

## Pre-Harvest Fruit Drop, Bunch Weight and Fruit Quality of 'Rothana' and 'Ghur' Date Palm Cultivars as Affected by Some Growth Regulators under Hot Arid Conditions

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**Abstract:** Pre-harvest fruit drop is a serious problem of some date palm cultivars growing in hot arid regions. During 2010 and 2011 seasons, the effect of growth regulators 2,4-D (50 and 100 ppm), NAA (100 and 150 ppm), GA<sub>3</sub> (100 and 150 ppm) and BA (100 and 150 ppm) application at 40 and 70 days from pollination on pre-harvest fruit drop and quality of 'Rothana' and 'Ghur' dates were studied. In both cultivars, the application of growth regulators at both rates significantly decreased, but not completely control, fruit drop in both cultivars. In this respect, 2,4-D and GA<sub>3</sub> were the most effective treatments followed by BA while NAA was the least effective. The reduction in fruit drop resulted in a higher bunch weight in the treated fruit than the control. The high rate of BA was more effective than the low rate in decreasing fruit drop in 'Ghur' cultivar. In 'Rothana' cultivar, bunch weight was higher with 2,4-D at both rates and in GA<sub>3</sub> and BA only at the high rate treatments than the control. While in 'Ghur' cultivar, bunch weight was higher in all growth regulators treatments than control. The ratab percentage was lower in NAA treatments than all the other treatments, except for control in 'Ghur' cultivar. Fruit and flesh weight of 'Rothana' cultivar were higher at the high rate of 2,4-D, the low rate of GA<sub>3</sub> and BA treatments than control. There were no consistence effects for growth regulators on the physical and chemical quality characteristics of fruit, possibly due to the large variations in fruit load among the treatments. It was concluded that, under hot arid conditions, the application of growth regulators of especially 2,4-D (50 ppm) and GA<sub>3</sub> (150 ppm) at both 40 and 70 days form pollination is recommended to inhibit, but not completely control, pre-harvest drop and improve fruit quality of both 'Rothana' and 'Ghur' date palm cultivars.

**Key words:** Date palm • Growth regulators • Yield • Quality • Fruit drop • *Phoenix dactylifera* L.

### INTRODUCTION

Date palm is the most successful and extremely important subsistence crop in most of the hot arid regions [1]. However, under such conditions date palms are facing environmental stress such as heat, drought and salinity which limit tree growth and productivity [2]. One of the major problems that face some date palm cultivars production in the Hada Al-Shame valley at the western region of the Kingdom of Saudi Arabia is the low fruit set and/or abnormal flowering accompanied with subsequent high fruit drop percentage [3]. Date palms, as most other fruit trees, have two waves of fruit drop. The first occurs few weeks following pollination. This drop is usually caused by lack of or incomplete pollination or fertilization.

The second drop is usually serious and more dramatic and occur five to six weeks later (end of the kimri and beginning of bisir stage) around mid May [4]. However, this second drop is called 'June drop' in many other fruit species because it usually occurs in early June [5-7]. Under the Hada Al-Shame valley conditions, both 'Ghur' and 'Rothana' date palm cultivars show normal flowering, fertilization and fruit setting. However, close to the maturation stage (bisir stage, color break around mid May) a severe fruit drop percentage (50-80%) occurs specifically in these two cultivars, in contrast to others, causing serious economic loss. This drop occurs very rapidly, often within a few days. Fruit drop is genetically, physiologically and environmentally regulated but plant stress and premature ethylene production is at the basis

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of true physiological drop [6-12]. Stress factors such as heat, drought, nutrient imbalance or deficiency and heavy crop load, all can contribute to drop [5-7]. It is well known that plant hormones such as auxins, cytokinins and gibberellins have critical role in fruit set and subsequent growth, maturation and ripening [13, 5]. After bloom, seeds formation positively contribute to fruit set, growth and retention primarily because they are the sites of hormones biosynthesis especially auxin (indole acetic acid, IAA). It is believed that fruit drop is mainly due to an imbalance between the level of IAA and ethylene within fruit tissues and abscission zone [8-12]. In this context, some growth regulators such as 2,4-dichlorophenoxyacetic acid (2,4-D), naphthalene acetic acid (NAA), benzyladenine (BA) and gibberellic acid (GA<sub>3</sub>) showed positive effects on fruit retention and are available tools to manage drop in many species, if applied at proper time [14, 6, 7, 15, 16, 17, 18, 19, 20]. In date palm, Al-Juburi *et al.* [21] reported that the application of GA<sub>3</sub>, ethephon or mixture of both growth regulators and NAA did not affect fruit set or subsequent fruit drop of 'Khaniezy' cultivar. They found also that NAA alone showed no effect on fruit set (measured at 45 days following pollination) but decreased fruit drop after 90 and 135 days from pollination compared to control. They observed that fruit drop was progressively increased with fruit development. However, the application of GA<sub>3</sub> on 'Sewy' dates at 50 or 100 ppm about 60 days from pollination had no effect on the number of fruits per bunch at commercial harvest [22]. The growth regulators namely 2,4-D; 2,4-5TP 2,4-5T; NAA and GA<sub>3</sub> which sprayed at concentrations of 25-100 ppm repeated three times at four weeks interval started at flowering, successfully produced normal seedless dates of identical quality to the seeded dates [23]. Therefore, the objective of this study was to evaluate the effect of some growth regulators on pre-harvest fruit drop in 'Rothana' and 'Ghur' date palm cultivars under hot arid conditions.

## MATERIALS AND METHODS

**Plant Materials and Experimental Procedure:** During 2010 and 2011 growing seasons 'Rothana' and 'Ghur' cultivars which show sever subsequent high pre-mature fruit drop were selected for this study. At pollination period, four uniform trees of each cultivar were selected and for each tree nine spadices were nominated for classical pollination with pollen strands collected from one male tree. All spadices were bagged with craft paper perforated bags directly following

pollination. These bags were removed after three weeks from pollination. After both 40 and 70 days form pollination (the kimri and the early beginning of bisir stage, respectively), one spadix on each female tree was sprayed with one concentration of the following growth regulators: 2,4-D at 50 and 100 ppm, NAA at 100 and 150 ppm, GA<sub>3</sub> at 100 and 150 ppm and BA at 100 and 150 ppm. On each palm one spadix was sprayed only with water and served as control. A non ionic wetting agent (Tween 20 surfactant) at 0.01% was added in all treatments including the control. All the selected palm trees received the normal cultural practices and normal fertilization and irrigation program. After 40 days from pollination, directly before the first application of growth regulators, 10 strands per bunch were selected, marked and total number of fruit were recorded. At the end of the bisir stage, the retained fruit on the selected strands were recorded and drop percentage was then calculated. At the beginning of the rutab stage for 'Ghur' and at the tamer stage for 'Rothana' (about mid June and late July for 'Ghur' and 'Rothana', respectively), bunch weight was recorded and fruit samples of 20 fruit for each bunch (replicate) were collected and kept at -20 °C for physical and chemical determinations as described below.

**Physical Characteristics, Total soluble solids, Acidity and Vitamin C Determinations:** Fruit, flesh and seed weight, flesh/seed ratio, fruit length and diameter were recorded independently in each of the 20 fruit per replicate at the bisir and the rutab stage. A homogeneous sample were prepared from these 20 fruit per replicate for measuring total soluble solids (TSS), acidity, vitamin C, total phenols and soluble tannins. Total soluble solids (TSS) were measured as Brix % in fruit juice with a digital refractometer (DR 6000, A. Kruss Optronic GmbH, Hamburg, Germany). Titratable acidity was determined in juice by titrating with 0.1N sodium hydroxide in the presence of phenolphthalene as indicator and the results were expressed as a percentage of malic acid. Ascorbic acid (vitamin C) was measured by the oxidation of ascorbic acid with 2,6-dichlorophenol endophenol dye and the results were expressed as mg/100 ml juice [24].

**Total Phenols Determination:** Total phenols were measured according to Velioglu *et al.* [25] using Folin-Ciocalteu reagent. Two hundred milligrams of fruit tissue (including skin and flesh) were extracted with 2 ml of 50% methanol for 2 h at ambient temperature. The mixture was centrifuged for 10 min and the supernatant was decanted into 4 ml vials. A 200 µl of the extract was mixed with

1.5 ml Folin-Ciocalteu reagent (previously diluted 10-fold with distilled water) and allowed to stand for 5 min before the addition of 1.5 ml of 20% sodium carbonate. After 90 min, absorbance was measured at 750 nm using a UV-Vis Spectrophotometer. The blank contains only water and the reagents. Total phenols were quantified from a calibration curve obtained by measuring the absorbance of known concentrations of gallic acid.

**Soluble Tannins Determination:** Soluble tannins were measured according to Taira [26]. Five grams of fruit tissue (including skin and flesh) was homogenised with 25 ml of 80% methanol in a blender and then centrifuged. The supernatant was collected and the precipitant was re-extracted with 80% methanol and centrifuged. The combined supernatant was brought to 100 ml with distilled water. One ml of sample solution was mixed with 6 ml distilled water and 0.5 ml Folin-Ciocalteu reagent (previously diluted 10-fold with distilled water). After 3 min, 1 ml of saturated sodium carbonate and 1.5 ml of distilled water was added and kept for 1 h at ambient temperature before measuring absorbance at 750 nm using

a UV-Vis Spectrophotometer. The blank contains only water and the reagents. Soluble tannins were quantified from a calibration curve obtained by measuring the absorbance of known concentrations of gallic acid.

**Statistical Analysis of Data:** The obtained data were statistically analyzed as a randomized complete block design with three replicates by analysis of variance (ANOVA) using the statistical package software SAS (SAS Institute Inc., 2000, Cary, NC., USA). Comparisons between means were made by *F*-test and the least significant differences (LSD) at *P* = 5%.

## RESULTS

**Fruit Drop Percentage, Bunch Weight and Rutab Percentage:** The application of growth regulators 2,4-D, NAA, GA<sub>3</sub> and BA at both the low and the high rates significantly decreased fruit drop percentage in both ‘Rothana’ and ‘Ghur’ cultivars (Table 1). In this respect, 2,4-D and GA<sub>3</sub> were the most effective treatments followed by BA while NAA was the least effective especially in

Table 1: Fruit drop percentage and bunch weight of ‘Rothana’ (at the tamer stage) and ‘Ghur’ (at the bisir and rutab stage) dates as affected by growth regulators application.

| Treatments              | Fruit drop (%) | Bunch weight (Kg) | Rutab%  |
|-------------------------|----------------|-------------------|---------|
| <b>‘Rothana’</b>        |                |                   |         |
| <i>Tamer stage</i>      |                |                   |         |
| Control                 | 51.5a          | 5.1c              | -       |
| 2,4-D 50 ppm            | 16.6cd         | 7.4a              | -       |
| 2,4-D 100 ppm           | 19.2bcd        | 7.1a              | -       |
| NAA 100 ppm             | 22.7bcd        | 5.1c              | -       |
| NAA 150 ppm             | 28.2b          | 5.6c              | -       |
| GA <sub>3</sub> 100 ppm | 18.2bcd        | 5.6c              | -       |
| GA <sub>3</sub> 150 ppm | 12.9d          | 7.3a              | -       |
| BA 100 ppm              | 23.8bc         | 5.9bc             | -       |
| BA 150 ppm              | 28.4b          | 6.9ba             | -       |
| <i>F-test</i>           | ***            | **                | -       |
| <i>LSD 0.05</i>         | 10.5           | 1.2               | -       |
| <b>‘Ghur’</b>           |                |                   |         |
| <i>Bisir stage</i>      |                |                   |         |
| Control                 | 83.2a          | 2.9e              | 17.8abc |
| 2,4-D 50 ppm            | 3.5e           | 11.0a             | 19.7a   |
| 2,4-D 100 ppm           | 4.9e           | 8.3cd             | 18.7a   |
| NAA 100 ppm             | 23.7c          | 9.1bc             | 11.6c   |
| NAA 150 ppm             | 32.7b          | 7.4d              | 12.1c   |
| GA <sub>3</sub> 100 ppm | 8.1de          | 8.2cd             | 23.2a   |
| GA <sub>3</sub> 150 ppm | 8.1de          | 10.5ab            | 21.3a   |
| BA 100 ppm              | 21.8c          | 7.1d              | 22.2a   |
| BA 150 ppm              | 12.4d          | 8.6cd             | 22.6a   |
| <i>F-test</i>           | ***            | ***               | **      |
| <i>LSD 0.05</i>         | 6.5            | 1.6               | 7.0     |

Data are the mean of 2010 and 2011 seasons. For each cultivar, means within each column followed by the same letter are not significantly different at level *P* = 0.05. \*, \*\*, \*\*\*, significant at *P* = 0.05, 0.01 and 0.001, respectively; (NS), not significant; (-), not calculated.

Table 2: Physical quality characteristics of 'Rothana' (at the tamer stage) and 'Ghur' (at the bisir and rutab stage) dates as affected by growth regulators application.

| Treatments              | Fruit weight (g) | Flesh weight (g) | Flesh/Seed (ratio) | Diameter (cm) | Length (cm) |
|-------------------------|------------------|------------------|--------------------|---------------|-------------|
| <b>'Rothana'</b>        |                  |                  |                    |               |             |
| <i>Tamer stage</i>      |                  |                  |                    |               |             |
| Control                 | 6.4c             | 5.5c             | 6.8                | 2.2           | 2.6         |
| 2,4-D 50 ppm            | 6.9abc           | 6.1abc           | 7.2                | 2.2           | 2.7         |
| 2,4-D 100 ppm           | 7.3a             | 6.5a             | 7.8                | 2.2           | 2.6         |
| NAA 100 ppm             | 6.8abc           | 5.9abc           | 7.0                | 2.3           | 2.6         |
| NAA 150 ppm             | 6.6bc            | 5.7bc            | 6.9                | 2.2           | 2.6         |
| GA <sub>3</sub> 100 ppm | 7.2a             | 6.4a             | 7.4                | 2.3           | 2.9         |
| GA <sub>3</sub> 150 ppm | 6.7abc           | 6.0abc           | 8.1                | 2.2           | 2.7         |
| BA 100 ppm              | 7.1ab            | 6.3ab            | 7.6                | 2.2           | 2.6         |
| BA 150 ppm              | 6.6bc            | 5.8bc            | 7.1                | 2.2           | 2.7         |
| <i>F-test</i>           | *                | *                | NS                 | NS            | NS          |
| <i>LSD 0.05</i>         | 0.59             | 0.55             | -                  | -             | -           |
| <b>'Ghur'</b>           |                  |                  |                    |               |             |
| <i>Bisir stage</i>      |                  |                  |                    |               |             |
| Control                 | 7.3ab            | 6.5ab            | 8.2bc              | 1.7b          | 2.9ab       |
| 2,4-D 50 ppm            | 6.2dc            | 5.5de            | 8.2bc              | 1.7b          | 2.8ab       |
| 2,4-D 100 ppm           | 7.5a             | 6.8a             | 10.3a              | 1.9a          | 3.0a        |
| NAA 100 ppm             | 6.4bc            | 5.6cde           | 6.9cde             | 1.6cd         | 2.7bc       |
| NAA 150 ppm             | 6.6bc            | 5.9cde           | 8.5b               | 1.6cd         | 2.8abc      |
| GA <sub>3</sub> 100 ppm | 7.1ab            | 6.3abc           | 8.3bc              | 1.7bc         | 3.0a        |
| GA <sub>3</sub> 150 ppm | 6.8abc           | 6.1abcd          | 7.9bcd             | 1.6bcd        | 2.8ab       |
| BA 100 ppm              | 5.5d             | 4.6f             | 5.9e               | 1.7bcd        | 2.7bc       |
| BA 150 ppm              | 6.0cd            | 5.2ef            | 6.7de              | 1.6d          | 2.6c        |
| <i>F-test</i>           | **               | ***              | ***                | **            | **          |
| <i>LSD 0.05</i>         | 0.88             | 0.81             | 1.38               | 0.11          | 0.18        |
| <b>'Ghur'</b>           |                  |                  |                    |               |             |
| <i>Rutab stage</i>      |                  |                  |                    |               |             |
| Control                 | 4.3b             | 3.6b             | 5.5b               | 1.3b          | 2.2d        |
| 2,4-D 50 ppm            | 4.9ab            | 4.4ab            | 8.7a               | 1.4b          | 2.5bc       |
| 2,4-D 100 ppm           | 6.2a             | 5.6a             | 8.9a               | 1.7a          | 2.9a        |
| NAA 100 ppm             | 5.4ab            | 4.7ab            | 6.5ab              | 1.3b          | 2.4cd       |
| NAA 150 ppm             | 6.0a             | 5.3a             | 7.5ab              | 1.4b          | 2.6b        |
| GA <sub>3</sub> 100 ppm | 5.2ab            | 4.5ab            | 6.8ab              | 1.4b          | 2.5cb       |
| GA <sub>3</sub> 150 ppm | 5.2ab            | 4.5ab            | 6.8ab              | 1.4b          | 2.4cb       |
| BA 100 ppm              | 4.3b             | 3.7b             | 6.6ab              | 1.4b          | 2.5cb       |
| BA 150 ppm              | 4.3b             | 3.6b             | 5.2b               | 1.4b          | 2.4cd       |
| <i>F-test</i>           | *                | *                | *                  | *             | ***         |
| <i>LSD 0.05</i>         | 1.3              | 1.3              | 2.5                | 0.22          | 0.19        |

Data are the mean of 2010 and 2011 seasons. For each cultivar, means within each column followed by the same letter are not significantly different at level  $P = 0.05$ . \* and \*\*, significant at  $P = 0.05$  and  $0.01$ , respectively; (NS), not significant; (-), not calculated.

'Ghur' cultivar (Table 1). The high rate of BA was more effective than the low rate in decreasing fruit drop only in 'Ghur' cultivar. In 'Rothana' cultivar, the bunch weight was significantly higher with 2,4-D at both rates and with GA<sub>3</sub> and BA only at the high rate treatments than the control. While in 'Ghur' cultivar, the bunch weight was higher with all of the growth regulators treatments than control. NAA application decreased the rutab percentage compared with all other treatments, except the control in 'Ghur' cultivar (Table 1).

**Physical Quality Characteristics of Fruit:** Fruit and flesh weight of 'Rothana' cultivar, were higher at the high rate of 2,4-D, the low rate of GA<sub>3</sub> and BA treatments than the control (Table 2). However, the flesh/seed ratio, diameter and length of the fruit were not affected by any of the treatments. While in 'Ghur' cultivar, at the bisir stage, both fruit and flesh weight were lower at low rate of 2,4-D and at low and high rates of BA treatments than the control (Table 2). The flesh/seed ratio and fruit diameter were higher at the high rate of 2,4-D than all other

Table 3: Chemical quality characteristics of ‘Rothana’ (at the tamer stage) and ‘Ghur’ (at the bisir and rutab stage) dates as affected by growth regulators application.

| Treatments              | TSS<br>(Brix %) | Acidity<br>(%) | Vitamin C<br>(mg/100g fw) | Total phenols<br>(mg/g fw) | Soluble tannins<br>(mg/g fw) |
|-------------------------|-----------------|----------------|---------------------------|----------------------------|------------------------------|
| <b>‘Rothana’</b>        |                 |                |                           |                            |                              |
| <i>Tamer stage</i>      |                 |                |                           |                            |                              |
| Control                 | 44.2c           | 0.41d          | 3.2bdc                    | 0.25c                      | 2.23                         |
| 2,4-D 50 ppm            | 47.6b           | 0.55bc         | 3.5bc                     | 0.28bc                     | 1.95                         |
| 2,4-D 100 ppm           | 52.9a           | 5.55bc         | 2.7d                      | 0.33ab                     | 2.20                         |
| NAA 100 ppm             | 47.1b           | 0.51c          | 4.2a                      | 0.32abc                    | 2.28                         |
| NAA 150 ppm             | 42.5d           | 0.61a          | 3.7ab                     | 0.30abc                    | 2.20                         |
| GA <sub>3</sub> 100 ppm | 42.5d           | 0.49c          | 3.0cd                     | 0.25c                      | 2.05                         |
| GA <sub>3</sub> 150 ppm | 37.0e           | 0.41d          | 3.0cd                     | 0.26c                      | 2.02                         |
| BA 100 ppm              | 45.6c           | 0.57ab         | 3.7ab                     | 0.36a                      | 2.10                         |
| BA 150 ppm              | 47.0b           | 0.49c          | 3.6abc                    | 0.35a                      | 2.05                         |
| <i>F-test</i>           | ***             | ***            | **                        | *                          | NS                           |
| <i>LSD 0.05</i>         | 1.4             | 0.06           | 0.70                      | 0.06                       | -                            |
| <b>‘Ghur’</b>           |                 |                |                           |                            |                              |
| <i>Bisir stage</i>      |                 |                |                           |                            |                              |
| Control                 | 30.0d           | 0.29a          | 6.2ab                     | 0.59                       | 10.3e                        |
| 2,4-D 50 ppm            | 33.0b           | 0.28ab         | 6.2ab                     | 0.70                       | 13.2bc                       |
| 2,4-D 100 ppm           | 36.0a           | 0.24bc         | 5.0bc                     | 0.70                       | 11.1de                       |
| NAA 100 ppm             | 28.0e           | 0.25abc        | 7.0a                      | 0.69                       | 11.9cd                       |
| NAA 150 ppm             | 24.0f           | 0.28ab         | 7.0a                      | 0.78                       | 10.1e                        |
| GA <sub>3</sub> 100 ppm | 27.0e           | 0.23c          | 6.7a                      | 0.96                       | 12.8c                        |
| GA <sub>3</sub> 150 ppm | 32.0bc          | 0.27abc        | 7.0a                      | 1.02                       | 14.5ab                       |
| BA 100 ppm              | 31.0cd          | 0.28ab         | 5.7abc                    | 0.94                       | 14.8a                        |
| BA 150 ppm              | 33.0b           | 0.25abc        | 4.5c                      | 0.76                       | 13.1bc                       |
| <i>F-test</i>           | ***             | **             | **                        | NS                         | ***                          |
| <i>LSD 0.05</i>         | 1.5             | 0.05           | 1.3                       | -                          | 1.4                          |
| <b>‘Ghur’</b>           |                 |                |                           |                            |                              |
| <i>Rutab stage</i>      |                 |                |                           |                            |                              |
| Control                 | 50.0g           | 0.32e          | 5.7ab                     | 0.49b                      | 5.8ab                        |
| 2,4-D 50 ppm            | 58.5d           | 0.43cb         | 3.2d                      | 0.51b                      | 4.7d                         |
| 2,4-D 100 ppm           | 64.5a           | 0.55a          | 4.0cd                     | 0.54b                      | 5.0cd                        |
| NAA 100 ppm             | 49.0g           | 0.36de         | 3.2b                      | 0.49b                      | 5.5abc                       |
| NAA 150 ppm             | 61.2b           | 0.37cde        | 5.2b                      | 0.48b                      | 4.7d                         |
| GA <sub>3</sub> 100 ppm | 58.3d           | 0.35de         | 6.2a                      | 0.66a                      | 5.0cd                        |
| GA <sub>3</sub> 150 ppm | 56.5e           | 0.36de         | 6.2a                      | 0.68a                      | 5.5abcd                      |
| BA 100 ppm              | 55.0f           | 0.40cd         | 4.2c                      | 0.49b                      | 6.1a                         |
| BA 150 ppm              | 60.0c           | 0.47b          | 5.7ab                     | 0.51b                      | 5.2bcd                       |
| <i>F-test</i>           | ***             | ***            | ***                       | ***                        | **                           |
| <i>LSD 0.05</i>         | 0.96            | 0.07           | 0.85                      | 0.09                       | 0.75                         |

Data are the mean of 2010 and 2011 seasons. For each cultivar, means within each column followed by the same letter are not significantly different at level P = 0.05. \*, \*\*, \*\*\*, significant at P = 0.05, 0.01 and 0.001, respectively; (NS), not significant; (-), not calculated.

treatments. The BA application at both rates significantly decreased the flesh/seed ratio compared to control. Fruit length was lower at the high rate of BA treatment than control (Table 2). At the rutab stage, fruit and flesh weight in ‘Ghur’ cultivar were higher at the high rate of 2,4-D and NAA than the control (Table 2). The flesh/seed ratio was higher at the high and low rates of 2,4-D than the control. Fruit length was higher in most of the growth regulator treatments than the control. The 2,4-D application at the high rate produced longer fruit than all other treatments (Table 2).

**Chemical Quality Characteristics of Fruit:** In ‘Rothana’ cultivar, the TSS concentration was higher than all other treatments (Table 3). The application of 2,4-D at high and low rate, NAA at low rate and BA at high rate significantly increased the TSS concentration compared to control, while the NAA at high rate and GA<sub>3</sub> at both high and low rate significantly decreased the TSS concentration. All the growth regulators treatments significantly increased acidity compared to control. The highest acidity was obtained at the high rate of NAA followed by the BA at the low rate treatments (Table 3).

Vitamin C concentration was higher at the low rate of NAA than all other treatments except for, the high rate of NAA and the BA at both studied rates. Total phenols concentration was higher at the high rate of 2,4-D and at the BA at both rates than the control. However soluble tannins concentration was not affected by any of the treatments. In 'Ghur' cultivar, at the bisir stage, the TSS concentration was higher at the high rate of 2,4-D than all other treatments (Table 3). Also, the 2,4-D at the low rate, GA<sub>3</sub> and BA at the high rate significantly increased the TSS concentration compared to control. While, the NAA at both rates and the GA<sub>3</sub> at the low rate significantly decreased the TSS concentration compared to all other treatments. Acidity concentration was not affected by the applied growth regulators compared to control except for, the high rate of 2,4-D and the low rate of GA<sub>3</sub> that decreased acidity concentration. Also, vitamin C concentration was not affected by the applied treatments compared to control except for, the high rate of BA that lowered vitamin C than the control (Table 3). The total phenols concentration was not affected by any of the treatments. However, the soluble tannins concentration was higher at the low rate of 2,4-D, the high rate of NAA and the low and the high rates of GA<sub>3</sub> and BA compared to control. At the rutab stage, the TSS concentration was higher in all treatments than the control except for, the high rate of NAA (Table 3). The high rate of 2,4-D and NAA showed the highest TSS concentration that was higher than all other treatments. Acidity concentration was higher at the high rate of 2,4-D than all other treatments. The application of BA at both rates and 2,4-D at low rate significantly increased acidity concentration compared to control. Vitamin C concentration was lower with 2,4-D at both rates and BA at low rate treatments than the control. Total phenols concentration was higher at the GA<sub>3</sub> at both high and low rates than all other treatments. Soluble tannins concentration was lower with 2,4-D at both rates, the high rate of NAA and the low rate of GA<sub>3</sub> and BA treatments than the control (Table 3). Generally, during ripening (changing from bisir to rutab stage) of 'Ghur' dates, the concentration of TSS and acidity was greatly increased while, soluble tannins, total phenols and vitamin C concentration was greatly decreased (Table 3). Also, the physical quality characteristics were decreased due to lose of moisture (Table 3).

## DISCUSSION

In contrast to other date palm cultivars growing in the Hada Al-Shame valley, both 'Rothana' and 'Ghur' are

severing heavy fruit drop percentage (about 50-80%) close to the maturation stage (around mid May). This phenomenon occurs very rapidly, often within a few days. In the current experiment, the application of growth regulators 2,4-D, NAA, GA<sub>3</sub> and BA at both low and high rates significantly decreased, but not completely control, fruit drop in both cultivars (Table 1). In this respect, 2,4-D and GA<sub>3</sub> were the most effective treatments followed by BA, while NAA was the least effective. The reduction in fruit drop resulted in a significantly higher bunch weight in the treated fruit than the control (Table 1). However, in other date palm cultivars that normally show a slight fruit drop close to maturation, GA<sub>3</sub> and ethephon sprayed alone or in combination with NAA had no effect on fruit set percentage or on subsequent fruit drop of 'Khenazy' cultivar [21]. They found, however, that the NAA application alone decreased fruit drop after 90 and 135 days from pollination compared to control. Also, the application of GA<sub>3</sub> on 'Sewy' dates at 50 or 100 ppm about 60 days from pollination had no effect on the number of fruits per bunch at commercial harvest [22]. The growth regulators, namely 2,4-D; 2,4-5TP 2,4-5T; NAA and GA<sub>3</sub> sprayed at concentrations of 25-100 ppm repeated three times started at flowering successfully produced normal seedless dates of identical quality to the seeded dates [23]. During the last few decades, the use of plant growth regulators, especially 2,4-D and NAA has become a widespread practice to control fruit drop of different species [14, 7]. It is believed that fruit drop is mainly due to an imbalance between the level of auxin and ethylene within fruit tissues. Ethylene trigger the system for abscission layer formation and the hydrolytic enzymes (cellulase and polygalacturonase) that break down the cell walls leading to fruit drop [15, 20]. Accordingly, the combination of Retain and naphthaleneacetic acid (NAA) controlled preharvest drop of several apple cultivars better than either chemical alone [7-10]. NAA alone controls the genes associated with abscission zone formation (MdPG2 and MdeG1) but stimulates ethylene production in the fruit which advances ripening caused by polygalacturonase (controlled by MdPG1 gene). In contrast, Retain acts by controlling ethylene biosynthesis and thus the genes associated with fruit abscission (MdPG2) and fruit softening (MdPG1) [7-12]. At the time of severe and fast fruit drop (around mid May) in both 'Rothana' and 'Ghur' date palm cultivars, the seed was almost developed and the endocarp lignified. The recorded air temperature in the orchard was generally higher by about 5.5 degrees than that in April. According to Racskó *et al.* [5] in one-seeded fruits, e.g. stone fruits, June drop increases when the growth of embryo is

intense. At this stage, the embryo consumes most of the endosperm which is coincident with a lag phase of fruit growth often associated with fruit drop. When the embryo completes its growth the secondary endosperm appears and start to produce auxin that required for inhibiting abscission. In the current experiment, none of the used growth regulators completely controlled fruit drop (Table 1). It has been also reported that in warm seasons (over 95°F in August) Retain suppressed ethylene production but did not adequately control pre-harvest drop of McIntosh apples [6]. The physiological responses of date palm fruit to environmental stress such as high temperature and drought might induce such a high degree of fruit drop. These climatic factors might induce specific changes within the abscission zone (e.g. lack of carbohydrate supply, reduced export of indole-3-acetic acid (IAA) out of the fruit; increased fruit ethylene synthesis) which subsequently leads to fruit drop. Initial results reported on mango fruit by Roemer *et al.* [27] showed a correlation between a reduced IAA export and fruit drop. The higher the temperatures at the time of McIntosh apples begin to ripen and to produce ethylene the more severe and earlier is pre-harvest fruit drop [28]. In apricot, fruit drop ensued most clearly close after the initial lignification of the stone (endocarp) at the mid of May. About the 34% of the fruits of the tree were dropped. Under conditions of drought, the water absorption of the leaf is stronger than that of the fruit therefore the fruits are more exposed to be dropped than leaves [5]. Our data showed that bunch weight was significantly higher with 2,4-D at both rates and GA<sub>3</sub> and BA treatments at the high rates than all the other treatments including the control (Table 1). In confirmation, the application of GA<sub>3</sub> on 'Sewy' dates at 50 or 100 ppm at 60 days from pollination increased bunch weight, fruit and flesh weight, fruit diameter and length and slightly increased total tannins concentration compared to the control [22]. The rutab percentage in 'Ghur' cultivar was lower with NAA treatments than all the other treatments, except for the control (Table 1). Also, at the bisir stage, NAA decreased the TSS concentration than all other treatments, indicating delayed in fruit ripening (Table 3). Delayed in fruit ripening of 'Zahdi' date palm up to one month by NAA application at 40 and 60 ppm has been previously reported [29]. In our study, there were no consistence effects for the applied growth regulators on the physical and the chemical quality characteristics of fruit (Tables 2 and 3). This might be due to the large variations in fruit load among the treatments in comparison with the control (Table 1). Despite the

relatively high fruit load in growth regulators treated bunches in both cultivars (Table 1), most of the treatments give even higher or similar fruit and flesh weight compared with the control except for, the low rate of 2,4-D and BA at both rates in 'Ghur' cultivar at the bisir stage (Table 2). The positive effect of growth regulators on fruit quality might be through influencing cell size and numbers and/or the movement of assimilates into the fruit. Cell size and numbers was increased in 'Sayer' date palm cultivar by the NAA application [30]. Also, GA<sub>3</sub> application has been found to increase both the rate and amount of assimilates moved into the grape berries resulted in higher berry weight and size [31]. BA may have affected the sink strength of individual fruit, directly by increasing sink activity or indirectly by stimulating fruit growth and increasing sink size in apples and other fruit [32-34]. Thus, the competition among the fruit within a bunch and among bunches on the same tree on assimilates might also influence fruit growth and retention. Ben Salah [4] reported that the minimum fruit drop percentage was obtained when 1/3 of strands were thinned out from the inside of the bunch of 'Khenazy' cultivar. It was concluded that, under the Hada Al-Shame valley conditions, the application of growth regulators, especially 2,4-D at 50 ppm and GA<sub>3</sub> at 150 ppm at both 40 and 70 days from pollination is recommended to reduce, but not completely control, fruit drop and improve quality of both 'Rothana' and 'Ghur' date palm cultivars. The possibility of re-applying the growth regulators alone or in combinations, with especially ethylene inhibitors and/or manipulating crop load, that may control fruit drop is worthy of further investigation.

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