

Microbiological Study of Periodontitis in the West of Algeria

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Abstract: In pathology, oral flora responsible for periodontal disease is polymorphic. The periodontal infection results either from the penetration of pathogenic microorganisms in the tissues, or even the activation of already existing germs, but not pathogenic under normal conditions. Of a population of 500 patients consulting in Periodontology, 149 were diagnosed as suffering from periodontitis. More than Twenty bacteria had been isolated and identified from subgingival plaques of patients affected with aggressive (28%) and chronic periodontitis (18.4%). In general, the bacterial flora of aggressive periodontitis had shown dominant gram-negative and motile morphotypes, among them, straight rods, curved, spirochetes and fusiform Gram negative. For chronic periodontitis the bacterial flora is characterized by the dominance of rods and motile to the rare gram-negative morphotypes. Certain bacteria known as periodontopathogens were identified in aggressive periodontitis as *Aggregatibacterium actinomycetemcomitans* (7.3%), *Prevotella intermedia* (4.2%), *Eikenella corrodens* (6.3%), *Bacteroides fragilis* (4.6%) and *Capnocytophaga* sp (4.7%).

Key words: Periodontitis disease • *Aggregatibacterium actinomycetemcomitans* • *Prevotella intermedia* • *Eikenella corrodens* • *Bacteroides fragilis* and *Capnocytophaga* sp.

INTRODUCTION

The periodontal infection results either from the penetration of pathogenic microorganisms in the tissues, or even the activation of already existing germs, but not pathogenic under normal conditions. This responsible flora is polymorphic, Gram-negative and microaerophilic or strictly anaerobic. Only ten or twenty species, regarded as pathogens, play a role in the pathogenesis of periodontal destruction [1]. Due to the fact that periodontitis is caused by bacteria, the most important causal periodontal treatment is the elimination of these bacteria [2]. Even on an individual tooth, periodontal disease does not progress uniformly. This can make two Hypotheses: either all tooth surfaces are not vulnerable to the same point, or pathogens are not distributed uniformly over the dental arches [2].

Each type of periodontal disease has a subgingival flora consists of a combination of micro-organisms of its own. The concept of bacterial specificity has been demonstrated that through advances in techniques for anaerobic cultures and the development of new selective culture media. Most

microorganisms involved in these disease are gram-negative bacilli, anaerobes (*Porphyromonas gingivalis*, *Prevotella intermedia*, *Fusobacterium nucleatum*, *Campylobacter rectus*) or capnophiles (*Aggregatibacterium actinomycetemcomitans*, *Eikenella corrodens*, *Capnocytophaga ochracea*...) [3].

The juvenile periodontitis were divided into two clinical entities: localized juvenile periodontitis and generalized juvenile periodontitis, each with a different microbiology. The microbiology of generalized juvenile periodontitis is more complex and association of *Porphyromonas gingivalis* (10 to 15%) and other gram-negative bacilli (*Eikenella corrodens*, *Capnocytophaga* sp. *Aggregatibacterium actinomycetemcomitans* [3].

The rapidly progressive periodontitis is an aggressive form of periodontitis. The subgingival flora is typically composed of significant proportions of *Porphyromonas gingivalis*. *Prevotella intermedia* and other bacteria of the genus *Bacteroides* but *Porphyromonas gingivalis* appears to be one of the essential causative microorganisms in rapidly progressing periodontitis [3].

In recent years researchers in bacteriology trying to identify the causative bacteria.

The aim of our work is to isolated identify the bacterial flora characteristic of aggressive and chronic periodontitis.

MATERIAL AND METHODS

232 patients aged 14 -35 years with periodontitis are presented in the department of Periodontology (Oran, Algeria) and were divided as aggressive or classification of Armitage [4]

Bacteriological Study

This Study Includes Several Steps

Selection of Site: The sampled sites were either molars and incisors, because these areas are most frequently affected in localized juvenile periodontitis.

Sampling: After removal of supragingival plaque by means of sterile cotton balls, subgingival plaque was collected on Gracey curette inserted to the depth of the periodontal (Figure 1). After sampling, samples of subgingival plaque were collected and deposited on tubes containing sterile saline or otherwise sterile distilled water [5]

Culture: the sample plate and collected shall be deposited in a tube containing 2 ml of saline (0.9% NaCl). The samples were mixed using a vortex for one minute to allow the dispersal of bacteria. Samples of subgingival plaque were shaken vigorously for 30 seconds using the mixer before being prepared for analysis.

Two other tests will be made for microscopic examination (one for fresh and one for Gram stain) [6].

The streaking were made on different selective agar media "trypticase soy agar with serum horse + bacitracin and vancomycin, TSBV) [7], *Bacteroides* bile esculin agar [8] and nonselective aerobic and anaerobic (Brucella agar and blood agar) and identify the bacteria found in these different clinical entities [9].



Fig. 1: subgingival sampling, CHU Oran.

After the bacteria are inoculated on solid media, Petri dishes were incubated under different atmospheres (ambient, microaerophilic and anaerobic) at 37°C for 2 to 7 days depending on the used media [6].

RESULTS

The study is based on results from a series of samples of subgingival plaque collected and treated with oral bacteriology laboratory, department of periodontology, Oran Algeria. 149 patients aged 14 - 35 years were included in this study.

Direct examination represented by fresh and Gram stain revealed in patients with aggressive periodontitis, a predominance of flora especially curved, fusiform motile rods and motile spirochetes and Gram negative bacilli and coccobacilli Gram negative and Gram-positive filaments (Figure2).

For chronic periodontitis, the flora is characterized by both motile and non motile morphology Gram negative and positive. The morphology of the most evocative are cocci and straight rods (Figure 3). The rate of coccobacilli is very negligible in this entity.

bacteria were isolated, some are identified as periodontal pathogens and others whose etiologic role in periodontal disease is not yet clarified. These bacteria are *Aggregatibacterium actinomycetemcomitans*, *Eikenella corrodens* *P. intermedia*, *Capnocytophaga*, *Campylobacter*, *H. pylori*, *Actinomyces*, *Eubacteriu* and *Peptostreptococcus* (Table 1).

Regarding aggressive periodontitis, bacteria that play a role in periodontal disease are *A. actinomycetemcomitans*, *A. actinomycetemcomitans*, *E. corrodens*, *P. intermdia*, *Capnocytophaga* sp. and *Bacteroides frgilis*.

Other bacteria have been identified including their involvement in periodontal disease is unclear such as: *Haemophilus aphrophilus*, *Campylobacter* species, *Eubacterium* and *Actinomyces*.

The enterobacteria were also identified in our study, such as *E.coli*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* For chronic periodontitis, the absence of *Bacteroides fragilis*, *Capnocytophaga* sp as important bacteria in periodontal disease is noted in this entity (Table 1).

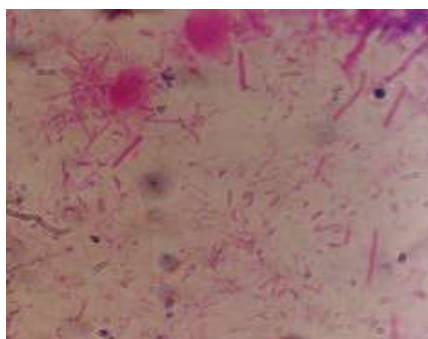


Fig. 2: Flora of aggressive periodontitis X1000.

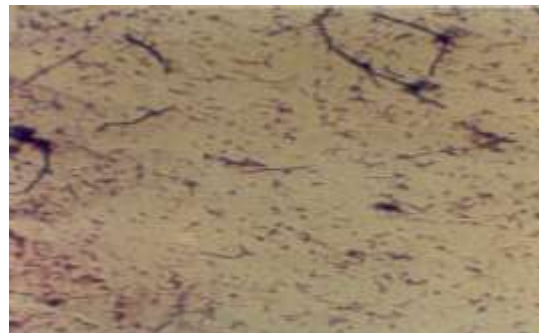


Fig. 3: Flora of chronic periodontitis X1000.

Table 1: cultivable flora of periodontitis

Bacteria	Agressive periodontitis (%)	Chronic periodontitis(%)
<i>A. actinomycetemcomitans</i>	8.4	0.3
<i>Eikenella corrodens</i>	10.8	20.7
<i>Prevotella intermedia</i>	8.0	5.4
<i>Bacteroides fragilis</i>	5.6	0.0
<i>Capnocytophaga</i> sp	2.4	0.0
<i>Campylobacter</i> sp	7.1	7.5
<i>Helicobacter pylori</i>	4.5	6.3
<i>Haemophilus aphrophilus</i>	7.1	2.1
<i>Eubacterium</i> sp	8.0	6.3
<i>Eubacterium nodatum</i>	1.4	0.0
<i>Actinomyces israelii</i>	1.7	1.8
<i>Actinomyces naeslundii</i>	1.7	0.9
<i>Peptostreptococcus anaerobius</i>	1.4	0.0
<i>Peptostreptococcus micros</i>	3.5	0.3
<i>Staphylococcus aureus</i>	4.2	0.0
<i>Streptococcus viridans</i>	11.8	38.7
<i>E. coli</i>	3.1	5.4
<i>Pseudomonas aeruginosa</i>	3.5	0.0
<i>Klebsiella pneumoniae</i>	3.1	0.0
<i>Corynebacterium matruchotii</i>	2.4	1.2
<i>Neisseria mucosa</i>	0.3	3.0

DISCUSSION

Oral microbiology did in the second half of the twentieth century considerable progress. The microbiological examination is recognized as indispensable in certain situations or to detect and quantify the presence of certain pathogenic bacteria that are suspected.

The flora of periodontal pockets are characterized by a high proportion of motile anaerobic Gram-negative bacteria (30%) and spirochetes [10]. In our study we found also a dominance of Gram negative -flora, motile as well as the high density of spirochetes.

Some Gram-negative anaerobes are associated with the incidence and progression of periodontal disease [11-13].

The Spirochetes were observed in all cases and play an important role in periodontitis [14]. Oral spirochetes are classified as periodontal pathogens [15]. Also, Loesche and Grossman[16] support this hypothesis. Other studies suggest that the predominance of *Treponema denticola* and other gram-negative bacteria in high numbers in periodontal pockets may play an important role in progression of disease [17, 18].

According, Loesche and Grossman [16] the oral spirochetes are often dominant bacterial types observed in subgingival plaque taken from sites with periodontal disease [19]. For Richard *et al.* spirochetes comprise a significant proportion (20-50%) of the total microscopic count of bacteria in inflamed pockets [15]. Also, Baehni and Guggenheim [20] have found spirochetes in periodontal pockets of patients with periodontitis. Sela has shown a positive relationship

between *Treponema denticola* and aggressive periodontitis [17].

Our study revealed that chronic periodontitis flora is diverse and composed of both motile and non motile types and gram positive and negative (cocci, rod-rights) as well as its rarity in gram-negative bacilli and gram-positive filaments.

We have found *Eikenella corrodens* in chronic (20.7%) and in aggressive periodontitis (10.8%). For Noiri *et al*, *Eikenella corrodens* is associated with various types of periodontitis [21]. Similarly, Nonnenmacher *et al*. [22] and the Academy report [14] *Eikenella corrodens* were detected in both groups.

The ecological niche of *Aggregatibacterium actinomycetemcomitans* is the oral cavity. It is mainly found in the subgingival flora [23].

Aggregatibacterium actinomycetemcomitans is detected in our study more in aggressive periodontitis (8.4%) compared to chronic periodontitis (0.3%); *Aggregatibacterium actinomycetemcomitans* has been implicated as primary causative agent in localized juvenile periodontitis [10, 3, 24]. Nonnenmacher *et al*. [22] have identified *Aggregatibacterium actinomycetemcomitans* in cases of localized juvenile periodontitis and rapidly progressing. The Academy report [14] was associated *Aggregatibacterium actinomycetemcomitans* cases of localized juvenile periodontitis and generalized. According to Loesche and Grossman, [16] *Aggregatibacterium actinomycetemcomitans* is present in all clinical entities. Baehni and Guggenheim [20] have reported its presence in subgingival sites of patients with periodontitis.

We identified *Capnocytophaga* only in aggressive periodontitis. According to Spratt *et al*. [27], *Capnocytophaga* sp. has been shown his involvement in some forms of periodontitis.

We found *Prevotella intermedia* and *Haemophilus aphrophilus* more in aggressive than in chronic periodontitis. According Gürsoy *et al*. [25], *Prevotella intermedia* is associated with periodontal disease. For Mayorga-Fayad *et al*. [26], the frequency increases *Prevotella intermedia* in patients with periodontitis. According Tempio and Slots [28] *Haemophilus aphrophilus* is present in low proportion in the subgingival plaques and play a role that are not clear in advanced periodontal disease. For the same researchers, Dental plaque is a primary ecological niche for *Haemophilus aphrophilus*.

In our study, *Eubacterium* is present in aggressive periodontitis (8%). Indeed, Poco and al. [29], have found

up to 54% of the microflora of subgingival plaque from periodontal pockets. For Hill *et al*. [30], this bacterium was isolated between 35 and 42%.

According Ximenez-Fyvie *et al*. [31] *Actinomyces* are an important component of supra and subgingival plaques. Cisar *et al*. [32] have found *Actinomyces naeslundii* in dental plaque may contribute to certain diseases such as periodontitis. Johnson *et al*. [33] noted *Actinomyces israelii* in periodontitis cases. We isolated *Actinomyces naeslundii* (1.7%) and *Actinomyces israelii* (1.7%) in aggressive periodontitis, *Actinomyces naeslundii* (0.9%) and *Actinomyces israelii* (1.8%) in chronic periodontitis.

We isolated *Staphylococcus aureus* (4.2%) and *Peptostreptococcus anaerobius* (1.4%) from the periodontal pockets of patients with aggressive periodontitis. Other studies have reported the presence of *Staphylococcus aureus* in such infections [14]. Nonnenmacher *et al*. [22], *Staphylococcus aureus* were observed in cases of periodontitis. Kumar [34] *et al*. Have reported that *Peptostreptococcus* is associated with periodontitis and the percentage of this organism is very high in these entities. Similarly, Mouton [35] attributed a causative role in *Peptostreptococcus* (*Peptostreptococcus anaerobius* and *Peptostreptococcus micros*) in cases of periodontal disease.

We noted the presence of *Campylobacter* in both types of periodontitis. Macuchi and Tanner [36], have identified at least seven species of *Campylobacter* in subgingival sites. According to Lakshman [37], *Campylobacter* spp. is associated with different forms of periodontitis.

The *Neisseria* were isolated with very low levels in aggressive periodontitis (0.3%). For the expert group - INSERM-[23], *Neisseria* are primary colonizers of the film but gained no exogenous pathogenic role they have not been assigned.

Streptococci were observed with high percentage in chronic periodontitis (38.7%). For Baehni and Guggenheim, [20], *Streptococci* are considered beneficial for the host and colonizing the pocket in large numbers, can delay the process of periodontal disease.

Helicobacter pylori is grown in chronic periodontitis (6.3%) more than aggressive periodontitis (4.5%). For Stolzenberg-Solomon *et al*. [38] *Helicobacter pylori* was found in dental plaque and associated with periodontal disease. According Namiot *et al*. [39] Studies have shown that *Helicobacter pylori* colonizes not only the stomach but also the oral cavity and there are strains of *Helicobacter pylori* that were isolated from dental plaque.

Stolzenberg-Solomon *et al.* [38] have shown that periodontal disease and especially the deep periodontal pockets associated with *Helicobacter pylori*.

E. coli was grown in both entities. According Betancourth *et al.* [40] micro-organisms of the family Enterobacteriaceae (gram negative rods) were considered as microorganisms unusual in patients with periodontitis. Steffens *et al.* [41] have mentioned that there are several studies that have associated enteric bacilli to periodontal disease.

Corynebacterium matruchotii is found in the case of aggressive (2.4%) and chronic (1.2%) periodontitis. Barrett *et al.* [42] have reported the possible role of *Corynebacterium matruchotii* in dental plaque and the pathogenesis of periodontitis.

REFERENCES

1. Darby, I.M. Curtis, 2001. Microbiology of periodontal disease in children and young adults. *Periodontology*, 26: 33-53.
2. Macuchi, P.J. and A.C.R. Tanner, 2000. *Campylobacter* species in health, gingivitis and periodontitis. *J. Dent Res.*, 79(2): 785-792.
3. Fine, D.H., D. Furgang, H.C. Schreiner, P. Goncharoff, J. Charlesworth, G. Ghazwan, P. Fitzgerald-Bocarsly and D.H. Figurski, 1999. Phenotypic variation in *Actinobacillus actinomycetemcomitans* during laboratory growth; implication for virulence. *Microbiology*, 145: 1335-1347.
4. Armitage C. Gary, 1999. Development of a Classification System for Periodontal Diseases and Conditions. *Ann. Periodontol.*, 4: 1-6.
5. Wu, Y., J. Yan, L. Chen and Z. GU, 2007. Association between infection of different strains of *Porphyromonas gingivalis* and *Actinobacillus actinomycetemcomitans* in subgingival plaque and clinical parameters in chronic periodontitis / J. Zhejiang Univ. Sci. B., 8(2): 121-131.
6. Lakhssassi, N. and M. Sixou, 2005. Variabilité de l'efficacité de l'érythromycine et de la spiramycine sur les pathogènes parodontaux dans les parodontites agressives. Étude in vitro comparative. *Pathologie Biologie*, 53: 527-535.
7. Bonta, Y., J. Zambon, R.J. Genco and M.E. neiders, 1985. Rapid Identification of Periodontal Pathogens in Subgingival Plaque: Comparison of Indirect Immunofluorescence Microscopy with Bacterial Culture for Detection of *Actinobacillus actinomycetemcomitans*. *J. Dent Res.*, 64(5): 793-798.
8. Winn, W.C. and E.W. Koneman, 2006. The anaerobic bacteria. In: Koneman's color atlas and textbook of diagnostic microbiology. Sixth Edition.
9. François Denis and Marie-Cécile Ploy, 2007. *Bactériologie médicale. Technique usuelle*.
10. Ouhayoun, J.P., D. Etienne and F. Mora., 0000. UFR d'odontologie. Sous-section de parodontologie.
11. Noiri, Y., L. Li and S. Ebisu, 2001. The localization of periodontal-disease-associated bacteria in human periodontal pockets. *J. Dent Res.*, 80(10): 1930-4.
12. Holt, S.C. and T.E. Bramanti, 1991. Factors in virulence expression and their role in periodontal disease pathogenesis. *Crit. Rev. Oral. Biol. Med.*, 2(2): 177-281.
13. Loos, B.G., D. Mayrand, R.J. Genco and D.P. Dickinson, 1990. Genetic heterogeneity of *Porphyromonas (Bacteroides) gingivalis* by genomic DNA fingerprinting. *Dent Res.*, 69(8): 1488-93.
14. Academy Report, 2005. The Role of Supra- and Subgingival Irrigation in the Treatment of Periodontal Diseases. *J. Periodontol.*, 76: 2015-2027.
15. Ellen, R.P. and V.B. Galimanas, 2005. Spirochetes at the forefront of periodontal infections. *Periodontol.*, 38: 13-32.
16. Loesche, W.J. and N.S. Grossman, 2001. Periodontal disease as a specific, albeit chronic, infection: diagnosis and treatment. *Clin. Microbiol., Rev.*, 14(4): 727-52.
17. Sela, M.N., 2001. Role of *Treponema denticola* in periodontal diseases. *Crit Rev. Oral. Biol. Med.*, 12(5): 399-413.
18. Holt, S.C. and T.E. Bramanti, 1991. Factors in virulence expression and their role in periodontal disease pathogenesis. *Crit Rev. Oral. Biol. Med.*, 2(2): 177-281.
19. Loesche, W.J., 1988. The role of spirochetes in periodontal disease. *Adv. Dent. Res. Nov.*, 2(2): 275-83.
20. Baehni, P.C. and B. Guggenheim, 1996. Potential of diagnostic microbiology for treatment and prognosis of dental caries and periodontal diseases. *Crit. Rev. Oral. Biol. Med.*, 7(3): 259-77.
21. Noiri, Y., L. Li and S. Ebisu, 2001. The Localization of Periodontal disease- associated Bacteria in Human Periodontal Pockets. *J. Dent Res.*, 80(10): 1930-1934.
22. Nonnenmacher, C., R. Muters and L. Flores de Jacoby, 2001. Microbiological characteristics of subgingival microbiota in adult periodontitis, localized juvenile periodontitis and rapidly progressive periodontitis subjects. *Clin. Microbiol., Infect.*, 7: 213-217.

23. INSERM - Expertise Collective, 1999.
24. Slots, J. And R.J. Genco, 1984. Black-pigmented *Bacteroides* species, *Capnocytophaga* species and *Actinobacillus actinomycetemcomitans* in Human Periodontal Disease: virulence Factors in Colonization, Survival and Tissue Destruction. J. Dent Res., 63(3): 412-421, March, 1984.
25. Gürsoy, M., G. Haraldsson, M. Hyvönen, T. Sorsa, R. Pajukanta and E. Könönen, 2009. Does the frequency of *Prevotella intermedia* increase during pregnancy? Oral. Microbiol. Immunol., 24(4): 299-303.
26. Mayorga-Fayad, I., G.I. Lafaurie, A. Contreras, D.M. Castillo, A. Barón and R. Aya Mdel, 2007. Subgingival microbiota in chronic and aggressive periodontitis in Bogotá, Colombia: an epidemiological approach Biomedica, 27(1): 21-33.
27. Spratt, D.A., J. Greenman and A.G. Schaffer, 1996. *Capnocytophaga gingivalis*: effects of glucose concentration on growth and hydrolytic enzyme production. Microbiol., 142(Pt 8): 2161-4.
28. Temprow, P.J. and J. Slots, 1986. Selective medium for isolation of *H. aphrophilus* from human periodontium and others sites and the low proportion of the organism in the oral flora. J. Clin. Microbiol., 23(4): 777-782.
29. Poco, S.E. Jr, F. Nakazawa, M. Sato and E. Hoshino, 1996. *Eubacterium minutum* sp. nov., isolated from human periodontal pockets. Int. J. Syst. Bacteriol., 46(1): 31-4.
30. Hill, G.B., O.M. Ayers and A.P. Kohan, 1987. Characteristics and sites of Infection of *Eubacterium nodatum*, *Eubacterium timidum*, *Eubacterium brachy* and Other Asaccharolytic Eubacteria. J. Clin. Microbiol., 25(8): 1540-1545.
31. Ximenez-Fyvie, L.A., A.D. Haffajee and S.S. Socransky, 2000. Microbial composition of supra and subgingival plaque in subjects with adults periodontitis. J. Clin. Periodontol., 27: 722-732.
32. Cisar, J.O., P.E. Kolenbrander and F.C. McIntire, 1989. Specificity of coaggregation reactions between human oral streptococci and strains of *Actinomyces viscosus* or *Actinomyces naeslundii*. Infect Immun., 24(3): 742-52.
33. Johnson, J.L., L.V. Moore, B. Kaneko and W.E. Moore, 1990. *Actinomyces georgiae* sp. nov., *Actinomyces gerencseriae* sp. nov., designation of two genospecies of *Actinomyces naeslundii* and inclusion of *A. naeslundii* serotypes II and III and *Actinomyces viscosus* serotype II in *A. naeslundii* genospecies 2. Int. J. Syst. Bacteriol., 40(3): 273-86.
34. Kumar, P.S., A.L. Griffen, M.L. Moeschberger and E.J. Leys, 2005. Identification of candidate periodontal pathogens and beneficial species by quantitative 16S clonal analysis. J. Clin. Microbiol., 43(8): 3944-55.
35. Mouton, C., 2003. Bactériologie et pathogénie des maladies parodontales. In: parodontologie du diagnostic à la pratique. Pierre Bercy, Henri Tenenbaum. 1th.
36. Macuchi, P.J. and A.C.R. Tanner, 2000. *Campylobacter* species in health, gingivitis and periodontitis. J. Dent Res., 79(2): 785-792.
37. Lakshman, S.P., 2006. Oral microbiology. In: Essential microbiology for dentistry. 3th edition.
38. Stolzenberg-Solomon, R.Z., K.W. Dodd, M.J. Blaser, J. Virtamo, P.R. Taylor and D. Albanes, 2003. Tooth loss, pancreatic cancer and *Helicobacter pylori*. Am. J. Clin. Nutr., 78(1): 176-81.
39. Namiot, D.B., Z. Namiot, A. Kemonia and M. Goebiewska, 2006. Peptic ulcers and oral health status. Adv. Med. Sci., 51: 153-5.
40. Betancourth, M., R. Arce, J. Botero, A. Jaramillo, C. Cruz and A. Contreras, 2006. Unusual microorganisms in gingival sulcus and periodontal pockets. Colombia Médica Vol. 37 N° 1.
41. Steffens, N.S., A.J. Gamonal and R.M. Gajardo, 2006. Occurrence of non fastidious Gram negative rods and yeasts in the subgingival microbiota of Chilean periodontitis patients. Original title: Ocurrencia de bacilos Gram negativos no fastidiosos y levaduras en la microbiota subgingival de pacientes chilenos con periodontitis Rev. Odont. Mex., 10(3): 119-125.
42. Barrett, S.L., B.T. Cookson, L.C. Carlson, K.A. Bernard and M.B. Coyle, 2001. Diversity within reference strains of *Corynebacterium matruchotii* includes *Corynebacterium durum* and a novel organism. J. Clin. Microbiol., 39(3): 943-8.