Extraction and Separation of Glucosinolates from Brassica Oleraceae var Rubra

J. Renuka Devi and E. Berla Thangam

Department of Biotechnology, School of Bioengineering, SRM University, Kattankulathur-603203

Abstract: Glucosinolates occur in a variety of Crucifers and among various classes of crucifers, genus Brassica contains a rich content of glucosinolates. Glucosinolates are found to have detrimental activity against various types of cancer such as breast, lung and colon. The intact glucosinolates were separated by cold maceration and the fractions were separated using Column, Thin layer (TLC) and Paper chromatographies. The fractions were further analyzed using chromogenic reagent and the Rf values were calculated. The purified fraction were further subjected to HPLC for purity assessment and confirmed with the standard graphs. It was concluded that the yield depends on the different stages of plant development. Any pure extraction study has to be performed by taking the plants life cycle stage and age of the plant as a parameter to optimize the yield.

Key words: Brassica oleraceae · Glucosinolates · Separation · Myrosinase · Chromatography

INTRODUCTION

In recent years, plant research has grown to a greater scale with scope in the treatment of various diseases. The nutrition campaign "five per day" (for better health) which is supported by governmental, economical authorities and health organizations in several countries encourages the population to eat five or more servings of fruit and vegetables every day [1]. Epidemiological studies demonstrated that vegetable consumption decreases risk of various diseases, especially cancers of various types [2]. Among the plant groups Brassica research continues to attain greater pace for the past 15 years [3]. Vegetables of the Brassica genus (broccoli, cabbage, cauliflower, radish, mustard, etc.) have received much attention, because they are reported to have anticancer activity both in vitro and in vivo [4].

Broccoli sprouts are widely consumed in many parts of the world. There have been no reported concerns with respect to their tolerance and safety in humans [5]. Agudo [6] reported that the consumption of cruciferous vegetables and hence of glucosinolates, is relatively low within Europe, which in turn is lower than in North America and several Asian populations.

A considerable number of epidemiological studies revealed an inverse relationship between consumption of Brassica vegetables (broccoli, red cabbage, Brussels sprout, kale, cauliflower, cabbage) and risk of cancer in various human organs [1]. Crucifers are important sources of glucosinolates (GLs), [7] whose de-generated products like isothiocyanates were attributed to chemo-preventive activity [8]. Glucosinolates are anionic, hydrophilic plant secondary metabolites and play an important role in the prevention of cancer and other chronic and degenerative diseases [9]. However Abdull Razis and Bagatta [8] showed that the intact glucosinolates are capable of carcinogen-metabolizing enzyme systems Γ101. Glucosinolates may breakdown to form isothiocyanates and/or nitriles in plant material during processing by the action of the endogenous enzyme myrosinase (thioglucoside glucohydrolase) within or gastrointestinal tract by the action of commensal microflora [11]. Aires et al. [12] demonstrated the potential of these glucosinolates and their respective enzymatic hydrolysis products on bacteria isolated from the human intestinal tract [13]. Various collaborative research states that Indole compounds will be produced as a consequence of breakdown of glucosinolates by enzyme myrosinase which is present in intact tissue. Many researchers support that the chemopreventive effect of brassica vegetables and their constituents in various animal and clinical experiments [14]. Red cabbage (Brassica oleraceae var rubra) contains similar amounts of glucoraphanin and glucobrassicin but, in addition, appreciable amounts of glucoiberin, progoitrin, sinigrin, gluconapin and glucoerucin, while neo-glucobrassicin occurred at trace levels [2].

Our study focus on the extraction of Brassica oleraceae var rubra using methanol by cold maceration and separation of the constituents by various chromatography techniques.

MATERIALS AND METHODS

Plant Materials: Brassica oleracea var rubra was obtained from the local grocer and the leaves were separated, washed and air dried. Plant material was lyophilized and subsequently macerated with 80% methanol (4ml/g original fresh weight) and extracted for 18 hr before filtration. The plant material was re- extracted twice with cold 80% methanol for 4hr each time with frequent swirling. The filtrates were combined, concentrated under reduced pressure at 30°C and subjected to column chromatography [14, 15].

Chromatographic Investigation

Column Chromatography: The obtained residue (total glucosinolate) of each plant was further purified on silica column using the mobile phase ethyl acetate.

Thin Layer Chromatography (TLC): The fractions were separated using the solvent mixture as hexane: ethyl acetate (80:20%) for separation of Brassica oleracea var rubra.

Paper Chromatography of the Glucosinolates: The purified glucosinolates of each plant (broccoli and red cabbage) was subjected to paper chromatography on (Whatmann 3mm) using the solvent system butanol: ethanol: water (4:1:4, two runs) and descending technique. The developed chromatograms were air dried and then the outer margins of each chromatogram were sprayed with ammoniacal silver nitrate reagent. The separated bands were cut off and diluted separately with methanol (80%) to obtain the pure compounds. The compounds were identified using chemical tests.

Isolation of Myrosinase from Mustard Cake: Myrosinase was isolated from *Aspergillus* spp using mustard extract medium. To 100 ml of potato dextrose broth, fresh culture of *Aspergillus* spp was added and shaken for 15 min with intermittent heating initially and continued further for 3 hrs. The culture was filtered and centrifuged; the supernatant was collected and stored. Protein test was done using ninhydrin test [16].

Chromogenic Reagents: Compounds were detected by means of ferric nitrate in solution of nitric acid and myrosinase solution (pH 7) as previously described [14, 15]. The compounds were developed on TLC plates and sprayed with myrosinase (pH 7) with ascorbic acid and allowed to dry for 2 hrs and spraying with ferric nitrate reagent.

HPLC Analysis: HPLC was performed with Zorpax Eclipse XDB C $_{18}$ 5 μ m 4.6 mm \times 150mm column. The mobile phase was acetonitrile: water (20: 80). The sample (20 μ l) was injected in to the column by means of an autoinjector and peaks were monitored at 229 nm. The chromatograms were recorded for 30 minutes. Mobile phases were pumped at 1.5 ml/ min [17-19].

RESULTS

Determination of Total Glucosinolates: The residue of glucosinolates after extraction was estimated to be 1.8 and 1.7 g, respectively. The fractions from the column were collected and were subjected to TLC for further analysis.

Thin Layer and Paper Chromatography: The chromatograms from the column chromatography were air dried and designated red cabbage as RC1, RC2, RC3 and RC4. The Rf values of different samples are given in Table 1.

Based on the Rf values of the fractions and upon spraying with ferric nitrate reagent the red brown colour may indicate the presence of glucosinolates.

Table 1: Chromatographic properties of Glucosinolates

Solvent system	Rf values for compounds			
Butanol:ethanol:water	RC1	RC2	RC3	RC4
	0.46	0.66	0.76	0.62
Chromogenic reagent				
Ferric nitrate in nitric acid	Brownish	Bluish	Bluish	Bluish green
Myrosinase (pH 7)+ Ferric nitrate in nitric acid	Brown	Reddish brown	Reddish brown	Reddish brown

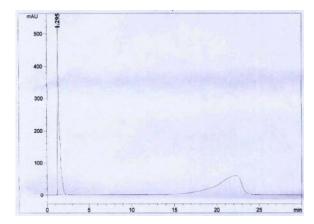


Fig. 1: HPLC for sample RC1

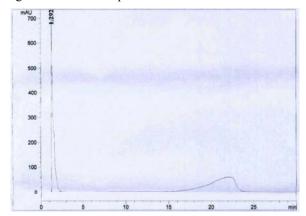


Fig. 2: HPLC for sample RC2

HPLC Analysis: The fractions from chromatography were individually subjected to analysis and the graphs are depicted in the picture. Since internal standards are not available the glucosinolates are confirmed by comparing the retention times of peaks with the standard graphs from the literature [20]. The HPLC graphs are shown below.

DISCUSSION

The objectives of the current extraction and purification were to analyze the extracted compounds further to study the anti-cancer activity. Marcelo Fonseca Xavier studied the optimization of column process and optimization conditions for extraction of anthocyanins from red cabbage [18]. Michael Meyer made a comparative study of glucosinolate levels in commercial broccoli and red cabbage. [1] McNaughton discussed about the creation of a food composition database for the estimation of dietary intakes of glucosinolates [21].

Kim and Durret states that the Levels of leaf glucosinolates are regulated during plant development.

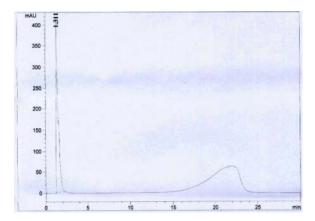


Fig. 3: HPLC for sample RC3

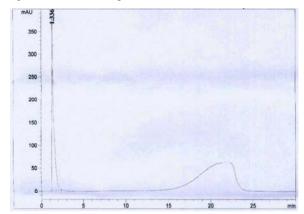


Fig. 4: HPLC for sample RC4

The level of glucosinolates increases in response to mechanical damage or insect feeding. [22] Zhou et al. used a three step process to purify main glucosinolate from Brassica oleracea. The steps involved, extraction with methanol, separation and purification by chromatographic column on alumina support; and followed by a reversed-phase separation by octadecyl (C18) silica [23]. Barillari et al. measured the two important (GRH) glucosinolate viz., Glucoraphasatin glucoraphenin (GRE) from Raphanus sativus sprouts and seeds during different stages of development. They found that in comparison to the seeds, the GRE content in sprouts decreased with development. Whereas, GRH content increased steadily up to a 25-fold increases. [24].

Different glucoraphenins have role in different stages of plant development and the extraction optimization has to be done based on the plants age and stage of development for commercial extraction studies, which is a scope for future study. Further scope also includes studying the various processes like anti-cancer and anti-microbial activity and also extraction of active compounds of importance.

REFERENCES

- Meyer, M. and S.T. Adam, 2008. Comparison of glucosinolate levels in commercial broccoli and red cabbage from conventional and ecological farming. Eur Food Res. Technol., 228: 1429-1437.
- 2. Young S Kim and J.A. Milner, 2005. Targets for indole-3-carbinol in cancer prevention. J. Nutritional Biochemistry, 16: 65-73.
- 3. Hecht, S.S., 2000. Inhibition of carcinogenesis by Isothiocyanates. Drug metabolism Reviews, 32: 395-411.
- Gerlinde Pappa, Maike Lichtenberg, Renato Iori, J. Barillari and H. Bartsch, 2006. Comparison of growth inhibition profiles and mechanisms of apoptosis induction in human colon cancer cell lines by isothiocyanates and indoles from Brassicaceae. Mutation Res., 599: 76-87.
- Shapiro, T.A., J.W. Fahey, A.T. Dinkova-Kostova, W.D. Holtzclaw and K.K. Stephenson, 2006. Safety, tolerance and metabolism of broccoli sprout glucosinolates and isothiocyanates: a clinical phase I study. Nutr Cancer., 55: 53-62.
- Agudo, A., R. Ibanez, P. Amiano, E. Ardanaz, A. Barricarte, *et al.*, 2008. Consumption of cruciferous vegetables and glucosinolates in a Spanish adult population. Eur. J. Clin. Nutr., 62: 324-331.
- Ugolini, L., G. De Nicola and S. Palmieri, 2008. Use of reverse micelles for the simultaneous extraction of oil, proteins and glucosinolates from cruciferous oilseeds. J. Agric. Food Chem., 56: 1595-1601.
- 8. Abdull Razis, A.F., M. Bagatta, G.R. De Nicola, R. Iori and C. Ioannides, 2010. Up-regulation of cytochrome P450 and phase II enzyme systems in rat precision-cut rat lung slices by the intact glucosinolates, glucoraphanin and glucoerucin. Lung Cancer.
- Fahey, J.W., K.L. Wade, K.K. Stephenson and F.E. Chou, 2003. Separation and purification of glucosinolates from crude plant homogenates by high-speed counter-current chromatography. J. Chromatogr. A, 996: 85-93.
- Abdull Razis, A.F., M. Bagatta, G.R. De Nicola, R. Iori and C. Ioannides, 2010. Intact glucosinolates modulate hepatic cytochrome P450 and phase II conjugation activities and may contribute directly to the chemopreventive activity of cruciferous Vegetables. Toxicol., 277: 74-85.

- Anita M. Kore, F. Gayland, A. Spencer and M. Wallig, 1993. Purification of the ω-(Methylsulfinyl)alkyl glucosinolate hydrolysis products: l-Isothiocyanato-3-(Methylsulfinyl) propane,l-Isothiocyanato-4(Methylsulfinyl)butane, 4-(Methylsulfinyl)butanenitrile and 5-(Methylsulfinyl) pentanenitrile from Broccoli. J. Agric. Food Chern., 41: 89-95.
- 12. McNaughton, S.A.G.C.M., 2003. Development of a food composition database for the estimation of dietary intakes of glucosinolates, the biologically active constituents of Cruciferous vegetables British J. Nutrition, 90: 687-697.
- Aires, A., V.R. Mota, M.J. Saavedra, E.A. Rosa and R.N. Bennett, 2009. The antimicrobial effects of glucosinolates and their respective enzymatic hydrolysis products on bacteria isolated from the human intestinal tract. J. Appl. Microbiol., 106: 2086-2095.
- Nebosa Ilic, Jennifer Normanly and J.D. Cohen, 1996.
 Quantification of free plus conjugated Indoleacetic acid in Arabidopsis Requires correction for the non enzymatic conversion of Indole nitriles. Plant Physiol., 111: 781-788.
- 15. Malcolm C. Elliott and B.B. Stowe, 1971. Indole compounds related to auxins and goitrogens of woad (*Isatis tinctoria* L.). Plant Physiol., 47: 366-372.
- Rakariyatham, N., Butr-Indr B Hataichanoke Niamsup Lalida Shank, 2006. Improvement of myrosinase activity of *Aspergillus* sp. NR4617 by Chemical Mutagenesis, 9(4).
- 17. Agnihotri, K.N., 1999. A High-performance liquid chromatographic method for separation and quantification of intact glucosinolates. Chromatographia, 49(5/6).
- 18. Marcelo Fonseca Xavier, Toni Jefferson Lopes, Mara Gabriela Novy Quadri and M.B. Quadri, 2008. Extraction of Red cabbage Anthocyanins: Optimization of the operation conditions of the column process. Brazilian archives of Biology and Technol., 51: 143-152.
- Du, Q., L. Zhang, H. Cui, K. Wang, L. Xiu, et al., 2008. Isolation of glucosinolates from broccoli seeds by use of high-speed countercurrent chromatography. Acta Chromatographica, 203: 489-496.
- Minchinton, I., J. Sang, D. Burke and R.J.W. Truscott, 1982. Separation of desulphoglucosinolates by reversed-phase high-performance liquid chromatography J. Chromatography, 247: 141-148.

- McNaughton, S.A. and G.C. Marks, 2003. Development of a food composition database for the estimation of dietary intakes of glucosinolates, the biologically active constituents of cruciferous vegetables. Br J. Nutr., 90: 687-697.
- 22. Kim, J.H., T.P. Durrett, R.L. Last and G. Jander, 2004. Characterization of the Arabidopsis TU8 glucosinolate mutation, an allele of Terminal Flower2. Plant Mol. Biol., 54: 671-682.
- 23. Zhou, J., J. Hu and A. Qiu, 2005. Separation and purification of the main glucosinolate from rapeseeds. Se. Pu., 23: 411-414.
- 24. Barillari, J., R. Cervellati, M. Paolini, A. Tatibouet, P. Rollin, *et al.*, 2005. Isolation of 4-methylthio-3-butenyl glucosinolate from Raphanus sativus sprouts (*Kaiware daikon*) and its redox properties. J. Agric. Food Chem., 53: 9890-9896.