

Scope of Nasal Drug Delivery: Current Challenges

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Abstract: Currently the pharmaceutical scientists has exploring the possibilities of intranasal delivery as an alternative to other routes in the view of increasing number of reports related to problems associated with oral, parenteral and rectal routes for therapeutic delivery of different drugs. The present attempt was focused on nasal drug delivery systems for different therapeutic potentials i.e. local, systemic, CNS delivery and vaccine delivery. The factors which affect the absorption of drugs or biomolecules through the nasal mucosa including biological, device-related and formulation factors are also discussed. This review may provide a complete and deep insight in relation to the nanotechnology based delivery systems for intranasal administration. A comparative study of conventional and novel delivery systems with respect to their advantages and shortcoming allows the in-depth information over the advancement of nasal delivery systems for different therapeutic purposes. The emphasis on current challenges and the possible solutions against them makes this attempt valuable for the researchers working on this platform.

Key words: Intranasal Drug Delivery • Nanoparticulate • Nasal Vaccination • Nose to Brain Targeting

INTRODUCTION

From the past few decades, much interest has been generated to the exploitation of the nasal route for systemic delivery of drugs to the specific site [1, 2]. Nasal drug delivery (NDD) conventionally has been used for treatment of local diseases such as nasal allergy, congestion and infections, respectively [3]. Besides it the drug can be delivered directly to the brain along the olfactory nerves [4]. Traditionally Peptide hormones (PHs) were administered by the parenteral route, like intramuscular (IM) or intravenous (IV) and subcutaneous injection (SC) [5]. But the Intranasal (IN) approach offers as an alternative route of administration for peptide hormones, like glucagon [6]. In order to improve the bioavailability of nasally administered drugs, nanoparticulate (NP) system has been explored in combination to nasal delivery [7]. NP drug delivery systems may enhance notably the transport of drugs (And vaccines) across the nasal mucosa [8]. The usage of nanoparticles for vaccine delivery provides worthy effect, by achieving satisfying immune responses [7]. The main advantages of IN delivery are

avoidance of first pass metabolism, lowered systemic exposure to drug and quick onset of action [9]. The nasal mucosa has been examined as a possible route of administration to accomplish a faster and surpass level of drug absorption [10]. Kaur investigated that when the drug administered intranasally, vaccines can vitalize both local and systemic immune responses, respectively [9]. Intranasal route along with formulation of biodegradable polymers, which are lipophilic by nature and nanometer range of particle size are capable to reach CNS at therapeutically effective dose [11]. A research established that there exists an undeviating anatomical connection between the nasal cavity and the CNS via paracellular and transcellular pathways as well as through trigeminal neurons which guides towards the development of CNS therapeutics for IN (intranasal) administration [12]. From such positive attributes, it's been sensible to appraise IN administration while developing the new therapeutics, for different possibilities (Fig. 1). The present attempt provides an overall development hierarchy of nasal administration of therapeutic along with the challenges and future possibilities.

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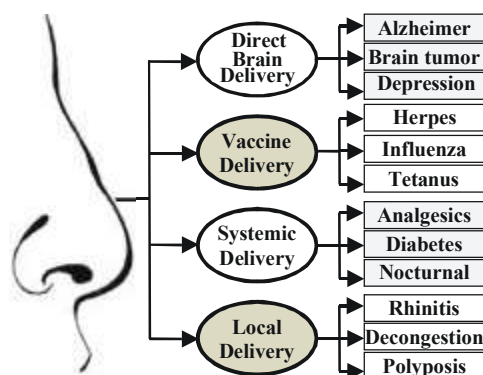


Fig. 1: Scope of nasal drug delivery[13]

Table 1: Marketed nasal products for topical/local delivery.

Product name	Drug	Indication	Manufacturer	References
Allergo-Comod	Cromolin sodium	Allergic rhinitis	Ursapharm	[16]
Lomusol®	Cromolyn sodium	Allergic rhinitis	Sanofi Aventis	[17]
Vividrin	Cromolyn sodium	Allergic rhinitis	Bausch and Lomb	[18]
Astelin®	Azelastine	Allergic rhinitis	Meda Pharm	[19]
Bactroban	Mupirocin	Eradication of nasal staphylococci	GlaxoSmithKline	[20]
Beconase	Beclomethasone dipropionate	Management of seasonal and perennial (Allergic) rhinitis	GlaxoSmithKline	[21]
Decadron*	Dexamethasone	Treatment of inflammatory nasal conditions or nasal polyposis	Merck and Co., Inc.	[22]
Flixonase	Fluticasone propionate	Management of seasonal and perennial (Allergic) rhinitis	GlaxoSmithKline	[23]
Livocab	Levocabastine	Allergic rhinitis	Janssen	[24]
Nasalchrom	Sodium cromoglicate	Management/treatment of symptoms of seasonal and perennial rhinitis	Mc Neil Consumer Healthcare	[25]
Nasivin	Oxymetazoline	Temporary relief of nasal congestion	Braco	[26]
Nasonex	Mometasone furoate	Management of seasonal and perennial (Allergic) rhinitis	Merck and Co., Inc.	[27]
Otrivin	Xylometazoline	Temporary relief of nasal congestion	Novartis	[28]
Patanase	Olapatadine	Management/treatment of symptoms of seasonal and perennial rhinitis	Alcon Laboratories, Inc.	[29]
Rhinex	Naphazoline	Decongestion	Stada	[30]
Rhinocort	Budesonide	Management of seasonal and perennial (Allergic) rhinitis	AstraZeneca	[31]
Syntaris	Flunisolide	Management of seasonal and perennial (Allergic) rhinitis	Piramal Healthcare	[32]

* Brand Discontinued in US

Nasal Route for Local/Topical Delivery: Protruding example for local/topical treatment by intranasally administered drugs are topical decongestants and topical steroids. At present it get grip of approx 2/3rd of the total market value of intranasal products [13]. As they are generally used for allergic rhinitis and as an antihistamines. IN administration is a logical delivery choice for the local/topical treatment of nasal symptoms, due to the fact that comparably low doses are efficient on topical administration with less systemic toxic effects. Factor that make an IN administration of antihistamines and corticosteroids more attractive as well as regularly preferred route of administration is that antihistamines do not cause impairment of psychomotor function or significant sedation as well. The clinical outcome of steroids is often objectionable, notably due to the poor

distribution to the various regions of nose, respectively. Hence, as an effect, the nanotechnology based treatment methods for chronic sinusitis and rhinitis, have the possible market hike for the new as well as the existing topical agents, which enhanced patient compliance and bioavailability. Table 1 gives an overview of some of the marketed nasal products for local delivery [14, 15].

Nasal Route for Systemic Delivery: It seems systemic availability of drugs via intranasal administration is an effective way as compared to oral and intravascular routes [14]. A study showed an alternative routes of administration of peptide hormones as a promising approach via intranasal (IN) route for the treatment of severe hypoglycemia as it provides the potential for rapid absorption and quick onset of action, although avoiding

hepatic first pass metabolism [7]. A new IN glucagon product AMG504-1 (Locemia Solutions, Montreal, Canada) currently being in clinical studies under the investigation process. AMG504-1 model, a glucagon delivery system consists of a dry powder glucagon formulation in compact, portable, single-use nasal powder dosing device that permits single step administration. The formulation contains glucagon along with phospholipid as absorption enhancing agent and a cyclodextrin as a bulking agent as well. Unlike insulin that is routinely injected by the individual with diabetes, glucagon is administered by an individual who never be a trained medical professional (e.g., child, spouse, friend, work colleague, sports coach, etc.) [6].

Nasal Route for Brain Delivery: IN delivery of drugs, targeting the CNS is an area of increasing interest where a rapid and/or specific targeting is achieved, as it can circumvent the BBB and delivers the therapeutic molecules to the brain. This route has emerged as a promising approach for the delivery of drugs to the brain [33]. It explores the possibility of direct nose to brain transport of administered drugs which are difficult to pass through BBB, such as peptides, proteins and hydrophilic small molecules and improve their access to the various regions of brain [34]. The management of condition such as Alzheimer's disease, Parkinson's disease, brain tumors, epilepsy and sleep disorders would be benefited from the development of IN delivery system, whose improved access to the brain region provides the opportunity of optimizing the efficacy of treatment, such as meningitis and stroke [35]. Studies on nose-to-brain transport shows that drug reached the CNS after a nasal instillation mainly through three pathways, via; (i) absorption across the nasal respiratory epithelium into the systemic circulation and from there, across the BBB into the brain (Systemic pathway), (ii) direct paracellular or transcellular transport via the olfactory neurons (Olfactory neural pathway) or the olfactory epithelial cells (Olfactory epithelial pathway), or (iii) transport via the trigeminal nerves (Trigeminal pathway) [36]. When drugs are administered nasally for the direct transfer from nasal crater to brain, the exact path it follows is still under an investigation [37].

Nasal Route for Vaccine Delivery: At present, IN delivery of vaccines has received some more attention and become popular area of research. It offers many distinguished advantages over parental route of administration by preventing infection with pathogens. IN immunization has reported to produce local as well as systemic immunity.

Furthermore, IN vaccination does not require sterile dosing technique or a sterile product [38]. Nasal mucosa as a site of vaccine administration is extremely rich in organized Lymphatic tissues as well as in specialized cells. In humans, immune responses from nasal mucosal surface are generally simulated through interactions initiated with nasopharyngeal associated lymphoid tissues (NALT) also known as Waldeyer's ring and these organized lymphatic tissues are concerned with the first line defense against airborne microorganisms. The majority of the invading infectious pathogens gets entry into the body via mucosal surfaces and the mucosa represent the first line body defenses against infection [13]. Nasal secretions are known to contain immunoglobulins (IgA, IgG, IgM&IgE), protective proteins such as complement as well as neutrophils and lymphocytes in the mucosa [15]. Examples of the human efficacy of intranasal vaccines include those against influenza A & B virus, proteosoma influenza, adenovirus vectored influenza, Group B meningococcal native, attenuated respiratory syncytial virus and Para influenza O3 viruses [39].

Following properties reveals the potential of vaccination method [13, 15].

- Economic, patient friendly, non-injectable and easily accessible.
- Highly vascularized.
- Availability of both mucosal and systemic immune responses.
- Doesn't require needles and syringes (a distinct advantage).
- Existence of numerous microvilli covers the nasal epithelium provides a large absorption surface area.

The nasal cavity itself stimulates the local secretory IgA as well as IgG antibodies which provides an additional first line of defense, which eliminates the pathogen before its establishment.

Nanoparticulates for Intranasal Delivery: The incorporation of drugs into the nanoparticles is a promising approach, since these colloidal formulations facilitate their transport across the mucosal barriers and protect them from the degrading milieu in the nasal cavity as well [7]. The use NP provides all sort of beneficial effects for IN delivery. The outcomes of these nanoparticulates-based systems have been intensely promising when combined with the nasal delivery. However it is not clear how this system enhance drug absorption and may cause cellular and subcellular damage

also to the biological system as well [13]. Such NP's incorporated in the form of Nanoemulsions, Nanogels, Liposomes, Carbon nanotubes, Microspheres and Quantum dots.

Nanoparticulates for Nose-To-Brain Delivery: The developed carboplatin-loaded PCL NP (Polycaprolactone Based Nanoparticles) were prepared by double emulsion solvent evaporation technique and characterized by entrapment efficiency, particle size, zeta potential, scanning electron microscopy and DSC (Differential scanning calorimetry), has improved *in vitro* anti-tumour activity as comparison to that of plain drug against human glioblastoma cells LN229 and thus for improved brain delivery, intranasal administration of carboplatin can be used, respectively [12].

The Donepezil loaded nanosuspension for direct olfactory administration was developed, which reached the brain and determined the safety profile in Sprague-Dawley rats. The developed nanosuspension was prepared by ionic cross-linking method and instilled intranasally into the nostrils of rats and drug reached into the brain via nose-to-brain pathway directly. The average size of NS was 150-200nm with a polydispersity index of 0.341 and using HPLC method its concentration was estimated in the brain homogenate. And after the study, it was anticipated that the donepezil loaded NS was capable of giving direct nose-to-brain delivery and eventually enhancing drug concentration in the brain [40]. Bian *et al.*, established with his work that M-C-PLA-NP is a novel brain-targeting agent for NDDS. The developed M-C-PLA-NP exhibited sustained release, smaller particle size and absolute zeta potential along with the high entrapment. M-C-PLA-NP showed its stability in the nasal cavity environment, with no significant change for 2 hrs during a period of incubation with the Lysozyme. And eventually from the in-vivo studies, it was indicated that, the M-C-PLA-NP on IN administration, not only provide a higher brain concentration of aniracetam but also lessen the spreading of the M-C-PLA-NP in other tissues as compared with the free aniracetam [41]. Jafarieh *et al.* [42] using polymeric NP (Nanoparticulate) developed and investigated the possibilities of targeting an Anti-parkinson's drug ropinirole (RH), which was loaded with chitosan NP (CSNPs), prepared by an ionic gelatin method. The RH-CSNPs were characterized for entrapment efficiency *in-vitro* release study, *in-vivo* distribution, loading capacity, particle size, polydispersity-index (PDI) and zeta-potential after nasal administration for brain-targeting. The study showed,

upto 18hrs sustained release profiles and eventually it concluded from the animal studies that novel formulation of RH loaded CSNPs showed superiority of nose-to-brain delivery than that delivered by RH solution with a consequent increase in bioavailability. Jain *et al.* developed and investigated artemether-loaded nanostructured lipid carriers (ARM-NLC) using central composite design for IN delivery. Microemulsion method was applied in preparation of ARM-NLC with optimized formulation having zeta-potential of -34.4mV and particle size of 123.4nm, respectively. As, solid and liquid lipid materials of lipid NP, Trimyristin™ and medium chain triglycerides (MCT) were chosen. The developed formulations were non-toxic and conformed by *in-vitro* cytotoxicity assay using nasal histopathological studies and SVG p12 cell line on sheep nasal mucosa and the developed formulation showed sustained release up to 96hrs and ex-vivo diffusion study revealed that ARM-NLC had significantly lower flux as compared to ARM-SOL. Thus the brain uptake studies and pharmacokinetic in rats showed significant higher concentration of drug upon administration of NLC by IN route in the brain was maintained up to 6hrs owing to slower release of drug [43]. Zhao *et al.* [44] developed IN phospholipid-based gelatin NP encapsulating fibroblast growth factor to the target the brain. Treatment effect assessed by quantifying rational behavior, along with monoamine neurotransmitter levels. Water-in-water emulsion method, after that, freeze dried method were used in the preparation of gelatin nanostructured lipid carrier (GNLs). As comparison with the gelatin nanoparticle (GNs), GNLs possessed better profile, with the zeta potential -38.2 ± 1.2 mv and particle size 143 ± 1.14 nm. Thus, GNLs showed obvious therapeutic effects on hemiparkinsonian rats and no adverse impact on the integrity of nasal mucosa and these are efficiently enriched exogenous bFGF in striatum and olfactory bulb respectively. Table 2 gives an overview of some of the brain targeted intranasal formulations.

Nanoparticulates for Gene Delivery: BT Harmon *et al.*, showed in their work that, IN delivery of unimolecularly compacted DNA NPs (DNA nanoparticles), successfully transfect cells in the rat brain, which consists of single molecules of plasmid DNA enhanced green fluorescent protein (eGFP) which compacted with 10kDa PEG (Polyethylene glycol) substituted Lysine 30mers (Ck30PEG10k). After 2-days of pCG (Plasmid) DNA NP injection, first experiment were conducted to conform whether PCG NPs were successfully transfect cells or not along with eGFP expression in the rat brain, respectively.

Table 2: Various brain-targeted intranasal formulations

Drug	Indication	Category	Formulations	References
Amiloride	Antiepileptic	Diuretics	Nanoemulsion	[45]
Bromocriptine	Parkinson's diseases	Dopamine D2 agonist	Chitosan-loaded nanoparticles CS-BRC-NPs	[46]
Buspirone HCL	Anxiety	Anxiolytic agent	Bus-chitosan nanoparticles	[46]
Clonazepam	Status epileptics	Benzodiazepine derivative	Mucoadhesive microemulsion	[46]
Deferoxamine	Cerebral ischemia	High-affinity iron chelator	Nasal solution	[46]
Didanosine	HIV infection	HIV reverse transcriptase inhibitor	Chitosan-loaded nanoparticles	[46]
Doxorubicin	Antitumour	Anthracycline antibiotics having antitumour Activity	Niosomes	[47]
Duloxetine	Depression	Serotonin and norepinephrine reuptake inhibitor (SNRI)	Nanostructured lipid carrier (NLC)	[46]
Erythropoietin	Cerebral ischemic	Hematopoietic Growth Factor	Nasal solution	[46]
Estradiol	Alzheimer's diseases	Steroid hormone	E2-loaded chitosan nanoparticles	[46]
Hexarelin	Stimulate GH secretion	Growth hormone-releasing neuropeptide	Nasal solution	[46]
Interferon-Beta	Multiple sclerosis	Anti-inflammatory cytokines	Nasal solution	[46]
Lorazepam	Insomnia, anxiety and epilepsy	General anaesthetics slower acting drugs	Microemulsion Gel	[48]
Melatonin	Sleep disorders	Sedative-Hypnotics	Gel suspension	[49]
Methylprednisolone	Multiple sclerosis (MS)	Corticoids	Liposomes	[50]
Neurotoxin-1	Movement disorders	Analgesic peptide	Nanoparticles	[51]
Odorranalectin	Parkinson's disease	Smallest peptide with lectin-like activity	Cubosomes	[52]
			Nanoparticles	[53]
Olanzapine	Schizophrenia	Second-generation or atypical antipsychotic	Transfersomes	[54]
			Nanoparticles	[55]
Ondansetron HCl	Management of chemotherapy induced postoperative nausea and vomiting	Serotonin (5-hydroxy tryptamine) subtype (5HT3) receptor Antagonist	Nanoparticles	[56]
Risperidone	Schizophrenia	Dopamine agonist	Mucoadhesive nanoemulsion & solid lipid nanoparticles (SLNs)	[57]
Rivastigmine	Alzheimer's disease	Acetylcholinesterase (AChE) inhibitor	Chitosan-loaded nanoparticles CS-RHT-NPs	[58]
Ropinirole	Parkinson's disease	Dopamine D2 agonist	Mucoadhesive formulation	[59]
Sumatriptane	Migraine	Selective 5-HT 1D agonist	Micellar nanocarrier	[60]
Tacrine	Parkinson's disease	Anti-Parkinsonism Cholinergic activator	Nanoparticles	[61]
Tramadol HCl	Post-surgical pain, obstetric pain, cancer pain and chronic pain of mechanical and neurogenic origin	Synthetic opioid of amino cyclohexanol group	Microspheres	[62]
Valproic acid	Epilepsy, bipolar disorders, migraine and cancer	Aliphatic carboxylic acid with a broad spectrum anticonvulsant action	Nanostructured lipid carriers	[63]
Venlafaxine	Depression	Serotonin and norepinephrine reuptake inhibitor (SNRI)	Chitosan-loaded nanoparticles VLF-CS-NPs	[46]
Ziprasidone HCl	Schizophrenia	Fifth generation antipsychotic	Nanoemulsion	[64]
Zolmitriptan	Migrane	-	Micellar nanocarriers	[65]

The rats were killed 2 days after injection and by using eGFP-IHC and fluorescence microscopy eGFP expression was visualized. Throughout the rostral-caudal axis of the rat brain, eGF-positive cell were found sufficiently. And thus through this it confirmed that intranasal delivery of DNA NPs can transfect and express the encoded protein in the rat's brain and can bypass the BBB, by non-invasive approach for gene therapy of CNS disorders [66].

Nanoparticulates for Systemic Delivery: Kumar M. *et al.*, developed streptomycin sulfate (STRS) loaded (STRS-SLNs) solid lipid nanoparticles for IN delivery. It is an aminoglycoside, a bactericidal antibiotic potent against both Gram +ve and Gram -ve organisms as well as Mycobacterium, respectively. By using patented nanocolloidal aqueous dispersion technique (Indian Patent Application 3093/DEL/2012), it was prepared with

entrapment efficiency (54.83±2.1%) and small particle size (140.1±7.0nm). In comparison to free (F)-CSTRS, bio distribution studies using ^{99m}Tc showed 3.15 to 11.0 times greater concentration in the brain blood of mice, upon IN administration of STRS-SLNs. Lower concentration (3.3 times) incriminate lower nephrotoxicity in kidneys, along with 4 and 12 times lower levels of drug in spleen and liver and it also specifies its lesser accumulation in reticuloendothelial system organs [67]. As per literature search, PLGA-NPs (Polylactic co-glycolic acid) for RIS (Risedronate sodium) could be a drug delivery system for the prevention and treatment of osteoporosis via nasal route using NP approach, in order to reduce peripheral toxic effects. NPs of RIS were prepared by nanoprecipitation method. Table 3 gives an overview of some of the marketed nasal products for systemic delivery.

Table 3: Marketed nasal products for systemic delivery

Product name	Drug	Indication	Manufacturer	References
Aerodiol	Estradiol	Management of menopause symptoms	Servier	[68]
Atrona	Ipratropium bromide	Treatment of bronchospasm	Boehringer Ingelheim, Belg.	[69]
Imigran®	Sumatriptan	Management of migraine	GlaxoSmithKline	[70]
Miacalcic®	Calcitonin	Post-menopausal osteoporosis	Novartis Pharmaceutical Ltd.	[71]
Miacalcin®	Salmon calcitonin	Osteoporosis	Novartis Pharma	[72]
Migranal®	Dihydroergotamine mesylate	Management of migraine	Novartis Pharma	[73]
Desmospray	Desmopressin acetate	Nocturnal Enuresis	Ferring Pharmaceuticals, United Kingdom	[74]
Minrin®, Octostim®	Desmopressin acetate	Nocturnal enuresis, Management of diabetes insipidus, Hemophilia A, Von Willebrand's disease (type 1)	Ferring, Mexico	[75]
Nascobal®	Cyanocobalamine	Vit-B12 deficiency	Endo Pharmaceuticals	[76]
Nicotrol®	Nicotine	Smoking cessation	Pfizer	[77]
Stadol NS®	Butorphanol tartrate	Management of pain/Migraine	Bristol Myers Squibb	[78]
Suprecur®, Profact®, Suprefact®	Buserelin (acetate)	Prostate carcinoma, endometriosis	Sanofi-Aventis	[79]
onzetra® xsail®	Sumatriptan	Treatment of acute migraine	Avanir Pharmaceuticals, Inc.	[80]
Synarel®	Nafarelin acetate	Treatment of symptoms (dysmenorrhea, dyspareunia and pelvic pain) associated with endometriosis.	Pfizer Inc.	[81]
Syntocinon®	Oxytocin	Stimulates milk ejection in breast feeding mothers	Novartis Pharma	[82]
Zomig®	Zolmitriptan	Management of migraine	Cipla	[83]

Nanoparticulates for Vaccine Delivery: A suitable way is provided by the nanocarriers for the nasal delivery of antigenic molecules. The optimized vaccine nanocarriers offer a promising way for nasal mucosal vaccination. Nanoparticulate delivery systems provide more effective antigen recognition by immune cells along with improved protection and facilitated transport of the antigen, respectively [84].

Lipid-based nanocarriers and Polymeric nanocarriers are responsible for nasal antigen delivery.

ISCOMs, Liposomes, Synthetic Biomimetic Supramolecular Biovector TM and other lipid-based nanocarriers come under Lipid-based nanocarriers.

Biodegradable polyester nanoparticles, Lectin-decorated polymeric particles, Polysaccharide-based nanocarriers come under Polymeric nanocarriers, respectively.

Antigens associated with ISCOMs were efficiently processed by different antigen presenting cells (DCs, B-cells, epithelial cells, etc.) [85]. Chitosan: TPP and chitosan-PEG:TPP nanoparticles are more advantageous than the electrostatic chitosan-DNA. Complexes, as the carrier possibly provides the nucleic acids with better protection and sustained release characteristics as well [86]. Table 4 provides a list of products under investigation for IN vaccination.

Table 4: Products under investigation for IN vaccination

Vaccine/Antigen	Delivery system	References
Chlamydia Vaccine	Nanoemulsion	[87]
Pneumococcal Vaccine	C-terminal fragment of Clostridium perfringens enterotoxin (C-CPE)	[88]
Ghrelin vaccine	Nanogel	[89]
DNA vaccine (Chitosan)	Chitosan nanoparticles	[90]
Human sperm surface antigen – CD52	Liposomes	[91]
Influenza A vaccine	Nanoemulsion	[92]
Influenza subunit vaccine	N-trimethyl Chitosan nanoparticles	[93]
Influenza vaccine	PLGA nanoparticles	[94]
Foot-and-mouth disease (FMD) vaccine	Gel	[95]
Multivalent group A streptococcal vaccines	Liposomes	[96]
Ovalbumin	Liposomes	[97]
pDNA vaccine	Chitosan nanoparticles	[98]
Peptide T vaccine	PLGA microspheres	[99]

Researcher's findings in laboratory animal models and some of them being tested under clinical trial in humans are listed in Table 5.

Table 5: Marketed/Under clinical trial nasal products for vaccination

Product name/Antigen	Indication	Delivery System	Status	Manufacturer	References
Human influenza vaccine (Nasalfu Berna)	Influenza	Virosomes (Spray)	Marketed	Berna Biotech	[100]
Flu Avert®	Influenza	Drops	Marketed	Intervet/Merck	[101]
FluNsure™	Influenza	Proteosomes (Nanoparticulate)	Marketed	ID Biomedical	[13]
FluMist®	Influenza	Spray	Marketed	Astrazeneca	[102]
Maxi/Guard Nasal Vac	Bordetella bronchiseptica vaccine	Drops	Marketed	Addison Biological Laboratory	[103]
Nobivac BP®	Bordetella bronchiseptica Diseases	Suspension drops	Marketed	Intervet	[13]
Nasovac-S-(H1N1)	Swine flu	Bioadhesive delivery system	Marketed	Serum Inst. Of India Pvt. Ltd.	[104]
Calcitonin 'Novartis' 100	Post-menopausal osteoporosis	Nasal spray	Marketed	Novartis Pharma	[105]
AS01™	Malaria	Liposomes	Completed	GlaxoSmithKline	[106]
AS02™	Malaria	Oil-in-water emulsion	Completed	GlaxoSmithKline	[107]
DeltaFLU	Pandemic influenza	Nasal spray	Clinical trail (Phase II)	Green Hills Biotechnology AG	[108]
Influenza (H7N9 inactivated virus)	Flu	Matrix-M1™	Clinical trail	Novavax	[109]
MEDI 534	Parainfluenza virus type 3/respiratory syncytial virus	-	Clinical trail completed	MedImmune	[110]
MEDI 559	Respiratory Syncytial Virus	-	Clinical trail (Phase I/IIa)	MedImmune	[111]
MEDI 560	Parainfluenza virus type 3	-	Terminated	MedImmune	[112]
Norwalk VLP	Norovirus	Powder	Phase I-II completed	Takeda Pharmaceuticals Ltd.	[113]
StrepAvax®	Group A streptococcus diseases	Proteosomes (Nanoparticulate)	Phase II	ID Biomedical	[114]
Feline trivalent vaccine	Against calici herpes-1 and parvovirus	Drops	Marketed	Heska	[115]

A good number of patents received on Nasal drug delivery formulations, even though some of them have not reached the market yet. Some of the patents related to Nasal drug delivery system are presented in Table 6.

Table 6: Patents on Intranasal Drug Delivery Systems

Drug	Indication	Delivery System	Patent	References
Benzodiazepines	Epilepsy	Nasoadhesivemicroemulsion	A Misra, TK Vyas. 1061/MUM/2004	[116]
Epinephrine	Anaphylaxis	Nasal spray	Nigel Ten Fleming US20150005356 A1 (2015)	[117]
Ketamine	Depression	Nasal sprays and inhalers	Dennis S. Charney, Sanjay J. Mathew, Hussein K. Manji, Carlos A. Zarate, John H. Krystal US20170181966 A1 (2017)	[118]
Ketorolac®	Analgesic/anti-inflammatory	Drops or spray	Giancarlo Santus, Giuseppe Bottoni, Ettore Bilato US6333044 B1 (2001)	[119]
Naloxone	Septic shock	Nasal sprays	Roger Crystal, Michael Brenner Weiss. US9211253 B2 (2015)	[120]
Proteasomes	Neurodegenerative disorders	Nanoemulsion	D Frenkel, Rmaron, D Burt, HL Weiner. US20060229233A1 (2006)	[121]
Glatiramer	Neuroendocrinologic disorders, such as Female Sexual Disorder (FSD)	Gel formulation	Claudia Mattern. US20170189414 A1 (2017)	[122]
Noseafix®				
Triptans, Caffeine	Migraine	Nasoadhesivemicroemulsions	A Misra, TK Vyas. 1125/MUM/(2004)	[123]
Zolpidem	Insomnia	Cyclodextrin/chitosan Sols	JD Castile, YH Cheng, PG Jenkins. US0140981A1 (2007)	[124]

The Nasal drug delivery devices do exist and some are under development. Table 7 provides list of patents on Nasal drug delivery devices.

Table 7: Patents on Intranasal Drug Delivery Devices

Delivery devices	Patent	References
Pressurized Olfactory Delivery (POD) device	John D. Hoekman, Michael Hite, Alan Brunelle, Joel RELETHFORD, Rodney J. Y. HO. WO2012119153 A2. (2012)	[125]
Pfeiffer/Aptar single-dose device	Roger Crystal, Michael Brenner Weiss. US9211253 B2. (2015)	[120]
Nasal breathing device	Luigi Corsaro. US 5727543. (1998)	[126]
Nasal dilator	Martin O'Connell, Keith Yeager. US 20170027736.	[127]
Nasal inserts	Adva Beck Amon. US 8517026. (2013)	[128]
Nasal inserts	Adva Beck Arnon. 8839790. (2014)	[129]
Ergonomic nasal cannula	Darin B. Atherton. US 20170007794. (2017)	[130]
Nasal filter structure	Joseph K. Moore. US 9132300. (2015)	[131]
Nasal cavity insertion device fixture	Kenji Hioki, Hiroshi Yamada. 20140094840. (2014)	[132]

Effect of Physical Form of Delivery System

Solid: Solid drugs are chemically more stable in comparison with the solutions [133].

Powder Formulations: Powder formulations have many advantages over other formulations viz., solutions and gels, such as application in higher dosages and improved drug stability along with better drug absorption [134]. It has been shown in literature that IN delivery of Sumatriptan powder with a novel Breath Powered was more productive form of drug delivery, which provided a higher peak and rapid exposure with a lower delivered dose as comparison with speedy absorption then nasal spray [135].

Semi-Solid: For a more precise drug delivery through nasal cavity, gel approaches have been assessed in recent time [13].

Gel Formulations: Gels are more preferable than other physical form as they provide larger drug transport along with longer residence time of the formulation at the site of absorption. Gel lowers the post-nasal spread out and anterior run-off by holding the dosage form in nasal mucosa, as a result, the residence time increases and mucociliary clearance decreases [136].

An investigation suggested that nasal absorption of progesterone can be enhanced by using Carbopol nasal gel, which was done in rabbits and could be improved by using β cyclodextrin (CD) as a nasal absorption enhancer [137]. The study showed that Natesto Testosterone Nasal Gel has an advantage of self-administration with multiple dose dispenser and having a minimum incidence of local reaction [138]. The researcher's prepared (Solution, gel and lyophilized powder) nasal dosage forms of metoclopramide hydrochloride (MTC) by using sodium carboxy methylcellulose (NaCMC), a mucoadhesive polymer. And it was observed that the NaCMC-based gel delivery system of MTC holds higher bioavailability than those of solution and powder [139]. Dukovski *et al.* [140]

developed a nanoparticle loaded *in-situ* gelling system worthy for corticosteroid nasal delivery, responsible for the treatment of chronic rhino sinusitis with nasal polyps. They proposed lipid/alginate nanoparticle, whose size ranges from 252.3 ± 2.4 nm, polydispersity index 0.241, with zeta-potential -31.7 ± 1.0 mv and dexamethasone (Dex) content of 255 ± 7 μ g ml respectively, which eventually dispersed in a pectin solution of 5mg ml⁻¹ that gone through a sol-gel phase transition which were triggered by Ca²⁺ available in nasal mucosa and allow delivery of corticosteroid beyond the nasal valve. The proposed study gave an *in vitro* proof-of-concept of developed system that ensure efficient corticosteroid delivery with prolonged local action due to prolonged contact time with nasal mucosa upon gelation [140].

Liquid: Liquid (suspension and solution) sprays more convenient over powder sprays for the reason that the powder sprays simply provoked the nasal mucosal irritation [141]. The nasal drug solutions are administered as nasal drops, sprays and as metered dose nebulizer. An active ingredient's dose administered depends upon the concentration of drug in the formulation and the volume of drug, respectively.

Sprays: The case study provided a guideline to quality by design (QbD) approach for suspension type nasal spray products development e.g. beclomethasone nasal spray suspension, ciclesonide nasal spray suspension, fluticasone nasal spray suspension and mometasone nasal spray suspension. This novel model can surpass generic nasal spray developments by reducing cost, development time and manpower and thus reducing number of experiments required to procure *in vitro* equivalency with innovator drug products as well [142].

Drops: Study showed that drops administered in the right manner should reach the middle meatus in greater quantities and also improved penetration of nasal valve [143].

Table 8: Challenges related to nasal drug delivery

Challenges	Resolution to the challenges
Less retention time	Increase viscosity/bioadhesive systems
Permeability of drug	Optimum drug concentration, decrease MCC
Irritation and irreversible damage of the cilia on nasal mucosa	Formulation's should be at nasal pH
Disrupt and sometimes dissolution of nasal membrane	High concentration of absorption enhancers should be reduced
Adversely affected by pathological conditions(cold and cough)	Simultaneous use of compatible decongestants along with other therapeutic drugs.
Solubility of hydrophilic drug	Incorporation of solubilizer

Advancements in Nasal Delivery: Dr Djupesland (Opti Nose AS, Norway) came with a new concept for targeting the nasal cavity, with the device known as (OptiMist), consists of two nozzles to be inserted into a nostril. The major advantage of the device is that it minimizes the lung deposition i.e., <1% of the total dose gets collected in the lungs after bidirectional delivery and maximizes the distribution of aerosolized droplets in the nose [144]. The novel concept of Breath powered Bi Directional™ has been came into existence that responsible to overcome many of the inherent limitations of traditional nasal devices [145]. Impel NeuroPharma's Pressurized Olfactory Delivery (POD) device is an example of clinical evaluation, responsible for successful delivery of aerosolized drugs to the nasal cavity [33].

Challenges in Nasal Delivery: The transnasal delivery getting explored now a days with great interest for delivering the non-peptide small and macromolecules and neuropeptides bypassing the BBB, prevent local toxicities and achieve rapid, improved and prolonged drug delivery [146]. One of the limiting factors in nasal drug delivery, is the restricted volume of the nasal cavity, which hindered the amount of formulation that can be administered [147] while other challenges with their resolution are listed in Table 8. Without an absorption modifier, nasal administration of peptides (Such as calcitonin and insulin) and proteins results in factor called low bioavailability across the nasal mucosa, possibly owed to enzymatic degradation of the molecule during passage through the epithelial barrier and/or mucociliary elimination from the nasal cavity [15, 147]. Particle size of the droplet or powder responsible for drugs deposition, the ideal size for the retention in the nasal cavity is between 5-7 μm . It would be exhaled if particle size is <0.5 μm whereas if particle size is <10 μm , then it would be deposited in the upper respiratory tract [148]. At present, very limited adjuvants are being used in marketed vaccines, the most probable reason behind this is that, for vaccine approval, clinical trial is necessary, which are often very lengthy and difficult. Ideally, an adjuvant must be capable to stimulate cellular, mucosal and humoral immune responses

discretely or concurrently, depending on the desired treatment [149]. In recent times, treatment of neurological diseases has become one of the most remarkable challenge and in response to this challenge, nanotechnology provided promising solutions to the same.

There is a need to address some questions in order to meet the possible opportunities for the success of IN route deliver, like- it is unknown if there is a any size limit which governing what can be delivered to the brain via the intranasal route [150]. Study with dextrans suggested that there was an inverse relationship between CSF and the MW concentration following IN administration [151]. Generally, solution cannot stay for a longer period of time into the nasal cavity which leads to less retention time due to mucociliary clearance is another major challenge for delivered a drug into a nasal cavity. Repeated uses of nasal drops cause loss of ciliary layer, loss of epithelial cell and shrinkage of mucosal layer that might be incurred by inactive or incompatible active ingredient of formulations, respectively [152]. Use of Gelling agents results in the increase in retention time of the drug along with decrease in drainage [153]. The drugs can be absorbed efficiently and quickly across the nasal membrane, when it is lipophilic and having a MW of less than 1kDa and thus polar drugs not easily transport across nasal membrane [14]. Drug's permeation can be affected by the pH of the formulation, as well as that of nasal surfaces. pH of the nasal formulation should be adjusted to 4.5-6.5 to avoid nasal irritation. In addition, prevents the growth of bacteria along with efficient drug permeation. Also permeation of the compound normally increases through nasal mucosa on increasing lipophilicity [153]. Nasal absorption and aqueous solubility of the insoluble compounds of the drug increase by the use of solubilizer. Specialized systems such as lipid emulsion [154] microspheres (Using carbopol, chitosan, 934P and lactose), films, liposomes, niosomes, proliposomes, provides a better chance of permeation for the drugs. In the perspective of sound researchers, the nasal drug delivery in combination with nano-technology undo the future understanding and therapeutics of brain function and CNS diseases.

CONCLUSIONS

The advantages of nasal administration put this cavity as a distinguish option for future drug delivery. The permeability of nasal mucosal surface offers the bioavailability comparable to systemic routes for some drugs that fact can be utilized for the development of non-invasive option of such parenteral therapeutics. The possibility of direct nose-to-brain transfer of nasally administered therapeutics also open the opportunities for the development of new formulations for better or equivalent bioavailability with lower doses by minimizing the pre-systemic and systemic losses. Despite of number of researches reported for nasal delivery of various molecules for different purposes i.e. local, systemic, nose-to-brain and vaccine delivery, the scientific community is still lacking to utilize the maximum therapeutic benefits. The challenges associated with nasal deliverance and lack of knowledge about the mechanism responsible for disposition of nasally administered drug, restricted the development of new dosage forms for different pathological conditions especially in case of nose-to-brain transport.

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REFERENCES

- Pardridge, W.M., 1991. In: Peptide Drug Delivery to the Brain. Raven Press. New York, pp: 99-122.
- Thorne, R.G., C.R. Emory, T.A. Ala and W.H. Frey, 1995. Quantitative analysis of the olfactory pathway for the drug delivery. Brain Res., 692: 278-282.
- Illum, L., 2003. Nasal drug delivery - possibilities, problems and solutions. J. Controlled. Rel., 87: 187-198.
- Illum, L., I. Jabbal-Gill and M. Hinchcliffe, 2001. Chitosan as a novel nasal delivery system for vaccines. Adv. Drug Deliv Rev., 51: 81-96.
- Pontiroli, A.E., M. Alberetto, A. Calderara, E. Pajetta and G. Pozza, 1989. Nasal administration of glucagon and human calcitonin to healthy subjects: a comparison of powders and spray solutions and of different enhancing agents. Eur. J. Clin Pharmacol., 37: 427-430.
- Pontiroli, A.E. and M.D., 2015. Intranasal glucagon: a promising approach for treatment of severe Hypoglycemia. J Diabetes Sci and Tec., 9: 38-43.
- Ali, J., M. Ali, S. Baboota, J.K. Sahni, C. Ramassamy, L. Dao and Bhavna, 2010. Potential of Nanoparticulate Drug Delivery System by Intranasal Administration. Current Pharm Design, 16: 1644-1653.
- Urrusuno, R.F., P. Calvo, C.R. Lopez, J.L.V. Jato and M.J. Alonso, 1999. Enhancement of Nasal Absorption of Insulin Using Chitosan Nanoparticles. Pharm. Res., 16: 1576-1581.
- Kaur, P., T. Garg, G. Rath and A.K. Goyal, 2015. In situ nasal gel drug delivery: A novel approach for brain targeting through the mucosal membrane. Artificial Cells, Nanomedicine and Biotechnology, pp: 1-10.
- Garg, T., R.S.R. Murthy, A.K. Goyal, S. Arora and B. Malik, 2012a. Development, optimization & evaluation of porous chitosan scaffold formulation of gliclazide for the treatment of type-2 diabetes mellitus. Drug Deliv Lett., 2: 251-261.
- Patel, T., J. Zhou and J.M. Piepmeier, 2012. Polymeric nanoparticles for drug delivery to the central nervous system. Adv. Drug Deliv Rev., 64: 701-705.
- Alex, A.T., A. Joseph, G. Shavi, J.V.Rao and N. Udupa, 2014. Development and evaluation of carboplatin-loaded PCL nanoparticles for intranasal delivery. Drug Deliv., pp: 1-10.
- Kumar, A., A.N. Pandey and S.K. Jain, 2015. Nasal-nanotechnology: revolution for efficient therapeutics delivery. Informa Healthcare USA, Inc, pp: 1-13.
- Rahisuddin, P.K. Sharma, G. Garg and M. Salim, 2011. Review on Nasal Drug Delivery System with Recent Advancemnt. Int. J. Pharmacy Pharm. Sci., 3: 6-11.
- Jadhav, K.R., M.N. Gambhire, I.M. Shaikh, V.J. Kadam and S.S. Pisal, 2007. Nasal Drug Delivery System-Factors Affecting and Applications. Current Drug Therapy, 2: 27-38.
- Ursapharm. Allergo-Comod Nasal. <http://www.pharmaturca.com/marke/ursapharm.aspx?tabId=123>, (Accessed on 14.08.17).
- Drugs.com. <https://www.drugs.com/international/cromoglicic-acid.html>, (Accessed on 14.08.17).
- <https://www.drugs.com/uk/pdf/leaflet/928905.pdf>, (Accessed on 14.08.17).
- https://www.accessdata.fda.gov/drugsatfda_docs/label/2012/020114s023lbl.pdf, (Accessed on 14.08.17).
- https://au.gsk.com/media/222812/bactroban_nasal_oointment_cmi_au_004_approved.pdf, (Accessed on 14.08.17).

21. https://www.gsksource.com/pharma/content/dam/GlaxoSmithKline/US/en/Prescribing_Information/Becanase_AQ/pdf/BECONASE-PI-PIL.PDF, (Accessed on 14.08.17).
22. Decadron. <https://online.epocrates.com/drugs/151910/Decadron/Monograph>, (accessed on 14.08.17).
23. <https://au.gsk.com/en-au/products/our-consumer-healthcare-products/respiratory/flixonase/>, (Accessed on 14.08.17).
24. http://www.janssen.com/netherlands/sites/www_janssen_com_netherlands/files/livocabneus_b_0.pdf, (Accessed on 14.08.17).
25. <http://www.clinicaladvisor.com/nasalcrom/drug/3095/>, (Accessed on 14.08.17).
26. https://www.google.co.in/search?q=nasivin+nasal+spray+Braco&tbm=isch&tbo=u&source=univ&sa=X&ved=0ahUKEwjn47vtHVAhVLqI8KHY0VA50Q7AkINw&biw=1383&bih=645#imgc=kJ_8f2Mv7XjttM: (Accessed on 14.08.17).
27. <https://www.fda.gov/downloads/Drugs/DrugSafety/DrugShortages/UCM463501.pdf>, (Accessed on 14.08.17).
28. <http://www.thehindubusinessline.com/todays-paper/tp-economy/now-novartis-otrivin-spray-will-be-available-over-the-counter/article1065586.ece>, (Accessed on 14.08.17).
29. <https://www.patanase.com/>, (Accessed on 14.08.17).
30. <http://www.mims.com/thailand/company/info/stada>, (Accessed on 14.08.17).
31. <https://www.jnj.com/media-center/press-releases/rhinocort-allergy-spray-now-available-over-the-counter-nationally>, (Accessed on 14.08.17).
32. <http://www.medindia.net/drug-price/benzalkonium-chloride/syntaris.htm>, (Accessed on 14.08.17).
33. Costantino, H.R., L. Illum, G. Brandt, P.H. Johnson and S.C. Quay, 2007. Intranasal delivery: Physicochemical and therapeutic aspects. *Int J. Pharm.*, 337: 1-24.
34. Illum, L., 2000. Transport of drugs from the nasal cavity to the central nervous system. *European J. Pharm S.*, 11: 1-18.
35. Illum, L., 2003. Transport of drugs from the nasal cavity to the central nervous system. *J Controlled Rel.*, 87: 187-198.
36. Illum, L., 2015. Intranasal Delivery to the Central Nervous System. In: *Blood-Brain Barrier in Drug Discovery: Optimizing Brain Exposure of CNS Drugs and Minimizing Brain Side Effects for Peripheral Drugs*, 1st ed. John Wiley & Sons, Inc., pp: 535-565.
37. Illum, L., N.F. Farraj and S.S. Davis, 1994. Chitosan as a Novel Nasal Delivery System for Peptide Drugs. *Pharm Res.*, 11: 1186-1189.
38. Yingying, X.U., P.W. Yuen and J.K.W. Lam, 2014. Intranasal DNA Vaccine for Protection against Respiratory Infectious Diseases: The Delivery Perspectives. *Pharmaceutics*, 6: 378-415.
39. Mujawar, N., S. Ghatage, S. Navale, B. Sankpal and S. Patil, 2014. Nasal Drug Delivery: Problem Solution and Its Application. *J Current Pharma Res.*, 4: 1231-1245.
40. Bhavna, S.M.D., M. Ali, R. Ali, A. Bhatnagar, S. Baboota and J. Ali, 2014. Donepezil nanosuspension intended for nose to brain targeting: *In vitro* and *in vivo* safety evaluation. *Int J. Biological Macromolecules*, 67: 418-425.
41. Bian, J., Z.Yuan, X. Chen, Y. Gao, C. Xu and J. Shi, 2016. Preparation of surface multiple-coated polylactide acid drug-loaded nanoparticles for intranasal delivery and evaluation on its brain-targeting efficiency. *Drug Deliv.*, 23: 269-27.
42. Jafarieh, O., S. Md, M. Ali, S. Baboota, J.K. Sahni, B. Kumari, A. Bhatnagar and J. Ali, 2014. Design, characterization and evaluation of intranasal delivery of ropinirole-loaded mucoadhesive nanoparticles for brain targeting. *Informa Healthcare USA, Inc*, pp: 1-8.
43. Jain, K., S. Sood and K. Gowthamarajan, 2014. Optimization of artemether-loaded NLC for intranasal delivery using central composite design. *Informa Healthcare USA, Inc*, pp: 1-15.
44. Zhao, Y.Z., X. Li, C.T. Lu, M. Lin, L.J. Chen, Q. Xiang, M. Zhang, R.R. Jin, X. Jiang, X.T. Shen, X.K. Li and J. Cai, 2014. Gelatin nanostructured lipid carriers-mediated intranasal delivery of basic fibroblast growth factor enhances functional recovery in hemiparkinsonian rats. *Nanomedicine: Nanotechnology, Biology and Med.*, 10: 755-764.
45. Jain, N., S. Akhter, G.K. Jain, Z.I. Khan, R.K. Khar and F.J. Ahmad, 2011. Antiepileptic Intranasal Amiloride Loaded Mucoadhesive Nanoemulsion: Development and Safety Assessment. *J. Biomedical Nanotechnology*, 7: 142-143.
46. Mittal, D., A. Ali, S. Md, S. Baboota, J.K. Sahni and J. Ali, 2013. Insights into direct nose to brain delivery: current status and future perspective. *Drug Deliv.*, pp: 1-12.
47. Bragagni, M., N. Mennini, C. Ghelardini and P. Mura, 2012. Development and characterization of niosomal formulations of doxorubicin aimed at brain targeting. *J. Pharm. Pharm. Sci.*, 15: 184-96.

48. Shah, V., M. Sharma, R. Pandya, R.K. Parikh, B. Bharatiya, A. Shukla and H.C. Tsai, 2017. Quality by Design Approach for an Insitu Gelling Microemulsion of Lorazepam via Intranasal route. *Materials Science & Engineering C*, pp: 1-46.
49. Jayachandra, B.R., P.P. Dayal, K. Pawar and M. Singh, 2011. Nose-to-brain transport of melatonin from polymer gel suspensions: a microdialysis study in rats. *J. Drug Target*, 19: 731-40.
50. Gaillard, P.J., C.C.M. Appeldoorn, J. Rip, R. Dorland, S.M.A. Van Der Pol, G. Kooij, H.E. Vries and A. Reijerkerk, 2012. Enhanced brain delivery of liposomal methylprednisolone improved therapeutic efficacy in a model of neuroinflammation. *J. Controlled Rel.*, 164: 364-369.
51. Ruan, Y., L. Yao, B. Zhang, S. Zhang and J. Guo, 2011. Antinociceptive properties of nasal delivery of Neurotoxin-loaded nanoparticles coated with polysorbate-80. *Peptides*, 32: 1526-1529.
52. Wu, H., J. Li, Q. Zhang, X. Yan, L. Guo, X. Gao, M. Qiu, X. Jiang, R. Lai and H. Chen, 2012. A novel small Odorranalectin-bearing cubosomes: Preparation, brain delivery and pharmacodynamic study on amyloid-b25-35-treated rats following intranasal administration. *European J. Pharm. and Biopharm.*, 80: 368-378.
53. Wen, Z., Z. Yan, K. Hu, Z. Pang, X. Cheng, L.R. Guo, Q. Zhang, X. Jiang, L. Fang and R. Lai, 2011. Odorranalectin-conjugated nanoparticles: Preparation, brain delivery and pharmacodynamic study on Parkinson's disease following intranasal administration. *J. Controlled Rel.*, 151: 131-138.
54. Salama, H.A., A.A. Mahmoud, A.O. Kamel, M.A. Hady and A.S.G. Awad, 2012. Brain delivery of olanzapine by intranasal administration of transfersomal vesicles. *J. Liposome Res.*, pp: 1-10.
55. Seju, U., A. Kumar and K.K. Sawant, 2011. Development and evaluation of olanzapine-loaded PLGA nanoparticles for nose-to-brain delivery: *In vitro* and *in vivo* studies. *Acta Biomaterialia*, 7: 4169-4176.
56. Joshi, A.S., H.S. Patel, V.S. Belgamwar, A. Agrawal and A.R. Tekade, 2012. Solid lipid nanoparticles of ondansetron HCl for intranasal delivery: development, optimization and evaluation. *J Mater Science Mater Med.*, 4702-7.
57. Kumar, M., A. Misra, A.K. Babbar, A.K. Mishra, P. Mishra and K. Pathak, 2008. Intranasal nanoemulsion based brain targeting drug delivery system of risperidone. *Int. J. Pharma.*, 358: 285-291.
58. Fazil, M., S. Md, S. Haque, M. Kumar, S. Baboota, J.K. Sahni and J. Ali, 2012. Development and evaluation of rivastigmine loaded chitosan nanoparticles for brain targeting. *Eur. J. Pharma Science*, 47: 6-15.
59. Khan, S., K. Patil, N. Bobade, P. Yeole and R. Gaikwad, 2010. Formulation of intranasal mucoadhesive temperature-mediated in situ gel containing ropinirole and evaluation of brain targeting efficiency in rats. *J. Drug Targeting*, 18: 223-234.
60. Jain, R., S. Nabar and P. Dandekar, 2010a. Formulation and evaluation of novel micellar nanocarrier for nasal delivery of sumatriptan. *Nanomedicine (Lond)*, 5: 575-87.
61. Luppi, B., F. Bigucci and G. Corace, 2011. Albumin nanoparticles carrying cyclodextrins for nasal delivery of the anti-Alzheimer drug tacrine. *Eur. J. Pharm Science*, 44: 559-65.
62. Belgamwar, V.S., H.S. Patel and A.S. Joshi, 2011. Design and development of nasal mucoadhesive microspheres containing tramadol HCl for CNS targeting. *Drug Delivery*, 18: 353-60.
63. Eskandari, S., J. Varshosaz, M. Minaiyan and M. Tabbakhian, 2011. Brain delivery of valproic acid via intranasal administration of nanostructured lipid carriers: in vivo pharmacodynamic studies using rat electroshock model. *Int J. Nanomedicine*, 6: 363-71.
64. Bahadur, S. and K. Pathak, 2012a. Buffered nanoemulsion for nose to brain delivery of ziprasidone hydrochloride: preformulation and pharmacodynamics evaluation. *Curr Drug Delivery*, 9: 596-607.
65. Jain, R., S. Nabar, P. Dandekar and V. Patravale, 2010b. Micellar nanocarriers: potential nose-to-brain delivery of zolmitriptan as novel migraine therapy. *Pharm Res.*, 27: 655-64.
66. Harmon, B.T., A.E. Aly, L. Padegimas, O.S. Laird, M.J. Cooper and B.L. Waszczak, 2014. Intranasal administration of plasmid DNA nanoparticles yields successful transfection and expression of a reporter protein in rat brain. *Macmillan Publishers Limited*, 21: 514-521.
67. Kumar, M., V. Kakkar, A.K. Mishra, K. Chuttani and I.P. Kaur, 2014. Intranasal delivery of streptomycin sulfate (STRS) loaded solid lipid nanoparticles to brain and blood. *International Journal of Pharmaceutics*, 461: 223- 233.
68. <http://www.medicines.org.au/files/secaerod.pdf>, (Accessed on 15.08.17).
69. <http://www.jodrugs.com/tradenames/250035-atronase.aspx>, (Accessed on 15.08.17).

70. https://www.gsk.com.au/resources.ashx/prescriptionmedicinesproductschilddata/downloads/606/File/F95A7B5D1F831E109833892131FA503A/CMI_Imigran_NasalSpray.pdf, (Accessed on 15.08.17).
71. <http://www.medicines.org.uk/emc/PIL.7884.latest.pdf>, (Accessed on 15.08.17).
72. https://www.accessdata.fda.gov/drugsatfda_docs/label/2012/020313s033lbl.pdf, (Accessed on 15.08.17).
73. <http://adisinsight.springer.com/drugs/800009972>, (Accessed on 15.08.17).
74. <https://www.medicines.org.uk/emc/company/50/Ferring%20Pharmaceuticals%20Ltd>, (Accessed On 15.08.17).
75. <http://mynetmeds.comprar-medicina.com/Stimate-Nasal-Spray.htm>, (Accessed on 15.08.17).
76. <https://www.nascobal.com/>, (Accessed 15.08.17).
77. https://www.pfizer.com/files/products/uspi_nicotrol_inhaler.pdf, (Accessed 15.08.17).
78. <https://dailymed.nlm.nih.gov/dailymed/archives/fdaDrugInfo.cfm?archiveid=69218>, (Accessed on 15.08.17).
79. https://books.google.co.in/books?id=4ICGAwAAQBAJ&pg=PA193&lpq=PA193&dq=Suprecur_+Profact_+Suprefact+Sanofi-Aventis&source=bl&ots=rHFuezfo4L&sig=JgN99iVnzZJ0rBtTmQfet-Cy4ng&hl=en&sa=X&ved=0ahUKEwi8j9zKmNvVAhUKY08KHe8rCTsQ6AEIJzAA#v=onepage&q=Suprecur_%2C%20Profact_%2C%20Suprefact%20Sanofi-Aventis&f=false, (Accessed on 15.08.17).
80. <https://www.avanir.com/press/avanir-pharmaceuticals-announces-fda-approval-onzetatm-xasaltm-avp-825-acute-treatment>, (Accessed on 15.08.17).
81. <http://labeling.pfizer.com/ShowLabeling.aspx?id=515>, (Accessed on 15.08.17).
82. <http://adisinsight.springer.com/drugs/800039450>, (accessed on 15.08.17).
83. <http://zomig.com/>, (Accessed on 15.08.17).
84. Csaba, N., M.Garcia-Fuentes and M.J.Alonso, 2009. Nanoparticles for nasal vaccination. *Adv Drug Delivery Reviews*, 61: 140-157.
85. Morein, B., M. Villacres-Eriksson, J. Ekstrom, K. Hu, S. Behboudi and K. Lovgren-Bengtsson. In: *ISCOM: A Delivery System for Neonates and for Mucosal Administration*. Advances in Veterinary Medicine. Academic Press, pp: 405-413.
86. Csaba, N., M. Garcia-Fuentes and M.J. Alonso, 2009. Nanoparticles for nasal vaccination. *Adv Drug Deliv Reviews*, 61: 140-157.
87. NanoBio's Chlamydia Vaccine Improves Clearance of Bacteria and Prevents Pelvic Inflammatory Disease in Mice. The next generation of vaccines. <http://www.nanobio.com/chlamydia-vaccine-update/> 2015 (Accessed on 16.08.17).
88. Suzuki, H., A. Watari, E. Hashimoto, M. Yonemitsu, H. Kiyono, K. Yagi, M. Kondoh and J. Kunisawa, 2015. C-Terminal Clostridium perfringens Enterotoxin-Mediated Antigen Delivery for Nasal Pneumococcal Vaccine. *Public Library of Science (PLOS) One*: 1-13.
89. Azegami, T., Y. Yuki, S. Sawada, M. Mejima, K. Ishige, K. Akiyoshi, H. Itoh and H. Kiyono, 2016. Nanogel-based nasal ghrelin vaccine prevents obesity. *Advance Online Publication*, pp: 1-10.
90. Xu, J., W. Dai, Z. Wang, B. Chen, Z. Li and X. Fan, 2011. Intranasal Vaccination with Chitosan-DNA Nanoparticles Expressing Pneumococcal Surface Antigen A Protects Mice against Nasopharyngeal Colonization by Streptococcus pneumonia. *Clin Vaccine Immunology*, 18: 75-81.
91. Hasegawa, A., Y. Fu and K. Koyama, 2002. Nasal immunization with diphtheria toxoid conjugated-CD52 core peptide induced specific antibody production in genital tract of female mice. *American J Reproductive Immunology*, 48: 305-311.
92. Myc A., J.F. Kukowska-Latallo, A.U. Bielinska, P. Caoa, P.P. Myc, K. Janczak, T.R. Sturm, M.S. Grabinski, J.J. Landers, K.S. Young, J. Changa, T. Hamoudad, M.A. Olszewski and J.R. Baker. Jr. Development of immune response that protects mice from viral pneumonitis after a single intranasal immunization with influenza A virus and nanoemulsion. *Vaccine*, 21: 3801-3814.
93. Kulkarni, A.D., H.M. Patel, S.J. Surana, Y.H. Vanjari, V.S. Belgamwar and C.V. Pardeshi, 2017. N, N, N-Trimethyl chitosan: An advanced polymer with myriad of opportunities in nanomedicine. *Carbohydrate Polymers*, 157: 875-902.
94. Lemoine, D., M. Deschuyteneer, F. Hogge and V. Preat, 1999. Intranasal immunization against influenza virus using polymeric particles. *J Biomater Sci Polym Edition*, 10: 805-825.
95. Cokcaliskan, C., F. Ozyoruk and R.N. Gursoy, 2014. Chitosan-based systems for intranasal immunization against foot-and-mouth disease. *Pharm. Dev. Technology*, 19: 181-8.
96. Hall, M.A., S.D. Stroop and M.C. Hu, 2004. Intranasal immunization with multivalent group A streptococcal vaccines protects mice against intranasal challenge infections. *Infection and Immunity*, 72: 2507-12.

97. Patel, G.B., A. Ponce, H. Zhou and W. Chen, 2008. Structural characterization of archaeal lipid mucosal vaccine adjuvant and delivery (AMVAD) formulations prepared by different protocols and their efficacy upon intranasal immunization of mice. *J Liposome Research*, 18: 127-43.
98. Iqbal, M., W. Lin and I. Jabbal-Gill, 2003. Nasal delivery of chitosan-DNA plasmid expressing epitopes of respiratory syncytial virus (RSV) induces protective CTL responses in BALB/c mice. *Vaccine*, 21: 1478-85.
99. Marazuela, E.G., R. Rodriguez and H. Fernandez-Garcia, 2008. Intranasal immunization with a dominant T-cell epitope peptide of a major allergen of olive pollen prevents mice from sensitization to the whole allergen. *Mol Immunology*, 45: 438-45.
100. Glück, R., 2002. Intranasal Immunization against Influenza. *J Aerosol Medicine*, 15: 221-228.
101. http://www.merck-animal-health-usa.com/products/130_120677/productdetails_130_121151.aspx/, 2017 (Accessed on 17.08.17).
102. AstraZeneca. FluMist.Quadrivalent.<https://www.flumistquadrivalent.com/>, 2017 (Accessed on 17.08.17).
103. Addison Biological Laboratory. MAXI/GUARD Nasal Vac®. <https://addisonlabs.com/product/maxiguard-nasal-vac/>, 2016 (Accessed on 17.08.17).
104. Serum Institute of India Pvt. Ltd. Influenza vaccine (Human, live attenuated), (freeze-dried) pandemic (h1n1) (Intranasal). http://www.seruminstitute.com/product_influenza_vaccines.php, 2017 (Accessed on 17.08.17).
105. Novartis. Calcitonin "Novartis" 100 IU - Nasalspray. <https://medikamio.com/de-at/medikamente/calcitonin-novartis-100-ie-nasalspray/pil>, (Accessed on 17.08.17).
106. ClinicalTrials.gov. AS01™. <https://clinicaltrials.gov/ct2/results?cond=&term=AS01%E2%84%A2&cntry1=&state1=&Search=Search>, (Accessed on 17.08.17).
107. ClinicalTrials.gov. AS02™. <https://clinicaltrials.gov/ct2/results?cond=&term=AS02%E2%84%A2&cntry1=&state1=&recrs=>, (Accessed on 17.08.17).
108. PRNewswire. AVIR Green Hills Biotechnology AG: deltaFLU - Innovative Vaccines for Intranasal Spray Delivery or Oral Administration. 2011. (Accessed on 17.08.17).
109. Novavax. A(H7N9) VLP Antigen Dose-Ranging Study With Matrix-M1™ Adjuvant. <https://clinicaltrials.gov/ct2/show/NCT02078674?term=Novavax&rank=8>, (Accessed on 18.08.17).
110. ClinicalTrials.gov. MEDI 534. <https://clinicaltrials.gov/ct2/results?cond=Parainfluenza+virus+type+3%2Frespiratory+syncytial+virus&term=MEDI+534&cntry1=&state1=&recrs=>, (Accessed on 17.08.17).
111. ClinicalTrials.gov. MEDI 559. <https://clinicaltrials.gov/ct2/show/NCT00767416?term=MEDI+559&rank=1>, (Accessed on 17.08.17).
112. ClinicalTrials.gov. MEDI 559. <https://clinicaltrials.gov/ct2/show/results/NCT00508651?term=MEDI+560&rank=1>, (Accessed on 17.08.17).
113. Baehner, F., H. Bogaerts and R. Goodwin, 2016. Vaccines against norovirus: state of the art trials in children and adults. *Clinical Microbiology and Infection*, 22: 136-139.
114. Nandkumar, P.P., S.D. Manohar and S.R. Bhanudas, 2015. Nasal drug delivery system an overview. *World J Pharma Research*, 4: 372-398.
115. Drugs.com. HESKA Feline UltraNasal FVRC Vaccine. <https://www.drugs.com/vet/heska-feline-ultranasal-fvrc-vaccine.html>, 2017 (Accessed on 17.08.17).
116. Misra, A. and T.K. Vyas, 2005. Drugs Loaded Nasoadhesive Microemulsions For Brain Targeted Brosk Delivery In Acute Epilepsy. 1061/MUM/2004.
117. Fleming, N.T., 2015. Intranasal Formulation of Epinephrine for the Treatment of Anaphylaxis. US 20150005356 A1.
118. Charney, D.S., S.J. Mathew, H.K. Manji, C.A. Zarate and J.H. Krystal, 2017. Intranasal administration of ketamine to treat depression. US 20170181966 A1.
119. Santus, G., G. Bottoni and E. Bilato, 2001. Therapeutic compositions for intranasal administration which include KETOROLAC®. US 6333044 B1.
120. Crystal, R. and M.B. Weiss, 2015. Nasal drug products and methods of their use. US 9211253 B2.
121. Frenkel, D., R. Maron, D. Burt and H. Weiner, 2006. Compositions and methods for treating neurological disorders. US 20060229233 A1.
122. Mattern, C., 2017. Controlled release delivery system for nasal applications and method of treatment. US 20170189414 A1.

123. Misra, A. and T.K. Vyas, 2005. Drugs Loaded Intranasal Nasoadhesive Microemulsions for Brain Targeted Delivery In Migrane. 1125/MUM/2004.
124. Castile, J., Y.H. Cheng, P. Jenkins, A. Smith and P. Watts, 2007. Intranasal compositions. US 20070140981 A1.
125. Hoekman, J.D., M. Hite, A. Brunelle, J. Relethford and Y.H.O. Rodney, 2012. Nasal drug delivery device. WO 2012119153 A2.
126. Corsaro, L., 1998. Nasal breathing device. US 5727543 A.
127. Connell, M.O. and K. Yeager, 2017. Nasal dilator. US 20170027736 A1.
128. Amon, A.B., 2013. Nasal inserts. US 8517026 B2.
129. Arnon, A.B., 2014. Nasal inserts. US 8839790 B2.
130. Atherton, D.B., 2017. Ergonomic Nasal Cannula. US 20170007794 A1.
131. Moore, J.K., 2017. Nasal filter structure. US 9132300 B2.
132. Hioki, K. and H. Yamada, 2014. Nasal cavity insertion device fixture and nasal cavity insertion device set including the same. US 20140094840 A1.
133. Tanaka, A., T. Furubayashi, Y. Enomura, T. Hori, R. Shimomura, C. Maeda, S. Kimura, D. Inoue, K. Kusamori, H. Katsumi, T. Sakane and A. Yamamoto, 2017. Nasal Drug Absorption from Powder Formulations: Effect of Fluid Volume Changes on the Mucosal Surface. Biol. Pharm. Bulletin, 40: 212-219.
134. Pozzoli, M., P. Rogueda, B. Zhu, T. Smith, P.M. Young, D. Traini and F. Sonvico, 2016. Dry Powder Nasal Drug Delivery: Challenges, Opportunities and a study of the commercial Teijin Puvlizer Rhinocort® device and formulation. Drug Development and Industrial Pharmacy.
135. Obaidi, M., E. Offman, J. Messina, J. Carothers, G. Djupesland and R.A. Mahmoud, 2013. Improved Pharmacokinetics of Sumatriptan With Breath Powered™ Nasal Delivery of Sumatriptan Powder. Headache, 53: 1323-1333.
136. Alsarra, I.A., A.Y. Hamed, G.M. Mahrous and G.M.E. Maghraby, 2009. Mucoadhesive Polymeric Hydrogels for Nasal Delivery of Acyclovir. Drug Development and Ind Pharmacy, 35: 352-362.
137. Rathnam, G., N. Narayanan and R. Ilavarasan, 2008. Carbopol-Based Gels for Nasal Delivery of Progesterone. Am. Assoc. of Pharm. Science, 9: 1078-1082.
138. Rogol, A.D., N. Tkachenko and N. Bryson, 2015. Natesto™, a novel testosterone nasal gel, normalizes androgen levels in hypogonadal men. Am Soc of Andrology and Eur Acad of Andrology, 4: 46-54.
139. Tas, C., C.K. Ozkan, A. Savaser, Y. Ozkan, U. Tasdemir and H. Altunay, 2009. Nasal administration of metoclopramide from different dosage forms: in vitro, ex vivo and in vivo evaluation. Drug Delivery, 16: 167-175.
140. Dukovski, B.J., I. Plantic, I. Cuncic, I. Krtalic, M. Juretic, I. Pepic, J. Lovric and A. Hafner, 2017. Lipid/alginate nanoparticle-loaded in situ gelling system tailored for dexamethasone nasal delivery. Int. J. Pharmaceutics, pp: 1-30.
141. Duchateau, G.S., 1987. Studies on nasal drug del. Pharm Weekbl Science, 9: 326-8.
142. Chudiwal, S. and M.H.G. Dehghan, 2016. Quality by Design Approach for Development of Suspension Nasal Spray Products: A Case Study on Budesonide Nasal Suspension. Drug Dev and Ind Pharmacy.
143. Aggarwal, R., A. Cardozo and J.J. Homer, 2004. The assessment of topical nasal drug distribution. Clin Otolaryngol Allied Science, 29: 201-205.
144. Suman J.D., 2003. Nasal Drug Delivery. Exprt Opin Biol Therapeutic, 3: 519-523.
145. Djupesland, P.G., J.C. Messina and R.A. Mahmoud, 2013. Breath Powered Nasal Delivery: A New Route to Rapid Headache Relief. Headache, 53: 72-84.
146. Jogani, V., K. Jinturkar, T. Vyas and A. Misra, 2008. Recent Patents Review on Intranasal Administration for CNS Drug Delivery. Drug Del Formulation, 2: 25-40.
147. Ishikawa, F., M. Murano, M. Hiraishi, T. Yamaguchi, I. Tamai and A. Tsuji, 2002. Insoluble Powder Formulation as an Effective Nasal Drug Delivery System. Pharm Research, 19: 1097-1104.
148. Huang, Y. and M.D. Donovan, 1998. Large molecule and particulate uptake in the nasal cavity: the effect of size on nasal absorption. Advanced drug delivery reviews, 29: 147-155.
149. Peek, L.J., C.R. Middaugh and C. Berkland, 2008. Nanotechnology in vaccine delivery. Advanced Drug Delivery Reviews, 60: 915-928.
150. Lochhead, J.J. and R.G. Thorne, 2014. Intranasal Drug Delivery to the Brain. In: M. Hammarlund-Udenaes et al. eds. Drug Delivery to the Brain, AAPS Advances 401 in the Pharm. Sci. Series, 10: 401-431.

151. Sakane, T., M. Akizuki, Y. Taki, S. Yamashita, H. Sezaki and T. Nadai, 1995. Direct Drug Transport from the Rat Nasal Cavity to the Cerebrospinal Fluid: the Relation to the Molecular Weight of Drugs. *J. Pharm. Pharmacology*, 47: 379-381
152. Chatterjee, B., 2017. Nose to Brain Drug Delivery: A Recent Update. *J Formul Sci Bioavailability*, 1: 105.
153. Arora, P., S. Sharma and S. Garg, 2002. Permeability issues in nasal drug delivery. *Drug Discovery Today*, 7: 967-975.
154. Mitra, R., I. Pezron, W.A. Chu and A.K. Mitra, 2000. Lipid emulsions as vehicles for enhanced nasal delivery of insulin. *Int J Pharmaceutics*, 205: 127-134. Not found in the text, Revise??.