Academic Journal of Animal Diseases 5(1): 01-15, 2016 ISSN 2079-200X © IDOSI Publications, 2016 DOI: 10.5829/idosi.ajad.2016.01.15

Review on Contagious Bovine Pleuropneumonia and its Economic Impacts

¹Zelalem Abera, ¹Demitu Mengistu, ²Geremew Batu and ³Moti Wakgari

 ¹School of Veterinary Medicine, College of Medical and Health Sciences, Wollega University, P.O. Box 395, Nekemte, Ethiopia
²West Wollega Livestock and Fisheries Development and Resource office; Gimbi, West Wollega Zone, Oromiya, Ethiopia
³Bedelle Veterinary Regional Laboratory, Bedelle, Illubabor Zone, Oromiya, Ethiopia

Abstract: Contagious bovine pleuropneumonia (CBPP) is a bacterial disease of respiratory system which is caused by Mycoplasma mycoides subspecies mycoides SC variant (MmmSC) and characterized by pneumonia and serofibrinous pleurisy. This review was aimed to elucidate the epidemiology, control measures and economic significance of CBPP, to address the economic loss of the country and to state effective control and prevention method of the disease. CBPP also epidemiologically characterized by its ability to transmit through direct contact, long incubation period, possibility of early excretion of mycoplasmas (up to 20 days) before apparition of clinical sings during the course of the disease and after recovery in "lungers" up to two years. Closeness of contact, intensity of infection and the number of susceptible animals determine the rate of spread of the disease. The clinical manifestations and lesions are typical and are no different in Africa from those seen in other countries. The post mortem lesions of CBPP include thickening and inflammation of lung tissues. Diagnosis requires the isolation of the etiological agent. In carrying out CBPP diagnosis it is necessary to differentiate this disease from other diseases that may present similar clinical signs or lesions. Treatment is recommended only in endemic areas because the organisms may not be eliminated and carriers may develop. The main problems for control or eradication are the frequent occurrence of sub acute or subclinical infections, the persistence of chronic carriers after the clinical phase and the lack of extensive vaccine coverage. Generally, CBPP is one of the major threats to cattle health and production in most African countries including Ethiopia. Relevant control measures should be implemented to minimize the incidence of CBPP zonal and national levels.

Key words: Cattle · CBPP · Economic loss · Epidemiology · Prevention · Risk factors · Transmission

INTRODUCTION

In addition to their direct role in generating food and income [1, 2], livestock are a valuable asset, serving as a store of wealth, collateral for credit and an essential safety net during times of crisis throughout the developing world [3-6] and generally generate a livelihood for 1.0 billion poor people in the world [7].

The livestock sector accounts for about 30% of the agricultural GDP in sub-Saharan Africa (SSA) and nearly 60% of the value of edible livestock products is generated by cattle [8]. However, Ethiopia has high livestock population which provides draught power, milk, meat,

fibber, fuel and fertilizer and foreign currency from hide and skin, our country is not using from her livestock as much expected due to many animal disease circulating in animal population [9, 2].

Contagious bovine pleuropneumonia (CBPP), one of these animal diseases which cause threat to our livestock, is a highly infectious respiratory disease in cattle causing lung and occasionally joint disease [10- 12]. It is caused by *Mycoplasma mycoides subspecies mycoides* SC variant (MmmSC) [13- 12]. It is difficult to see the organism even with a light microscope but growth of the organism can be seen when infectious material is cultured in the laboratory [15]. CBPP is an economically important

Corresponding Author: Zelalem Abera ,School of Veterinary Medicine, College of Medical and Health Sciences, Wollega University, P.O. Box 395, Nekemte, Ethiopia. disease of cattle that affects domestic ruminants of the genus *Bos*, mainly *Bos taurus* and *Bos indicus* and is an important disease in many of the principal pastoral areas of Africa [14, 16].

Many researchers and the Pan African program for the Control of Epizootics (PACE) has identified CBPP as the second most important trans-boundary disease in Africa next to render pest and thus, CBPP is now a major focus of activity for the program [14].

In recent years, it has been found in countries like Botswana from where it was previously eradicated [17]. It is characterized by a morbidity rate of 75% - 90%, a mortality rate from 50% to 90% and a case-fatality rate of 50% [18]. Additionally, different authors were reported as it is a respiratory illness characterized by the presence of sero-fibrinous, interstitial pneumonia, interlobular edema and hepatization giving a marbled appearance of the lung and capsulated lesions termed as sequestra in the lungs of affected cattle. Contagious bovine pleuropneumonia is a highly infectious acute, sub-acute, or chronic disease, primarily of cattle, affecting the lungs and occasionally the joints [19, 16].

There is growing evidence to indicate that the incidence of the disease is increasing in endemic areas [20]. These recent increases can be attributed to uncontrolled movement of cattle, poor disease control strategies and application of sub-standard vaccines [21, 22]. There is sufficient consensus that efficacious vaccines could contribute substantially to an integrated control program for CBPP [23]. This would involve improvement of existing and development of a new generation of vaccines [24]. So, it has become a serious obstacle to livestock development in sub-Saharan Africa and Asia.

The occurrence of sub acute, symptom less infections and chronic carriers after the clinical phase of the disease are the major problems in the control of this disease. It affects cattle production through mortality and reduced productivity [10, 20, 22]. It also retards genetic improvement and limits the ability of cattle to work [22]. Unlike some parasitic animal diseases whose impacts are confined to a single farm, the impact of CBPP is often felt at and beyond a single farm. The outbreak of CBPP in one herd poses a threat to neighboring herds especially in a production system where there is poor control of cattle movements. The economic impact of CBPP should therefore be seen beyond the farm level.

According to the report of most sub-Saharan African countries, a total of 2,612 outbreaks of CBPP between 1995 and 2002 were observed and this value represented 96% of the total number of outbreaks reported by all the countries in West, Central and East Africa and thus, found to be one of the major threats to cattle production and the lives of millions of cattle owners. An outbreak of contagious bovine pleuropneumonia (CBPP) was reported in the Somali National Regional State, Eastern Ethiopia and confirmed by detailed physical clinical examination, autopsy and bacteriological examinations to be CBPP [25]. The traditional management system and cattle movement played vital role in the spread of the diseases [4]. When the disease spreads for the first time within a sensitive cattle population, it generally causes high mortality. In countries such as Ethiopia where CBPP was reported to be prevalent, the knowledge of the diseases and factors associated to such important disease is crucial. Therefore, the objectives of this paper are to review the epidemiology, control measures and economic significance of Contagious Bovine Pleuropneumonia (CBPP) and to find the least cost method to restore health and productivity.

Contagious Bovine Pleuropneumonia (CBPP)

The Disease: Contagious bovine pleuropneumonia (CBPP) is an insidious pneumonic disease of cattle sometimes referred to as lung sickness [12, 11, 26]. There is considerable variation in severity of signs observed in cattle affected by CBPP, ranging from hyper acute through acute to chronic and subclinical [27]. CBPP has been known to occur in Europe since the 16^{th} century but it gained a world-wide distribution only during the second half of the 19^{th} century because of increased international trade in live cattle [26, 10].

It was eradicated from many countries by the beginning of the 20th century through stamping-out policies. However, the disease persists in many parts of Africa, with minor outbreaks occurring in the Middle East. The situation in Asia is unclear. There have been no reported outbreaks in Europe since 1999. In natural conditions, MmmSC affects only the ruminants of the *Bos* genus, i.e. mainly bovine and zebu cattle. MmmSC (bovine biotype) has been isolated from buffaloes in Italy (Bubalus bubalus) [26, 28] and from sheep and goats in Africa and more recently in Portugal and in India [29, 30].

Among wild animals, one single case has been reported in American buffaloes (Bison bison) and none in African buffaloes (Syncerus caffer) or other wild ruminants. Wild animals do not play a role in the epidemiology of the disease. CBPP is manifested by anorexia, fever and respiratory signs, such as dyspnea, polypnoea, cough and nasal discharges. In the case of acute outbreaks under experimental conditions, the mortality rate may be as high as 50% in the absence of antibiotic treatment. When an outbreak first occurs in an area, the mortality will be high but is often lower in the field following the primary outbreak [10, 26].

Causative Agent: The bacterial agent of contagious bovine pleuropneumonia is *Mycoplasma mycoides* [29]. *Mycoplasma* belongs to the order *Mycoplasmatales* and class Mollicutes (soft skin). *Mycoplasmas* are unique in microbiology because of their extremely small size and their growth on complex but cell-free media. Members of the *M. mycoides* group, *M. capricolum* group and Leach's group 7 form the so-called *M. mycoides* cluster, which consists of six Mycoplasmaspecies, subspecies or groups of strains, originating from bovines and goats [31].

In natural conditions, two types of Mm are recognized: large colony (LC) and small colony (SC) [29, 31, 32]. They cannot be differentiated serologically but are different morphologically, culturally and in their pathogenicity and can be distinguished through mouse protection tests. Mycoplasma mycoides subspecies mycoides Small Colony type (MmSC) affects only the ruminants of the Bos genus (mainly bovine) [10, 29]. Mycoplasma mycoides subspecies mycoides is the cause of CBPP in cattle [29, 31, 32]. Large colony types occur almost exclusively in goats, rarely in sheep while SC types cause CBPP in cattle [31]. M. mycoides mycoides large colony (LC) type does not result in disease in cattle, but causes septicemia, polyarthritis, mastitis, encephalitis, conjunctivitis, hepatitis and occasionally pneumonia in sheep and goats [33, 10].

Epidemiology: *Mycoplasma mycoides* subspecies mycoides SC type, the etiological agent of contagious bovine pleuropneumonia (CBPP) [29, 31, 33]. It can be grouped into two major, epidemiologically distinct, clusters. One cluster contains strains isolated from different European countries since 1980 and a second cluster contains African and Australian strains collected over the last 50 years [33, 35].

Hosts: Bovine, both *Bos taurus* and *Bos indicus*, are the main species that are susceptible to CBPP. Infections have also been reported from Asian buffalo (*Bubalus bubalis*), captive bison (*Bison bison*) and yak (*Poephagus grunnien*, formerly *Bos grunnien*). Sheep and goats can also be naturally infected, but with no clear associated pathology. Wild bovids and camels seem to be resistant and, so far, do not appear to be important in the

transmission of CBPP [36, 10, 16]. The African water buffalo (*Syncerus caffer*) is refractory to CBPP. CBPP prevalence with respect to age was assessed and cattle over two years were found highly affected as compared to the younger animals with significant variation [37].

Incubation Period: Incubation period of the disease is usually 1-4 months, but can be longer. After experimental inoculation into the trachea, clinical signs may appear in 2-3 weeks [10, 26, 38].

Transmission: Contagious bovine pleuropneumonia is epidemiologically characterized by its ability to transmit through direct contact, long incubation period, possibility of early excretion of *mycoplasmas* (up to 20 days) before apparition of clinical sings during the course of the disease and after recovery in "lungers" up to two years [32, 34, 39]. The organism is also present in saliva, urine, fetal membranes and uterine discharges [40].

Closeness of contact, intensity of infection and the number of susceptible animals determine the rate of spread of the disease. It is spread mainly by inhalation of droplets from infected coughing animals, especially if they are in the acute phase of the disease. Because of large numbers of MmmSC are present in bronchial secretions, nasal discharges and exhaled air, it is transmitted to susceptible animals in close contact by droplets emanating from either cattle with clinical disease or subclinical carriers that are actively excreting the organism [10, 16, 26].

Closely stabled or trucked animals are therefore most prone to infection. However, aerosols containing infected droplets may spread the disease over distances of 20 m or more [41]. Close proximity is necessary for transmission, which occurs primarily through the inhalation of infected droplets from a coughing animal [40].

Cattle are the only species affected; there is no reservoir host in wild animals and cattle movements play a very important role in the maintenance and extension of the disease [42]. Airborne spread up to 200meters is thought to be possible. Conditions under which cattle are herded closely together favour rapid spread of the disease. Apart from shedding of MmmSC by clinically ill animals, there are other sources of infection [43].

Outbreaks and Distribution: Contagious bovine pleuropneumonia was introduced in the Cape Province of South Africa in 1853 through cattle imports from the Netherlands. Following the first outbreak, CBPP quickly spread to neighboring countries and is now present in many parts of Africa. In 1904, it was eradicated from Zimbabwe followed by South Africa in 1924 and Botswana in 1939. It is generally accepted that after its arrival in South Africa from Europe, CBPP spread into East Africa by Boer settlers who trekked their cattle to the Kenyan highlands around the turn of the 20th century. Others believe it may have been present in pre-colonial times and cite Thompsonäs description of a cattle disease resembling CBPP in Maasai cattle in the1880s in Eastern Africa [44].

Today, CBPP is present in Central, East, West and parts of Southern Africa but is absent in North Africa. After examination of the number of countries reporting the disease, a more accurate estimate of distribution was provided [22]. According to OIE reports, there are about 27 sub-Saharan African countries with cases of CBPP [45]. The epidemiology of CBPP in Africa is dominated by different factors [46]. During the Pan African Rinderpest Campaign (PARC), which started in 1986, fewer countries experienced outbreaks of CBPP, due in part, to the combined vaccination against render pest and CBPP [47].

Epidemiological and clinical observations indicate that the European outbreaks of CBPP are less virulent than the disease encountered in Africa. Furthermore, CBPP in Europe seems to be far more insidious, as it is usually chronic and affected cattle show few distinctive clinical signs and rarely die [34].

An alternative or additional route of infection into Africa was the introduction of CBPP into Ethiopia and Sudan with infected Indian cattle belonging to the British Expeditionary Force in the late 19th Century [48-49]. Today, CBPP is endemic in much of sub-Saharan Africa and its incidence is increasing. The disease would then have spread into Eastern and possibly Western Africa via well established trade routes [43, 13, 32, 26].

Morbidity of CBPP: Morbidity refers to the proportion of animals affected in a given population. It includes prevalence and incidence, both of which measure the risk that a susceptible animal in a population has of contracting a disease [50]. CBPP morbidity rates vary significantly between herds. A CBPP CFT seropositive rate of 8.1% was reported by [51] in Sudan. Using the standard procedure of the Kenya Veterinary Laboratory and an antigen from the Muguga (Kenya) Veterinary Research Laboratory to test sera, [52] reported a morbidity rate of 8.3% among cattle in Sudan. Other report surveys reveal rates above 25% in Chad, Ethiopia, Guinea and Tanzania [53, 54, 55, 56]. Morbidity rates below 5% have been reported in Burkina Faso and Uganda [57, 53].

Prevalence and Incidence: The prevalence and incidence of CBPP vary according to the cattle production system concerned. Prevalence rates tend to be higher in extensive cattle production systems compared to more intensive dairy and beef production systems where animals are confined. In Chad, [55] estimated a CBPP prevalence rate of 1.6% for cattle on transhumance and a rate of 1.2% for cattle rose in agro-pastoral production systems. In Nigeria, an estimated prevalence rate of 0.29% from post mortem examinations of lesions in 81 national abattoirs [58]. [59] also estimated a prevalence rate of 0.51% in Nigeria while [53] reported rates of 2.9% for Burkina Faso, 5.4% for Mauritania and 10.5% for Mali. [60-61] reported prevalence rates of 2.8% and 4.0% in Kenya and Ethiopia respectively.

Mortality: CBPP outbreaks have been associated with various levels of mortality. Due to the debilitating nature of the disease, mortality rates have been relatively low, particularly in endemic situations. Higher mortality rates are however not uncommon. In its acute form, the mortality rate can reach 50% [15]. Mortality rates above 10% have been reported in Guinea [53] and Ethiopia [54]. Rates between 5 and 10% have been reported in Chad and Cote d'Ivoire [53] while rates below 5% have been reported in Tanzania, Uganda, Burkina Faso, Ghana and Mali [57, 56, 62, 53].

Pathogenesis: Many authors were reported that CBPP is typical example of multi-factorial diseases, where factors such as inter-current infections, crowding, inclement climatic conditions, age, genetic constitution and stress from transportation, handling and experimentation are important determinants of the final outcome of infection. An essential part of the pathogenesis of the disease is thrombosis in the pulmonary vessels, probably prior to the development of pneumonic lesions [63, 26].

Natural infection is by inhalation and results in Bronchitis, alveolitis, bronchiolitis with predominantly neutrophils and mononuclear cellular response constitute the very early inflammation in *Mycoplasma* pneumonia [64-65]. It is lobar variety of pneumonia in which the interlobular septa are dilated and prominent due to a great out pouring of plasma and fibrin in to them and it this dilated septa that give the "marbling" effect to the lung in these areas [46]. CBPP is characterized by substantial unilateral pulmonary necrosis, sometimes sequestration and marked serosanguinous fluid accumulation in interstitial and pleura [64]. Vasculitis appears to be an important component of the pathological changes in this disease, explaining the marked exudation and pleurisy. Thrombosis can explain ischemic necrosis and infarcts of the lung. Death results from anoxia and presumably from toxemia [66].

The pathogenic determinants of *M. mycoides subsp.* Mycoides SC types have not yet been elucidated completely, so the pathogenesis of CBPP is still unclear. Subcutaneous inoculation of virulent cultures into susceptible cattle causes extensive oedema, the so-called Willems' reaction, but never the natural disease. The only way to induce genuine pleuropneumonia lesions is by endobronchial inoculation of a virulent culture or, preferably, of ground lesions taken from a cow with naturally obtained CBPP. A toxin has not yet been definitively demonstrated. The galactan may be of pathogenic significance. This mycoplasmal antigen seems to contribute to the severity of the lesions by its inflammatory action as well as by its thrombotic effect on capillaries [67]. It probably also stimulates the development of connective tissue around sequestra [68].

The antigenic relationship between the mycoplasma galactan and lung tissue galactan could cause autoimmunity, with precipitation of immune complexes and damage of the pulmonary cells [67]. The lesions develop first in the lymphatic system. Thrombi that develop in the lymphatics cause coagulation of lymph, distension of interlobular septa and focal perivascular round cell infiltration. The formation o cuffs of round cells around the arterioles is the only histological pathognomonic characteristic of CBPP. The secondary lesion is characterized by alveolar involvement due to the accumulation of exudate from the foregoing changes. Necrotic foci surrounded by a band of polymorpho nuclear granulocytes often develop. These foci may develop into sequestra in chronic cases [69].

Clinical Signs: There is considerable variation in severity of signs observed in cattle affected by CBPP, ranging from hyperacute through acute to chronic and sub-clinical forms [70] and summarized as follows.

Hyper Acute Forms: The clinical signs observed in the hyperacute form are much accelerated. Affected animals may die within a week exhibiting classical respiratory signs [36, 46, 43].

Acute Forms: The early stages of CBPP are indistinguishable from any severe pneumonia with pleurisy. After an incubation period of 3-6 weeks, animals show dullness, anorexia, a fall in milk, yield irregular



Fig. 1: Source: [13]



Fig. 1: Early CBPP: Marbled lung Source: [13]

rumination with moderate fever and may show signs of respiratory disease [46, 27]. Coughing is usually persistent and is slight or dry. Sometimes fever goes up to 40-42 °C and the animal prostrates with difficulty of movement. As the typical lung lesions develop, the signs become more pronounced with increased frequency of coughing and the animal becomes prostrate or stands with the back arched, head extended and elbows abducted. While classical respiratory signs may be evident in calves, articular localization of the causative agent with attendant arthritis usually predominates [71, 27].

Sub Acute Forms: Signs may be limited to a slight cough only noticeable when the animal is exercised [72, 32]. Cattle that recover naturally are extremely weak and emaciated. Many infected animals develop chronic or milder forms of the disease, which may be either symptomless or associated with only a slight temporary rise in body temperature and some loss of condition [22, 73]. In infected herds mortality was around 2-3% [74].

Necropsy Findings: The post mortem lesions of CBPP include thickening and inflammation of lung tissues. Large amounts of straw-colored fluid may be present in

the chest cavity. A characteristic marbled appearance of the affected lungs is caused by the presence of both acute and chronic lesions in the connective tissues. Fluid accumulation in the lungs progresses to excess tissue formation (fi brosis). Encapsulated areas of diseased tissue can be found even in recovered animals [13].

Abnormalities (lesions) are generally confined to the chest cavity except in young calves, where inflammation of the limb joints (usually the carpal and tarsal joints), with increased fluid, is sometimes seen. A most striking feature of the acute disease is the very large volume of yellow fluid (up to 30 litres) containing clots, which can accumulate in the chest. The lungs (almost always one) and pleura are affected. In most cases, only the diaphragmatic lobe is involved; it is firm and fleshy, resembling liver rather than healthy pink lung. It does not collapse when the chest is opened. In acute forms, the yellowish fluid in the chest cavity may solidify and cover the lining of the chest and surface of the lung (the pleura) with a yellow or yellowish grey coating resembling a fibrin [13].

Lymph nodes in the chest may be enlarged and wet (edematous), with small necrotic foci and pinpoint hemorrhages, the difference between cortex and medulla may be indistinguishable. In the kidney cortex, white spots of dead tissue of variable size, called infarcts, can sometimes be seen. Because the lesions are so characteristic, slaughterhouse monitoring is a powerful tool to use in detecting introduction and spread of the disease although in the Ethiopian context slaughtering is achieved in almost all small butchery at backyard slaughtering [46].

Diagnosis: Contagious bovine pleuropneumonia is difficult to diagnose based on clinical signs alone as there can be many causes of severe pneumonia in cattle. But, we can diagnosis CBPP based on a history of contact with infected animals, clinical findings, immunodiagnosis tests, necropsy findings and cultural examination. CBPP frequently results in disease in only one lung as compared with other types of pneumonia in which both lungs are affected. In a herd with signs of pneumonia in adults and polyarthritis in calves, CBPP should be considered. Post mortem lesions may be more useful in the diagnosis [40]. Confirmatory diagnosis is based on the isolation of *Mccp* from clinical samples of lung [65]. **Differential Diagnosis:** In carrying out CBPP diagnosis it is necessary to differentiate this disease from other diseases that may present similar clinical signs or lesions. Some sources of confusion are: Rinderpest, Foot-andmouth disease (FMD), Haemorrhagic septicaemia (HS), Bacterial or viral broncho-pneumonia, Theileriosis (East Coast Fever), Tuberculosis, Actinobacillosis and Echinococcal (hydatid) cysts [13]. Differentials for acute infections include acute bovine pasteurellosis and bronchopneumonia and pleuropneumonia resulting from mixed infections [40].

Treatment: Under practical field conditions, when the disease breaks out in a new area, treatment is not applicable and not recommended because of reasons of disease prevention []. Treatment is recommended only in endemic areas because the organisms may not be eliminated and carriers may develop, Tylosin (10 mg/kg, IM, bid, for six injections) and danofloxacin 2.5% (2.5 mg/kg/day for 3 consecutive days) have been reported to be effective [46], but in practice farmers are treating their animals when they have no other alternative. Although the *Mycoplasmas* are susceptible to a number of antibiotics *invitro*, treatment failures are common [66].

The use of antimicrobials is controversial because of fears that it could result in more carriers, but cattle owners nevertheless do use antimicrobials to cure clinical disease and save the lives of their cattle. Oxytetracyclines (OTC) are the antimicrobials most widely used in Africa to treat CBPP. In spite of widespread and probably sub-optimal use over a long period of time resistance has not been detected, but a number of other products have been investigated [75].

Control and Prevention: The old maxim that prevention is better than cure is very relevant to dealing with CBPP. Quarantine is the first line of defense against these diseases and all countries should devote an appropriate level of resources to ensure that they implement effective border and import quarantine policies and programmes to prevent the introduction of serious livestock diseases. Eradication of CBPP has been achieved in North America, Europe, Australia and apparently in Asia, with only sub-Saharan African continent remaining significantly infected [76].

Methods used have generally involved movement control and stamping out of infected herds, although eradication in China, in which infection was widespread, was finally achieved by the use of an attenuated vaccine involving numerous passages in rabbits that was apparently effective, although it took more than 30 years to achieve eradication [75]. The most important resource in the prevention of CBPP (or any other livestock disease) is the informed animal owner or manager. Cattle owners at all levels of production must be able to recognize CBPP and know what to do when they suspect it. This can only be achieved by intensive farmer training, using media that are easily understood, highly visual and that will serve as a constant reminder of the disease and its importance [21].

CBPP control is achieved by eliminating the whole cattle herd population, i.e. stamping out, wherever the disease is detected. However, this may not prove realistic and quarantine coupled with vaccination is the most frequently used CBPP control measure [43]. Lack of the necessary resources as well as resistance by cattleowning populations preclude the use of massive culling as a control measure for CBPP in most countries and vaccination is usually regarded as the only alternative. Tylosin is quite effective for prophylactic treatment for controlling CBPP but the cost per head of cattle is much more expensive than the cost of vaccination. Hence vaccination is a more affordable alternative for farmers [41].

Vaccination: It is widely agreed that vaccination will play a central role in reducing the impact of CBPP in Africa and that it may be necessary to combine vaccination with therapy using approved antimicrobials. For this approach to succeed, a more effective vaccine than those currently available would be a great advantage [77].

The only commercial vaccines available are live attenuated vaccines using the T1/44 and T1sr strains. The former is more widely used, as it provides coverage for a year, while the duration of immunity of the T1sr vaccine is shorter. The latter has the advantage of inducing fewer adverse reactions and being unlikely to cause clinical disease, as sometimes occurs with T1/44, where especially first time vaccination may induce a Willems reaction that is sufficiently severe to require treatment [65].

Vaccination programmes as components of a CBPP eradication campaign must be comprehensively and consistently applied until there is evidence from disease surveillance that the disease has either apparently disappeared or at least the incidence has fallen to an extremely low level. The target areas for vaccination should include all but proven CBPP-free zones. In endemic regions, countrywide programmes are usually needed [21]. Live, attenuated CBPP vaccines are used. These may involve some compromise between inocuity and immunogenicity. Vaccine strains that are currently in use are T1-44 and T1-SR. T1-44 is currently the preferred vaccine in most countries. However, it has been criticized in some countries for causing excessive local reactions in vaccinated animals [76].

Status of Cbpp in Ethiopia: The irregularity and low rate of vaccinations since 1993 seem to contribute to the increased incidence of the disease and its further spread [78]. The usual blanket coverage was around 50% and never reached the desired 80-100% level. According to eleven years (1992-2002 G.C.) disease outbreak reports by Federal Ministry of Agriculture, several CBPP epidemics have been recorded from the south, south-west, west, north-west and north-east regions of the country (Table 2). The passive disease outbreak reports from 1992-2002 shows 587 outbreaks, 16,806 cases and 3,262 deaths. The highest record was in 1998 when 187 outbreaks with 5,652 cases and 1071 deaths were reported [79].

However, this data cannot be used to determine, the level and geographic feature of the disease, determine the importance of the disease, set priorities for the use of resources for disease control activities, plan, implement and monitor diseases control program, or demonstrate disease status to trading activities. Due to the insidious nature of the disease, such official data do not necessarily convey the extent of the problem caused by CBPP in Ethiopia [32].

The recent change in administrative structure and new management services at district level urges one to optimize local interventions (like vaccination). Therefore, prevalence and incidence studies are still required at such scale. However, the choice between appropriate interventions (preventive versus curative) guided by prevalence and incidence of the disease, should be compared in terms of the relative cost effectiveness of both the technologies [80].

Recent studies conducted in Western Ethiopia [81-82], Northwest Ethiopia [25], Southern Ethiopia [83] and different regions of the country [84] revealed that CBPP is posing a major threat to cattle in many parts of the country thereby causing considerable economic losses through morbidity and mortality and warranting for serious attention [85]. The cattle population at risk of CBPP and livestock production systems in CBPP endemic and epidemic zones of Ethiopia is estimated to be a total

Area		Number of examined animals	Prevalence	Average prevalence	Sources
Western Ethiopia	West Wellega	651	48%	32.5%	[37]
	East Wellega and Illubabor	105	17%		
North Omo	Konso	240	35%	56%	[87]
	Derashe	263	78%		
Southern Ethiopia	Shashemene	955	6.07%	6.14%	[88]
	Arbaminch	595	6.21%		
North West Ethiopia	Awi	2073	27.3%	19.1%	[89]
	Western Gojjam	67	12%		
Borena	Liben	1014	6.8%	9.4%	[90]
	Dire	787	12%		
	Didatuyure	246	9.4%		
Western Wellega	Bodji	506	28%	28%	[61]
Somali	Shinille and Jijiga	793	39%	39%	[32]

Acad. J. Anim. Diseases 5(1): 01-15, 2016

Table 2: Prevalence of CBPP in some areas of Ethiopia	Table 2:	Prevalence	of CBPP in	some areas	of Ethiopia
---	----------	------------	------------	------------	-------------

Source: [91]

of 13,325,700 heads of cattle. All of them are considered to be at risk of CBPP, of which 5,510,700 are in endemic zones and 7,815,000 are in epidemic zones. Generally, based on the available information, the epidemiological situation of CBPP in various parts of Ethiopia can be summarized as follows [85]:

Western Ethiopia including Western Wollega and Assosa Zones: (and possible a part of Gambella Region) are considered endemic and epidemic Southern Ethiopia: (Southern Nation, Nationalities and People region, SNNPR). Borena Zone as a whole (Oromia Region), infected since long time, is an endemic area and characterized by pastoralism; South Omo, Konso Derashe and Amaro Zones (SNNP Region) are considered as endemic, with recent outbreaks in the neighboring Zones such as Bench Maji and North Omo Zones,

Gondar and Gojam Areas: Have declared numerous outbreaks since 1993 and South Gondar and West Gojam are categorized as epiendemic areas. West Gojam zone comprises of seven districts, namely Burie-Wonberma, Denbecha, Jabitenan, Dega-Damot, Quarit, Sekela and Achefer, of which the first two districts are considered CBPP endemic and the last four districts are considered CBPP free.

The highlands of North Shewa: Were considered as CBPP free, however, [86] reported sero-prevalence rate of 54% using CFT.

Southern Tigray: seems to be recently infected with serorevealence rate of 50% reported in 1996 [86] and this can be categorized as epiendemic.

Agew Awi Zone: Comprises of four districts, namey Dangela, Ankesha-guagusa, Gungua and Banja-shikudad, of which the first three are considered as CBPP endemic and the last is with sporadic occurrence. Here mixed crop-livestock production system is practiced and the dominant livestock species are cattle (Table 1) [25].

North Eastern Ethiopia: Afar Region as a whole and Northern Somali Region may be considered as endemic, with recent outbreaks encroaching on the edge of endemic are in Southern Tigray, North Wello, North Shewa, East Shewa, (Amhara Region) and Arsi Zone (Oromia Region).

Eastern Ethiopia: In Somali Region except one zone, Shinille, which is considered to be CBPP epidemic zone, the status of the disease in all the other zones is unknown. Once introduced to a new area, initial losses in pastoral communities can be very high and its eradication is very difficult requiring major expenditure for control.

Economic Impact of CBPP: In the affected countries, enormous losses are experienced each year from the death of animals and the loss of production during convalescence. The highly fatal nature of the disease, the ease of spread and the difficulty of detecting carrier also

Country	Losses					
	 Cattle death (no)	Beef (metric tones)	Milk (metric tones)	Drought power		
B.Faso	1606	216	1312	365		
Chad	3335	299	1927	506		
Cote d'Ivior	930	83	537	141		
Ethiopia	10112	1350	8500	1645		
Ghana	474	64	387	108		
Guinea	1395	188	1140	317		
Kenya	3033	373	2316	494		
Mali	2606	350	2129	593		
Mauritania	556	75	476	126		
Niger	785	106	672	179		
Tanzania	4499	526	3527	641		
Uganda	1542	180	1208	220		
Total	30873	3810	24132	5335		

Acad. J. Anim. Diseases 5(1): 01-15, 2016

Table 3: Estimated losses in cattle and cattle products caused by CBPP

Sources: [41]

mean that close restriction must be placed on the movement of animals from enzootic areas. The economic impact of CBPP is enormous resulting in heavy losses in cattle populations [92]. Over 100,000 cattle died within two years of the introduction of CBPP in to South Africa [19]. In the Netherlands nearly 65,000 cattle died of CBPP between 1833-1850 [48]. In the early 1860s when the disease spread rapidly throughout Australia, it behaved as a virulent epidemic with losses of up to 75% of animals in an affected herd amounting to 1.4 million head.

Consideration of the true costs of control and eradication of CBPP in central and southern Africa have been detailed recently [93]. Estimates provided by indicate that animal diseases cause losses of up to 30% of the annual livestock output in developing countries [94]. CBPP is considered as a disease of economic significance because of its ability to compromise food security through loss of protein and draft power, reduce output, increase production costs due to costs of disease control, disrupt livestock/product trade, inhibit sustained investment in livestock production and cause pain and suffering to animals [21, 95].

Mortality and Morbidity Losses: Mortality losses were estimated by applying the CBPP specific mortality rate to each class of cattle at risk. Cattle production in each of the countries considered involves large pastoral communities and the effective contact rate for pastoral transhumant production systems in East Africa was estimated [41]. Morbidity losses were considered as reductions in the productivity of milk, beef and draft power. The loss in milk was estimated from two components: reductions due to dead cows that no longer produce milk and reductions due to diseased milk cows that do not produce the same quantity of milk as before [96]. Based on expert opinion, infected milk cows were assumed to lose 90% of their milk during the entire lactation period.

[98] estimated the annual losses directly or indirectly attributable to CBPP to be around US\$ 2 billion. For some countries, the losses can disrupt the entire livestock subsector and other economic sectors that depend on it. In Botswana, [98] estimated that a generalized outbreak of CBPP would result in a closure of its access to the European Union (EU) market and that the economy-wide effects of such closure would be a 60% decline in beef and other export products. In Nigeria, [99] reported economic losses due to CBPP of US\$3.6 million. In the northern part of Nigeria, [100] estimated the direct economic cost of CBPP to be US\$1.5 million of its economic cost.

Production Losses: In cattle and cattle products can be lowered by this disease. The total number of dead cattle in all twelve countries was estimated at about 30,873 head, giving an average of 2,573 head per country (range from 474 in Ghana to 10,112 in Ethiopia). The estimated total loss in beef and milk was 3,810 (318 MT per country) and 24,132 (2,011 MT per country) respectively. The former was obtained by multiplying the proportion of oxen in the herd by the number of oxen with clinical disease [41].

The loss in draft power was estimated at 445,000 workdays per country. The loss in draft power was estimated as the product of the number of infected oxen and the number of workdays per year. All physical losses in cattle, beef, milk and draft power were valued using market prices (FAO, 2004). Beef production loss by infected animals was used as a proxy for the absence of weight gain since diseased animals are assumed to not

Country	Value of losses (10	Value of losses (1000 Euros)					
	Cattle death	Beef	Milk	Animal power	Total		
B.Faso	397	432	656	117	1601		
Chad	824	598	164	162	2547		
Coted' Ivoir	230	167	296	45	710		
Ethiopia	2521	2700	4250	823	10294		
Ghana	124	128	194	35	480		
Guinea	344	375	570	102	1391		
Kenya	867	933	1390	247	3437		
Mali	643	701	1064	190	2598		
Mauritania	137	149	238	40	565		
Niger	194	211	336	57	798		
Tanzania	1121	1051	1764	320	4258		
Uganda	384	360	604	110	1454		
Total	7786	7805	12299	2248	30136		
Average	649	650	1025	1875	2511		

Acad. J. Anim. Diseases 5(1): 01-15, 2016

Table 4: Estimated value of losses in cattle and cattle products caused by CBPP

gain weight. The loss in beef production was estimated from the number of infected cattle and not from the number of dead cattle [96, 54].

The total value of output loss incurred by all twelve countries was estimated at 30.1 million Euros, giving an average of 2.5 million Euros per country (Table 4). The value of output loss ranged from 0.5 million Euros for Ghana to 10.3 million Euros for Ethiopia. The losses due to morbidity (productivity reductions in beef, milk and draft power) accounted for 74% of the total value of loss while mortality losses accounted for 26%.

Economic Costs: Many governments in Africa were found to use public funds to carry out vaccination campaigns against CBPP [20]. The cost of CBPP control by vaccination in ten (10) African countries during the PARC period and found unit costs to vary from 0.27 Euros in Ethiopia to 0.71 Euros in Cote d'Ivoire with an average of 0.42 Euros was estimated [105]. And also [106] has stated that the cost of controlling CBPP using a regional mass vaccination program in countries of Central, Eastern and Western Africa to be quite high (Euros 300 million).

Conclusion and Recommendations: Contagious bovine pleuropneumonia (CBPP) is an insidious pneumonic disease of cattle sometimes referred to as lung sickness. It was identified as the second most important transboundary disease in Africa next to render pest which needs a major focus. CBPP is a disease that causes high morbidity and mortality losses to cattle. The major risk factors those responsible for the occurrence of the disease are host factors, pathogen factors and the type of husbandry or management system. The financial implications of these losses are of great significance to both cattle owners and to the nation. Moreover, CBPP has potential to be spread in to new areas that have been considered previously as free areas. Control of CBPP is therefore important as a way to salvage the losses and increase the incomes of cattle owners. CBPP constitutes a major disease problem, which justifies a specific internationally coordinated regional control programme at least for Africa. For these facts, the following recommendations were forwarded:

- A regular programme of mass vaccination and a proven treatment regime would have a major impact on prevalence, morbidity and mortality losses.
- Abattoir surveillance for CBPP lesions is a useful technique for the detection of CBPP
- Pertinent control measures to minimize the incidence of CBPP zonal and national levels are recommended.
- Comparison of serological tests, improvement of the national veterinary infrastructure and education of livestock owners should be carried out.
- Further study on the prevalence and distribution of the disease in the country are needed.

ACKNOWLEDGEMENTS

The authors are very much grateful to the inhabitants of all staff members of School of Veterinary Medicine, College of Medical and Health Science, Wollega University for provision of materials and necessary supports during our work. Next to that, our sincere appreciation is extended for all individuals that voluntarism to provide us with sufficient information to do our research.

REFERENCES

- Perry, B.D., T.F. Randolph, S. Ashley, R. Chimedza, T. Forman, J. Morrison, C. Poulton, L. Sibanda, C. Stevens, N. Tebele and I. Yngstrom, 2003. The impact and poverty reduction implications of foot and mouth disease control in South Africa with special reference to Zimbabwe", International Livestock Research Institute, Nairobi, Kenya, pp: 152. and CD-ROM.
- Bonnet, P., R. Lancelot, H. Seegers and D. Martinez, 2011. World organization for animal health. Seventy ninth General sessions World Assembly, Paris, pp: 22-27.
- Upton, M., 2001. Trade in livestock and livestock products: International regulation and role for economic development. Food and Agriculture Organization, Livestock information and policy branch, AGAL, Livestock Policy Discussion Paper, pp: 6.
- MoA, 2006. Ministry of Agriculture and Rural Development, the Status of Animal Health Services in Ethiopia. Addis Ababa, Ethiopia.
- 5. FAO, 2009. Livestock in balance. Food and Agriculture Organization of the United Nations, Vialedelleterme di Caracalla 00153 Rome, Italy.
- Forman, S.F., G. Belton, D. Evans, B. Francois, J.L. Murray, G. Sheesley, D.A. Vandersmissen and S. Yoshimura, 2009. Moving towards the global control of foot and mouth disease: an opportunity for donors. Health Programmes in Developing Countries. Food and Agricultural Organization Expert Consultation, Rome. Rev. Sci. et tech. Off. Internat. Epiz, 28(3): 883-896.
- Naqvi, S.M.K. and V. Sejian, 2011. Global climate change: Role of livestock. Asian J. Agric. Sci., 3(1): 19-25. Nkhori PA. The impact of transaction costs on the choice of cattle markets in Mahalapye district, Botswana. Dissertation submitted to University of Pretoria, Pretoria, South Africa.
- 8. AU-IBAR, 2010. African Union-Inter African Bureau for Animal Resources.
- Zeleke, T., 2009. Common defects of sheep and goat skin in Ethiopia and causes, Ethiopia sheep and goat productivity. Technical Bulletin, 19: 1-15.
- CFSPH, 2006. Center for Food Security and Public Health, College of Veterinary Medicine, Iowa State University Ames, Iowa, pp: 50011.

- Niang, M., A. Sery, O. Cisse, M. Diallo, M. Doucoure, M. Kone, C.F. Simbe, W. Amanfu and F. Thiaucourt, 2007. Effect of antibiotic therapy on the pathogenesis of CBPP: Experimental transmission of the diseasebby contact from infected animals treated with oxytetracycline, Rome.
- Amanfu, W., 2007. The use of antibiotics for CBPP control: The challenges. Animal Health Officer (Bacterial Diseases and Zoonoses), Food and Agriculture Organization of the United Nations, Rome, Italy.
- FAO, 2002. Animal health manual. Preparation of CBPP contingency plans, Rome, pp: 5-7.
- FAO, 2007. CBPP Control: Antibiotics to the Rescue? FAO-OIE-AU/IBAR-IAEA. Consultative Group Meeting on CBPP in Africa, Rome.
- Masiga, W.N., J. Domenench and R.S. Windsor, 1996. Manifestation and epidemiology of contagious bovine pleuropneumonia in Africa. Revue Scientifique et technique Office International des Epizooties, 15(4):1283-1308.
- Brown, C. and Torres A. Eds, 2008. USAHA Foreign Animal Diseases, Seventh Edition. Committee of Foreign and Emerging Diseases of the US Animal Health Association. Boca Publications Group, Inc.
- Amanfu, W., K.V. Masupu, E.K. Adom, M.V. Raborokgwe and J.B. Bashiruddin, 1998. An outbreak of contagious bovine pleuropneumonia in Ngamiland district of northwestern Botswana. Veterinary Record, 143: 46-48.
- Blood D.C. Radostits, 1994. Veterinary medicine. 5th Ed. Baillière Tindall, London, pp: 1135.
- Trichard, C.J.V., P.A. Basson, Lugt J. Jvan der and E.P. Jacobsz, 1989. An outbreak of contagious bovine pleuropneumonia in the Owambo Mangetti area of South West Africa/Namibia: microbiological, immunofluorescent, pathological and serological findings. Onderstepoort Journal of veterinary Research, 56(4): 277-284.
- 20. FAO, 2004. Animal production and health Proceedings. FAO, Viale delle Terme di Caracalla, 00100Rome, Italy.
- FAO, 2000. CBPP status in Africa. In Report of the second meeting of the FAO/OIE/OAU/IAEA consultative group meeting on Contagious Bovine Pleuropneumonia (CBPP) Rome, Italy, pp: 24-26.

- Masiga, W.N., J. Domenench and R.S. Windsor, 1996. Manifestation and epidemiology of contagious bovine pleuropneumonia in Africa. Revue Scientifique et technique Office International des Epizooties, 15(4): 1283-1308.
- Tuslane, J.J., J.K. Litamoi, B. Morein, L. Dedieu, V. J.Palya, M. Yami, I. Abusugura, Sylla, E.M. Vilei, E.M. Abdo, J. Nicolet, A. Botelho, R. Gonçalves and J. Frey, 2000. Genomic and antigenic differences between the European and African/Australian Clusters of Mycoplasma mycoides subsp. mycoides SC. Microbiology, 146: 477-486.
- Mbithi, F., H. Wesonga, F. Thiaucourt and E.L.N. Taracha, 2004. Immune responses in cattle vaccinated against contagious bovine pleuropneumonia: Preliminary results. Kenya Agricultural Research Institute, Biotechnology laboratory, Nairobi, Kenya.
- 25. Takele, G., 1998. Epidemiological Survey of CBPP in Awi and Western Gojam zone of Amhara Region and Comparison of CFT and C-ELISA for the Diagnosis of CBPP. Addis Ababa University and Free University of Berlin, MSc thesis.
- 26. OIE, 2008. OIE Terrestrial Manual 2008. Reference Laboratories for Contagious bovine pleuropneumonia.
- RSCAHAW, 2001. Report of the Scientific Committee on Animal Health and Animal Welfare. Diagnostic Tests for CBPP. European Commission, SANCO/AH/R25/2001.
- Santini, F.G., M. Visaggio, G. Farinelli, G. Di Francesco, M. Guarducci, A.R. D'angelo, M. Scacchia and E. Di Giannatale, 1992. Pulmonary sequestrum from Mycoplasma mycoides var. mycoides SC in a domestic buffalo; isolation, anatamo-histopathology and immuno-histochemistry. Veterinaria Italiana, 4: 4-10.
- Blood, D.C. and O.M. Radostitis, 1989. veterinary madicine: Atext book of cattle, sheep, pigs, goats and horses, 7th edition. ELBS, Baillier Tindasll, 2: 778-781.
- Srivastava, N.C., F. Thiaucourt, V.P. Singh, J. Sunder and V.P. Singh, 2000. Isolation of *Mycoplasma mycoydes* small colony type from contagious caprine pleuropneumonia in India. Vet. Rec., 147: 520-521.
- OIE, 2000. Chapter 2.1.6. Manual of standards for diagnostoc tests and vaccines, 4th edn.
- 32. Gedlu, M.G., 2004. Serological, Clinical and Participatory Epidemiological Survey of Contagious Bovine Pleuropneumonia in Somali Region, Ethiopia. MSC Thesis, Faculty of veterinary medicine, Addis Ababa University, Debrezeit, Ethiopia.

- 33. Coetzer, J.A.W., G.R. Thomson and R.C. Tustin, 1994. Infectious Diseases of Livestock 1485-1494: Oxford University Press. Report of the Joint Fao Empres and Oau Ibar Regional Workshop. Contagious bovine pleuropneumonia (CBPP): prevention and control strategies in eastern and southern Africa.Contagious bovine pleuropneumonia. Rev. Sci. Tech. off. Int. Epiz., 6: 625-679.
- 34. Vilei, E.M., El-M. Abdo, J. Nicolet, A. Botelho, R. Goncalves and J. Frey, 2000. Genomic and antigenic differences between the European and African/Australian clusters of Mycoplasmamycoidessubsp.mycoidesSC. Microbiol., 146: 477-86.
- 35. FAO, 2003. Food and Agriculture Organization of the United Nations. Contagious bovine pleuropneumonia. EMRESS Transboundary Animal Diseases Bulletin, 24: 2-7.
- Provost, A., P. Perreau, C. Breard, C. Le Goff, J.L. Martel and G.S. Cottew, 1987. animale. Veterinary Epidemiology and Economics Research Unit, Department
- Desta, 1997. seroepidemiologcal investigation of CBPP in Illubabor and Wellega (Western Ethiopia), DVM Thesis, Faculty of veterinary medicine, A AU, Debrezeit Ethiopia, pp: 242.
- 38. OIE, 2003. World Animal Health in 2003. Reports on Animal Health Status. OIE, Paris.
- 39. FAO, 1996. Animal production and health paper. Prevention and control of transboundary animal disease.Rome, Italy, pp: 87-89.
- Merck, 1998. Veterinary Manual, Contagious Bovine Pleuropneumonia 8th ed. Edited by S.E. Aiello and A. Mays. Whitehouse Station, NJ: Merck and Co, pp: 1078-1079.
- Mariner, J.C., J. McDermott, J.A.P. Heesterbeek, G. Thomson and S.W. Martin, 2006. A model of contagious bovine pleuropneumonia transmission dynamics in East Africa, Preventive Veterinary Medicine, 73/1: 55-74.
- 42. Bessin, R. and R. J. Connor, 2000. The PACE strategy for supporting the control of Contagious Bovine Pleuropneumonia (CBPP). In: Report of second meeting of theFAO/OIE/OAU/IAEA consultative group on Contagious Bovine Pleuropneumonia (CBPP). Rome, Italy, pp: 39-45.
- 43. OIE, 2001. Disease Status Information, handistatus, http://www.oie.int/

- Davies, G., W.N. Masiga, M. Shifrine and W.C. Read, 1994. The efficacy of T1 strain broth vaccine against bovine pleuropneumonai: preliminary in-contact trials. Vet. Rec., 83: 239-44.
- Windsor, R.S. and A. Wood, 1998. Contagious bovine pleuropneumonia. The costs of control in central/southern Africa. Annals of the New York Academy of Sciences, 894: 299-306.
- Radostits, O.M., D.C. Blood and C.C. Gay, 1994. Veterinary Medicine: A textbook of the diseases of cattle, sheep, pigs, goats and horses. 8th ed. Baillière Tindall, pp: 910-913.
- 47. Wesonga, H.O. and F. Thiaucourt, 2000. Experimental studies on the efficacy of T1SR and T1/ 44 vaccines of Mycoplasma mycoides subspecies mycoides (small colony) against a field isolate causing contagious bovine pleuropneunia in Kenya-effect of a revaccination. Revue Elev. Med. Vet. Pays Trop., 53: 313-318.
- Laak EAter, 1992. Contagious bovine pleuropneumonia, a review. Veterinary Quarterly, 14(3): 104-110.
- 49. OIE, 1993. Meeting of the ad hoc group on contagious bovine pleuropneumonia surveillance systems: recommended standards for epidemiological surveillance of contagious bovine pleuropneumonea. OIE, Paris, France.
- Toma, B., B. Dufour, M. Sanaa, J.J. Benet, F. Moutou, A. Louza and P. Ellis, 1999. Applied veterinary epidemiology and the control of disease in populations. AEEMA, Maisons-Alfort, France, pp: 17-20.
- 51. McDermott, J.J., K.A. Deng, Tl.N. Jayatileka and M.A. El Jack, 1987. A cross-sectional cattle disease study in Kongor Rural Council, southern Sudan. Prevalence estimates and age sex and breed associations for brucellosis and contagious bovine pleuropneumonia. Preventive Veterinary Medicine, 5: 111-123.
- Zessin, K.H. and M. Baumann, 1985. Analysis of baseline surveillance data on CBPP in the Southern Sudan. Prev. vet. Med., 3: 371-381.
- 53. Kane, M., 2002. Consultancy report produced for the African Union Interafrican Bureau for Animal Resources-Pan African Programme for the Control of Epizootics. Etudehistoriquesur la péripneumonie contagieuse bovine au Burkina Faso, Côte d'Ivoire, Guinee, Mali.

- Laval, G., 2001. Experiences from CBPP follow-up in Western Wellega, Ethiopia. CBPP dynamics modeling project in Ethiopia. CIRAD/ILRI/MOA/EARO.CBPP regional workshop for Eastern African Countries. 19- 21 November 2001. Addis Ababa, Ethiopia.
- 55. Maho, A., 2001. Etude historique sur la péripneumonie contagieuse bovine au Tchad. Consultancy report produced for the African Union Interafrican Bureau for Animal Resources-Pan African Programme for the Control of Epizootics. AU/IBAR-PACE, Nairobi.
- Msami, H.M., 2001. Background information on Contagious Bovine Pleuropneumonia (CBPP) in Tanzania. Consultancy report produced for AU-IBARPACE. Nairobi, Kenya.
- Byekwaso, F. and R. Nyamutale, 2001. Background study on Contagious Bovine Pleuropneumonia (CBPP) in Uganda. Consultancy study for AU-IBAR-PACE, Naoribi, Kenya.
- Aliyu, M.M., T.U. Obi and G.O. Egwu, 2000. Prevalence of Contagious Bovine Pleuropneumonia (CBPP) in northern Nigeria. Preventive Veterinary Medicine, 47: 263- 269.
- Nawathe, D.R., 1992. Resurgence of contagious bovine pleuropneumonia in Nigeria. Rev. Sci. Tech. Off. Int. Epiz, 11: 799-804.
- Wanyoike, S.W., 1999. Assessment and mapping of contagious bovine pleuropneumonia in Kenya: Past and present. M. Sc. Thesis, Frei University of Berlin and Addis Ababa University.
- Fikru, R., 2001. Herd prevalence of CBPP, Bovinetuberculosis and Dictyocaulosis in Budju woreda. West Wellega, DVM Thesis. Addis Ababa University, FVM Debre Zeit, Ethiopia, pp: 328-329.
- Turkson, P.K., 2001. Background information on Contagious Bovine Pleuropneumonia (CBPP) in Ghana. Consultancy report produced for AU-IBARPACE. Nairobi, Kenya.
- Rosendal, S., 1993. Mycoplasma. In: Gyles, C. L and Charles, O. T. Pathogenesis of bacterial infections in animals. 2nd ed. Ames, IA: Iowa State University, pp: 297-311.
- FAO, 1997. Recognising CBPP, A Field Manual for Recognition. EMPRES (Livestock), FAO Animal Health Service, Rome.
- Nicholas, R.A.J., R.D. Ayling and L. McAuliffe, 2009. Vaccines for mycoplasma diseases in animals and man. Journal of Comparative Pathology, 140: 85-96.

- Walker, L.R., 1999. Mollicutes: In Hirsh, D. C. and Zee, Y. C. Veterinary microbiology. Blackwell Science, Inc., pp: 165-172.
- Monnerat, M.P., F. Thiaucourt, J.B. Poveda, J. Nicolet and J. Frey, 1999. Genetic and Serological Analysis of Lipoprotein Lpp A in Mycoplasma mycoides subsp. mycoides LC and Mycoplasma mycoides subsp. Capri. Clin. Diagn. Lab. Immunol., 6(2): 224-230.
- Buttery, S.H., G.S. Cottews and L.C. Lloyd, 1980. Effect of soluble factors from Mycoplasma mycoides subsp. mycoides on the collagen content of bovineconnective tissue. J. Comp. Path., 90: 303-14.
- Nunes Petisca, J.L., J.F. Costa DurAo, M. Lage, J.M. Congalves and M.J. Azevedo Ramos, 1990. Pathogenesis and pathological features of contagious bovine pleuropneumonia (CBPP).
- 70. ECHCPDG, 2001. European Commission Health & Consumer Protection Directorate General.
- Martel, J.L., M. Perrin, P. Belli and F.J. Froget, 1983. The Clinical aspects of Contagious Bovine Pleuropneumonia, Qthe diagnosis of contagious bovine pleuropneumonia and other infections with *Mycoplasma mycoides* subsp. *Mycoides*, (Ed. Hall, S.A.), Rep. N° EUR 8654, European Commission, Luxembourg, pp: 4-7.
- Abdo, E.M., J. Nicolet, R. Miserez, R. Goncalves, J. Regalla, Ch. Griot, A. Bensaide, M. Krampe and J. Frey, 1998. Humoral and bronchial immune responses in Cattle Experimentally Infected With, 59: 109-122.
- 73. Regalla, J., R. Gonçalves, N. Ribeiro, L. Duarte and A. Penha Gonçalves, 1996b. Use of immunoblotting test for carrier state difinition in contagious bovine pleuropneumonia., Mycoplasmas of ruminants: pathogenicity, diagnostics, epidemiology and molecular genetics, (Ed. J. Frey and K. Sarris), Rep. N^o EUR 16934 EN, European Commission, Luxembourg, pp: 72-74.
- 74. Guadagnini, P.F., F. De Simone, G.F. Panina, A. Gaffuri, M. Bugnetti, M. Finazzi, G. Mandelli, G. Sironi and A. Belloli, 1991. La pleuropneumonite contagiosa deibovini (PPCB). Selezione Veterinaria, 32: 3-31.
- 75. Anon, 2012. Chapter 2.4.9. Contagious bovine pleuropneumonia. Terrestrial Manual of Diagnostic Tests and Vaccines.
- Nicholas, R.A.J., J.B. Bashiruddin, R.D. Ayling and R.J. Miles, 2000. Contagious bovine pleuropneumonia: a review of recent development. Vet. Bull., 70: 827-38.

- Mair, G., E.M. Vilei, A.Wade, J. Frey and H. Unger, 2013. Isothermal loop-mediated amplification (lamp) for diagnosis of contagious bovine pleuro-pneumonia. BMC Veterinary Research, 9: 108.
- MoA, 2003. Monthly Animal Health Status Report; Ministry of Agriculture Veterinary Services, Epidemiology Unit. Addis Ababa, Ethiopia.
- MoA, 2002. Monthly Animal Health Status Report; Ministry of Agriculture Veterinary Services, Epidemiology Unit. Addis Ababa, Ethiopia.
- Lesnoft, M., G. Laval, P. Bonnet, S. Abidch, A. Workalemahu, D. Kifle, A. Peyroud, R. lancelot and F. Thiaucourt, 2004. within herd spread of CBPP in Ethiopia high lands. Prev. Vet. Med, 64: 27-40.
- Regassa, F., 2001. Herd prevalence of Contagious Bovine Pleuropneumonia (CBPP), Bovine Tuberculosis and Dictyocaulosis in Bodji woreda, West Wellega. AAU, FVM, Debre Zeit, Ethiopia, DVM thesis.
- Beyene, D., 1997. Sero-epidemiological investigation of CBPP in Illubabor and Wollega. AAU, FVM, Debre Zeit, Ethiopia, DVM thesis.
- Wondimu, D., 1996. Contagious Bovine Pleuropneumonia (CBPP): Prevalence and Evaluation of Post-Vaccination immune response (North Omo, Konso and Dirashe Regions/Ethiopia). Addis Ababa University, Faculty of Veterinary Medicine, Debre zeit, Ethiopia, DVM thesis.
- NAHRC, 2000. Report on CBPP Study in Ethiopia. Serosurveillance results National Animal Health Research Centre Ethiopian Agricultural Research Organization (EARO) Sebeta, Ethiopia.
- Afework, Y., 2000. Analysis of CBPP situation in Ethiopia, Past and Present. Ministry of Agriculture, Addis Ababa, Ethiopia.
- Roger, F. and L. Yigezu, 1995. The Situation of CBPP in Ethiopia. CIRAD-EMVT and National Veterinary Institute, Ethiopia.
- 87. Dejene. 1996.
- Asmamaw, 2003. Situation of CBCC in selected district of southern Ethiopia. MSC thesis, FVM, AAU, Debrezeit, Ethiopia, pp: 371.
- 89. Gashaw, 1998. Epidemiological survey of CBPP in Awi and western Gojjam zone of Amhara region and comparison of CFT and ELISA for the diagnosis of CBPP, Msc. Thesis. Faculty of veterinary medicine, Addis Ababa University, Debrezeit, Ethiopia, pp: 5.

- 90. Ahmed, 2004. Epidemiological study of CBPP in Borena pastoral areas using Complement Fixation Test and Competitive Enzyme Linked Immunosorbent Assay. MSC Thesis, Facult of Veterinary medicine, Addis Ababa University, Debrezeit Ethiopia, pp; 58-58.
- 91. Abstracts of DVM and MSC thesis (1985-2005)
- Blood D.C. Radostits, 1994. Veterinary medicine. 5th Ed. Baillière Tindall, London, pp: 1135.
- Windsor, R.S. and A. Wood, 1998. Contagious Bovine Pleuropneumonia: The Costs of Control in Central/Southern Africa Ann. N.Y. Acad. Sci., 849: 299-306.
- FAO (Food and Agriculture Organization), 1990. Cost/ benefit analysis for animal health programmes in developing countries.
- 95. Paskin, R., 2003. Economic and social welfare importance of transboundary animal diseases. *In* Report of a workshop of Chief Veterinary Officers/Directors of Veterinary Services of SADC Member Countries on Transboundary Animal Diseases with special reference to foot and mouth disease and contagious bovine pleuropneumonia in Southern Africa, Pretoria, 21-22 July, South Africa.
- Huddart, J.E., 1960. Bovine contagious pleuropneumonia. A new approach to field control in Kenya. Veterinary Record, 72: 1253-1254.
- 97. Masiga, W.N., P. Rossiter and R. Bessin, 1998. Present situation of CBPP in Africa and epidemiological trends. In report of the FAO/O.I.E./OAU-IBAR Consultative Group Meeting on Contagious Bovine Pleuropneumonia held in Rome, Italy, pp: 25-31.

- Kusiluka, L.J.M. and F.F. Sudi, 2003. Review of successes and failures of contagious bovine pleuropneumonia control strategies in Tanzania. Preventive Veterinary Medicine, 59: 113-123.
- Osiyemi TIO, 1981. The eradication of contagious bovine pleuropneumonia in Nigeria: Prospects and problems. Bull. Anim Health Prod. Afr, 29: 95-97.
- 100. Egwu, G.O., R.A.J. Nicholas, J.A. Ameh and J.B. Bashiruddin, 1996. Contagious Bovine Pleuropneumonia (CBPP): An update. Vet. Bull., 66: 875-888.
- 101. G.R.M. International, 1994. Herd health and productivity monitoring study: Final report of findings of three years of observations. Queensland, Australia, pp: 139.
- 102. MALDM, 1996. Ministry of Agriculture, Livestock Development and Marketing.
- 103. MNAGRI (Ministry of Agriculture), 1995.
- 104. GRU, 1997. Government of the Republic of Uganda Meat. Production Master Plan Study. Inception Report Phase I. Government of the Republic of Uganda; and Key Economic Indicators, Statistical Department. Ministry of Finance and Planning, Entebbe, pp. 57.
- 105. Tambi, E.N., O.W. Maina, A.W. Mukhebi and T. Randolph, 1999. Economic impact assessment of inderpest control in Africa. Rev. Sci. Tech. Off. Int. Epiz., 18(2): 458-477.
- 106. Thompson, G., 2003. Contagious bovine pleuropneumonia: Possible future strategies for the control of the disease in the PACE region. In: Towards sustainable CBPP control programmes for Africa. Proceedings of FAO-OIE-AU-IBAR-IAEA Consultative Group on CBPP. Third Meeting, Rome 12-14 November 2003 pp: 201.