cm thick to check the presence of fasciolosis, which made gritty sounds and bile duct thickness, palpation pressure, exerted brownish fluid and immature Fasciola. Identification of the species based on the morphological features of the agent and classify into *F. gigantica* and *F. hepatica* [15, 16].

To calculate the total sample size, the following assumptions were made: 5% desired level of precision, 95% level of confidence and 50% expected the prevalence of cattle fasciolosis in DambiDolloMunicipal abattoir, the sample size was determined by using the formula given below [17, 31].

$$n = \frac{196^2 P_{exp}(1 - P_{exp})}{d^2}$$

where,

n = required sample size,

P_{exp} = Expected prevalence,

d = desired absolute precision. But in current stud, all slaughtered animal samples were taken for good precision.

Statistical Analysis: The obtained results were encoded and recorded in an excel database analyzed by descriptive statistics survey were performed using GraphPad Instant version 3 for determination of means and the analysis of variance between the different data. The treatment, in this study, was determined using standard error and analysis of variance (p<0.05).The incidence of fasciolosis was 8/24 (33.33%) and 1356/4400 (30.82%) for females and males cattle carcasses, respectively.

Table 1: Prevalence of liver fascioliasis in examined cattle and slaughtered at the DambiDollo municipal abattoir

Examined animals	Examined	Positive	Prevalence%	Overall Prevalence%
Cattle				
Females	24	8	33.33	
Males	4400	1356	30.82	
Total	4424	1364		30.83

Seasonal Liver Fascioliasis Condemnation Rates in Examined Cattle and Samples: As illustrated in Table 2, results revealed that cattlefascioliasis is higher during spring and summer. The highest fasciolosis infection found in winter followed by autumn, spring and summer. There was a significant difference in between different seasons while there was not any significance between males and females cattle. The highest fasciolosis infection found in winter followed by autumn, spring and summer. There was a significant difference in between different seasons.The cattle fasciolosis prevalence ratewas (35.04%, 22.73%, 18.48% and 23.75%) during winter, spring, summer and autumn, respectively.

Macroscopic Liver Fasciolosis in Examined Cattle Samples: Grossly regarding fasciolosis infection during slaughterhouse postmortem inspection (Table 2) showing the external smooth liver surface declared several white or creamy tunnels ranged from few millimeters to nearly 3 cm, represented the postmortem liver fibrosis appear from external liver surface. Fasciolosis tunnels which observed from intact liver surfaces oozing grassy blackish hemorrhagic exudates. Creamyleaf-like *Fasciola* spp. about 1.5-2.0 cm in length and about 1.0 cm in width were observed by naked eye from the liver of the slaughtered cattle.

RESULTS

Prevalence of Liver Fasciolosis in Cattle: The results obtained in Table 1 indicated that the overall prevalence rate of Fasciola infection occurs in the study area were 1364/4424 (30.83%) from the total slaughtered cattle carcass.

The sex based prevalence of fasciolosis was 8/24 (33.33%) and 1356/4400 (30.82%) for females and males cattle carcasses, respectively.

Table 2: Seasonal liver fascioliasis condemnation rates in examined cattle slaughtered at the municipal abattoir of DambiDollo town was significantly different (p>0.05)

Season	Winter	Spring	Summer	Autumn
Rate of infection	35.04	22.73	18.48	23.75

DISCUSSION

Fasciola species is a parasite threatening domestic ruminants and public health. Transmission of this trematode infection is depending on the presence of intermediate “lymnaea snail” host and final host. This snail host commonly presents in high density during rainfall period annually and/or in highly moist pastures soil [13, 18]. The overall prevalence rate of fasciolosis in the examined cattle slaughtered in DambiDollo townmunicipal abattoir was about 1364/4424 (30.83%) which nearly agreed with Morsy *et al.* [19], who previously found 25.5% in Egypt. On the other hand, higher incidences of fasciolosis have been recorded by Pfukenyi and Mukaratirwa [20], who reported 37.1% in Zimbabwe and Abraham and Jude [13] recorded 44.8% in Nigeria. However, there were some remarkable lower results reported by Mellau *et al.* [21], who found 16.3% in Tanzania, Haridy *et al.* [22] noted 21.8% in Gambia Governorate, Afrakhosravi [23] reported 11.09% in Iran and Mungube *et al.* [24] recorded 26% in Kenya. Human fasciolosis was been occurred after the consumption of encysted cercaria and not by eating of animal livers infected by adult *Fasciola* spp.

The ingestion of watercress vegetables grown along contaminated water by snails and domestic ruminant fecal matters with adult parasite [25, 32]. Our reported seasonal liver fascioliasis condemnation rates revealed that is lower during winter and autumn than in spring and summer.

Accordingly, the study found that (35.04%, 23.75%, 22.73% and 18.48%) during winter, autumn, spring and summer, respectively. This finding might be attributed to raining season and presence of fresh green grazing pasturing. This finding was supported by the previous findings reported by Adedokun *et al.* [26] who reported in winter (52.3%) and in dry season (21%) in Nigerian cattle, while, fasciolosis was highest in winter (around the raining periods) and/or dampness area due to spreading of the snails host [13, 23, 27, 28].

Fasciolosis occurs mainly not only in children living in rural settings but also in people living in urban areas by metacercarial of the fluke is ingested along with watercress salad and vegetables grown along banks of water reservoirs inhabited by potential snail hosts. About 2.4 million people infected worldwide and 180 million are at risk of the infection fasciolosis commonly asymptomatic children infection with mild anemia. Humans' fasciolosis is mainly correlated with highly eggs excreted areas and not related with highly animals'

fasciolosis and sometimes infection transmitted by human stool contamination [29].

In this study, the routine macroscopic postmortem fasciolosis inspection revealed that infected liver have numerous injuries with congestion, enlargement with very hard fibrosis. Postmortem visually examination of intact liver also showing the presence of different sizes (1.5-2.7 cm) of *Fasciola* spp. impeded on the hepatic tissue with characteristic white or creamy color. Hepatic postmortem incision is showing thick wall fibrosis by fasciolosis tunnels which oozing grassy blackish exudates and debris. The trials to opening this tunnel exerted leaf-like liver flukes that diminished infected liver and carcass value and resulted in rejection of liver by consumers.

Similar lesions were observed by authors in Bangladesh [18] and in Nigeria [2, 13]. According to Ethiopian veterinary authorities, detection of fasciolosis in liver should be removed total liver condemnation or partial affected lobes after performing boiling tests and rapid phase according to parasitic infestation density and extension. The rest carcass was been released for human consumption [25]. Controlling fasciolosis mainly by anthelmintics, these only acts against at mature stages. Triclabendazole is the only drug, which affects against both immature and mature stages fascioliasis. Anthelmintic administered during December/January and from April/May for controlling chronic fasciolosis, a third dose should be given in August. However, molluscicides were been recommended for snail control [20, 30].

CONCLUSION AND RECOMMENDATION

The present study revealed a moderate fasciolosis infestation in cattle in the municipal abattoir of DambiDollo town and the study is recommended that it is important to enhance snail and fasciolosis control at farm levels to diminish the economic losses due to infection. Thorough meat inspection should also be taken on abattoir by experts.

REFERENCES

1. Oladele-Bukola, M.O. and I.A. Odetokun, 2014. Prevalence of bovine fasciolosis at the Ibadan municipal Abattoir, Nigeria. *Afr. J. Food Agric. Nutr. Dev.*, 14(4): 9055-9070.
2. Kalu, E., U. Akpabio and D.I. Gloria, 2015. A case of chronic fascioliasis in cattle slaughtered at Ubakala Abattoir. *J. Vet. Adv.*, 5(6): 1017-1022.

3. Haridy, F.M., T.A. Morsy, N.I. Gawish, T.N. Antonios and G.A. Abdel, 2002. The potential reservoir role of donkeys and horses in zoonotic fascioliasis in Gharbia governorate, Egypt. J. Egypt. Soc. Parasitol., 32(2): 561-570.
4. Dietrich, C.F., A. Kabaalioglu, E. Brunetti and J.Z. Richter, 2015. *Fascioliasis*. Z. Gastroenterol., 53: 285-290.
5. Amer, S., A. El-Khatam, S.H. Zidan, Y. Feng and L. Xiao, 2016. Identity of *Fasciola* spp. in sheep in Egypt. Parasites Vectors, 9: 623.
6. Farag, H.F., 1998. Humanfascioliasis in some countries of the Eastern mediterranean region. East Mediterr. Health J., 4(1): 156-160.
7. Walker, S.M., A.E. Makundi, F.V. Namuba, A.A. Kassuku, J. Keyyu, E.M. Hoey, P. Prodohl, J.R. Stothard and A. Trudgett, 2008. The distribution of *Fasciola hepatica* and *Fasciola gigantica* within southern Tanzania-constraints associated with the intermediate host. Parasitology, 135(4): 495-503.
8. Bazh, E.K., N.A. Beder, M. Ayoub and K. Sadek, 2012. *Fasciola* infection among cattle and buffaloes at Behera Governorate, Egypt. Zagazig Vet. J., 40: 125-136.
9. Mason, C., 2004. Fasciolosis associated with metabolic disease in a dairy herd and its effects on health and productivity. Cattle Pract., 12: 7-13.
10. Phiri, I.K., A.M. Phiri and L.J. Harrison, 2006. Serum antibody isotype responses of *Fasciola*-infected sheep and cattle to excretory and secretory products of *Fasciola* species. Vet. Parasitol., 141(3-4): 234-242.
11. Eman, K.A., M.B. Sherif and S.F. Reda, 2016. Molecular characterization of *Fasciola hepatica* infecting cattle from Egypt based on mitochondrial and nuclear ribosomal DNA sequences. Res. J. Parasitol., 11: 61-66.
12. Biu, A.A., M.I. Ahmed and S.S. Mshelia, 2006. Economic assessment of losses due to parasitic diseases common at the Maiduguri Abattoir, Nigeria. Afr. Sci., 7(3): 143-145.
13. Abraham, J.T. and I.B. Jude, 2014. Fascioliasis in cattle and goat slaughtered at calabar Abattoirs. J. Biol. Agric. Healthc., 4(18): 34-41.
14. Rokni, M., H. Mirhendi, M. Behnia, M. Harandi and N. Jalalizand, 2010. Molecular characterization of *Fasciola hepatica* isolates by RAPD-PCR and ribosomal ITS1 sequencing. Iran. Red Crescent Med. J., 12: 27-32.
15. Soulsby, E.J., 1982. Helminths, Arthropods and Protozoa of Domesticated Animals. 7th ed. Bailliere, Tindal and Cassel Ltd., London, pp: 1-300.
16. Urquhart, G.M., J. Duncan, L. Armour, J. Dunn and A.M. Jennings, 1996. Veterinary Parasitology. 2nd ed. Blackwell Science, UK, pp: 103-113.
17. Thrusfield, M., 2005. Veterinary Epidemiology. 3rd ed. University of Edinburgh, Blackwell Sciences Publishing, Oxford, pp: 626.
18. Talukder, S., M.J. Bhuiyan, M.M. Hossain, M.M. Viddin, S. Paul and M.M. Howlader, 2010. Pathological investigation of liver fluke infection of slaughtered black bengal goat in a selected area of Bangladesh. Bangladesh J. Vet. Med., 8(1): 35-40.
19. Morsy, T.A., H.S. Salem, F.M. Haridy, M.M. Rifaat, N.Y. Abo Zenadah and M. Adel El-Kadi, 2005. Farm animals' fascioliasis in Ezbet El-Bakly (*Tamya center*) Al-Fayoum governorate. J. Egypt. Soc. Parasitol., 35: 825-832.
20. Pfukenyi, D.M. and S. Mukaratirwa, 2004. A retrospective study of the prevalence and seasonal variation of *Fasciola gigantica* in cattle slaughtered in the major Abattoirs of Zimbabwe between 1990 and 1999. Onderstepoort J. Vet. Res., 71: 181-187.
21. Mellau, L.S.B., H.E. Nonga and E.D. Karimuribo, 2010. A slaughterhouse survey of liver lesions in slaughtered cattle, sheep and goats at Arusha, Tanzania. Res. J. Vet. Sci., 3: 179-188
22. Haridy, F.M., G.T. El-Sherbiny and T.A. Morsy, 2006. Some parasitic flukes infecting farm animals in Al-Santa center, Gharbia governorate, Egypt. J. Egypt. Soc. Parasitol., 36: 259-264.
23. Afrakhosravi, E.B., 2011. Epidemiology of *Fasciola hepatica* in Iran. Int. J. Biol., 4(4): 87.
24. Mungube, E., S. Bauni, B.A. Tenhagen, L. Wamae, J. Nginyi and J. Mugambi, 2006. The prevalence and economic significance of *Fasciola gigantica* and *Stilesia hepatica* in slaughtered animals in the semi-arid coastal Kenya. Trop. Anim. Health Prod., 38: 475-483.
25. Soliman, F.M., 2008. Epidemiological review of human and animal fascioliasis in Egypt. J. Infect. Dev. Ctries., 2(3): 182-189.
26. Adedokun, O.A., A.B. Ayinmode and B.O. Fagbemi, 2008. Seasonal prevalence of *Fasciola gigantica* infection among the sexes in Nigerian cattle. Vet. Res., 2(1): 12-14.

27. Oryan, A., M. Maryam, M. Moazeni, B. Nikahval and S. Barband, 2011. Liver distomatosis in cattle, sheep and goats of Northeastern Iran. *Glob. Vet.*, 6(3): 241-246.
28. Mochankana, M.E. and I.D. Robertson, 2016. A retrospective study of the prevalence of bovine fasciolosis at major abattoirs in Botswana. *Onderstepoort J. Vet. Res.*, 83(1): a1015.
29. WHO, 2007. Report of the WHO Informal Meeting on use of Triclabendazole in Fascioliasis Control. WHO, Geneva, Switzerland.
30. Spithill, T.W., P.M. Smooker and D.B. Copeman, 1999. *Fasciola gigantica*: Epidemiology, control, immunology and molecular biology. In: Dalton JP, editor. Fasciolosis. CABI Publisher, Wallingford, Oxon, UK., pp: 465-525.
31. Tesfaye, I.D., 2018. Prevalence and Risk Factors of Gastrointestinal Nematodes Infections in Small Ruminants in Sayyo District, KellemWollega Zone; Ethiopia; © IDOSI Publications, 2018: *Advances in Biological Research*, 12(2): 85-90.
32. Gemechu, B.K and I.D. Tesfaye, 2020. Prevalence and Associated Risk Factors of Gastro Intestinal Parasites in Small Ruminants in and Around Hirna Town, Western Hararghe, Ethiopia; © IDOSI Publications, 2020: *European Journal of Biological Sciences*, 12(1): 26-34.