

## Oxidative Stress Status in Type 2 Diabetic Patients in Eastern Algeria

<sup>1</sup>Mouad Benrebai, <sup>1</sup>Nacera Abidli, <sup>2</sup>Soad M. Nasr and <sup>3</sup>Cherefa Benlatreche

<sup>1</sup>Département des Sciences de la Nature et de la Vie. Université Mentouri,  
Constantine 25000, Algeria

<sup>2</sup>Department of Parasitology and Animal Diseases, Veterinary Division,  
National Research Center, Dokki, Cairo, Egypt

<sup>3</sup>Laboratoire de biochimie, CHU Ibn Badis, Constantine 25000, Algeria

**Abstract:** The present study was carried out to evaluate the oxidative status in Urban Algerian patients suffering from type 2 diabetes mellitus (T2DM) and treated with hypoglycaemic agents. Investigations had been conducted on 29 adult T2DM patients as compared with 11 healthy adult subjects (as control). Plasma glucose level and haemoglobin A1C (HbA1C) were determined, as well as total antioxidant capacity (TAC), catalase (CAT), glutathione reduced (GSH) and lipid peroxides (MDA). Kidney function tests were also assessed depending upon blood urea and creatinine values. The result revealed that fasting plasma glucose level was higher ( $P<0.01$ ) and HbA<sub>1</sub>C was markedly increased ( $P<0.05$ ) in T2DM group as compared to healthy subjects.. Patients revealed higher concentration of MDA ( $P<0.01$ ) and low activity of CAT ( $P<0.05$ ) as compared to the control group. GSH and TAC concentrations decreased ( $P<0.001$ ) in the tested group than the healthy control group. A negative correlation was found between fasting plasma glucose levels, HbA<sub>1</sub>C on one hand and TAC, CAT, GSH, MDA values in T2DM patients on the other hand. no alteration in kidney functions was found as indicated by urea and creatinine values. It was concluded that T2DM patients are undergo an important oxidative stress, even under hypoglycaemic control, they were considered to be poorly controlled.

**Key words:** Type 2 diabetes mellitus · Oxidative stress · Total antioxidant capacity · Catalase · GSH · MDA · Algeria

### INTRODUCTION

Investigations on diabetes mellitus had showed the involvement of reactive oxygen species (ROS) including free radicals in the genesis of chronic complications related to the disease, as cardiovascular affections, renal failure and neurodegenerative changes [1-3]. Oxidative stress is more obvious in type 2 diabetes and this appears to underlie the development of diabetic complications [4]. Hyperglycemia leads to metabolic disorders, characterized by alterations in the metabolism of carbohydrate, protein and lipid. Diabetes induced disturbance in lipid profiles, especially an increased susceptibility to lipid peroxidation [5], which is responsible for increased incidence of atherosclerosis [6]; a major complication of diabetes mellitus. Macrovascular complications, which manifest in about 80 percent of patients with type 2 diabetes, are a leading cause of morbidity and mortality worldwide [7].

In non-insulin-dependent (type 2 diabetes mellitus), oral hypoglycaemic agents are used to stimulate the pancreatic beta cells to secrete insulin and/or increase the sensitivity of peripheral insulin receptors to the action of endogenous insulin [8] with the hope of achieving better glycaemic control and attenuating related complications.

Among hypoglycaemic agents; Rosiglitazone used currently by type 2 diabetic patients due to its efficacy for improving the sensitivity of tissues to insulin and reduces insulin resistance [9,10], which in turn improves markers; HbA<sub>1</sub>c is the most representative marker of hyperglycemia for assessing glycaemic control and glycated albumin) of cardiovascular complication [11,12,13]. Metformin, is another hypoglycaemic drug used in clinical treatment of type-2 diabetes for over 35 to 40 years, it enhances the sensitivity of both hepatic and peripheral tissues to insulin, it inhibits gluconeogenesis in the liver and lowering plasma triglyceride and low-density

lipoprotein (LDL) cholesterol and total lipid levels. This drug is most often associated with sulphonylurea to exert an evident lowering effect, on blood glucose level [14,15].

Previous studies have shown that diabetes mellitus can worsen antioxidant status, hence deficiencies in some vitamins and trace elements related to the extrinsic antioxidants, such as vitamin C [16, 17] vitamin E [18] and zinc [19] can aggravate the several complications of diabetes.

This study was conducted to evaluate the oxidative status in a group of Urban Algerian with type 2 diabetes mellitus (T2DM) treated with hypoglycaemic agents (non-insulin-dependent diabetes mellitus) compared to a normoglycemic group.

## MATERIALS AND METHODS

**Subjects:** Twenty-nine patients suffering from T2DM including 20 males and 9 females with a mean age of  $57.57 \pm 2.39$  (range 35-82) years were randomly selected from the Center of Diabetology Al-Kantara, Constantine, Algeria. Patients were informed of the purposes of the study. Eleven age and gender-matched healthy subjects including 5 males and 6 females with a mean age of  $49.80 \pm 4.49$  (range 27-73) years, who came to the center for check up, with no known family history of T2DM, were also enrolled to the study as the control group.

Key exclusion criteria included smokers, pregnant women, persons receiving trace element or antioxidants supplements in the previous three months, persons with gastric or diuretic treatment, patients with acute renal failure (creatinine  $>1.20$  mg/dl) and patients with a recent surgery or acute infection. Only patients with fasting blood glucose  $\geq 150$  mg/dl and HbA1c  $\geq 7.5\%$  were included.

**Blood Samples and Biochemical Assays:** Blood samples were drawn from each T2DM patient and control healthy subject after an overnight fast at the Center of Diabetology Al-Kantara, Constantine. Three blood samples were collected from each subject. The first sample was collected in a tube containing sodium fluoride for plasma glucose determination. The second sample was collected in a tube containing heparin for estimation of plasma antioxidants, creatinine and urea. The third sample was anticoagulated by ethylenediamine tetraacetic acid (EDTA) and was used for determination of HbA1c. Fasting plasma glucose was estimated enzymatically using glucose oxidase [20], creatinine value using Semi-micromethod [21] and urea using urease enzyme [22]

were determined using an Auto-analyser ; ADVIA 1650 *Chemistry System Bayer-Diagnostic* Laboratory of biochemistry CHU IBN BADIS Constantine. Glycated haemoglobin (HbA1c) was measured using *BIO-RAD D-10™ UNITED STATES, Bio-Rad* Laboratories, Inc Hercules CA 94547. The D-10 Hemoglobine A<sub>1</sub>C program utilises principles of ion-exchange high performance liquid chromatography (HPLC).

**Antioxidant Markers Assessment:** Plasma lipid peroxidation product (MDA) and antioxidants status were measured using specific kits purchased from *Bio-diagnostic*, Dokki, Egypt.

Plasma MDA was determined according to the method of [23]. This technique is based on thiobarbituric acid reacts with MDA in acidic media at temperature of  $95^{\circ}\text{C}$  for 30min. to form thiobarbituric reactive product the absorbance of the resultant pink product can be measured at 534 nm.

Plasma glutathione reduced (GSH) was estimated according to the method of [24]. The method depends on the reduction of 5,5'-dithiobis (2-nitrobenzoic acid) (DTNB) with glutathione to produce a yellow compound. The reduced chromogen directly proportional to (GSH) concentration and its absorbance can be measured at 405nm.

Plasma total antioxidant capacity (TAC) was determined according to the method of [25]. This determination is performed by the reaction of antioxidants in the plasma with a defined amount of exogenously provide hydrogen peroxide ( $\text{H}_2\text{O}_2$ ). The antioxidants in the sample eliminate a certain amount of the provided  $\text{H}_2\text{O}_2$ . The residual  $\text{H}_2\text{O}_2$  is determined by an enzymatic reaction which involves the conversion of 3,5, dichloro-2-hydroxybenzenesulphonate to a coloured product and read at 505nm.

Plasma catalase (CAT) was estimated by the method of [26]. The method depends on catalase reacts with a known quantity of  $\text{H}_2\text{O}_2$ . The reaction is stopped after exactly one minute with catalase inhibitor. The presence of peroxidase, remaining  $\text{H}_2\text{O}_2$  reacts with 3,5 Dichloro-2 hydroxybenzene sulfonic acid and 4-Aminophenazone to form a chromophore with a colour intensity inversely proportional to the amount of CAT in the sample and read at 510nm.

**Statistical Analysis:** All data were subjected to statistical analysis including the calculation of the mean and standard error (mean $\pm$ SE). Student *t*-test was used for the evaluation of data. Differences were considered

significant at  $P < 0.05$  level [27], using SPSS version 10 computer programme.

### RESULTS

The individual characteristics; age, gender and duration of the disease (in diabetic patients) and healthy subjects are shown in Table 1.

Fasting blood glucose level of T2DM patients was higher ( $P < 0.01$ ) than that in healthy adult subjects. Also, the HbA1c exhibited clear increase ( $P < 0.05$ ). There was no significant difference in the level of plasma creatinine and urea in T2DM patients and healthy adult subjects during the period study (Table 2, Fig. 1).

The plasma lipid peroxidation end product, malondialdehyde (MDA) increased in T2DM patients

Table 1: Age, gender and duration of diabetes in healthy subjects and type 2 diabetes mellitus patients. (Mean±SE)

Parameters	Groups	
	Healthy subjects (Control, n=11)	Type 2 Diabetes (n=29)
Age (years)	49.80±4.49 (27.00-73.00)	57.57±2.30 (35.00-82.00)
Gender (Male/Female)	5/6	20/9
Duration of diabetes (years)	Non	1-20

SE = Standard error.

Table 2: Values of some plasma biochemical parameters in healthy subjects and type 2 diabetes mellitus patients (Mean±SE)

Parameters	Groups	
	Healthy subjects (Control, n=11)	Type 2 Diabetes mellitus (n=29)
Glucose (mg/dl)	95.27±7.18 (73.00-151.00)	154.52±11.13** (76.00-277.00)
HbA <sub>1c</sub> (g %)	6.27±0.23 (4.92-7.60)	7.96±0.39* (5.00-12.60)
Urea (mg/dl)	36.70±2.54 (26.00-54.00)	39.79±2.19 (21.00-71.00)
Creatinine (mg/dl)	1.14±0.06 (0.92-1.48)	1.12±0.03 (0.90-1.61)

SE = Standard error. HbA<sub>1c</sub> = glycated hemoglobin. \* =  $P < 0.05$ . \*\* =  $P < 0.01$ .

Table 3: Values of some serum oxidative stress markers in healthy subjects and type 2 Diabetes mellitus patients (Mean±SE)

Parameters	Groups	
	Healthy subjects (Control, n=11)	Type 2 Diabetes mellitus (n=29)
Lipid peroxides (nMol/ml)	42.87±1.37 (29.47-45.26)	51.21±1.61** (34.74-71.58)
Glutathione reduced (mMol/l)	7.99±0.93 (4.44-11.10)	4.45±0.25*** (2.22-6.66)
Total antioxidant capacity (mMol/l)	3.81±0.15 (2.71-4.59)	2.91±0.09*** (1.95-3.95)
Catalase activity(Unit/l)	317.81±18.34 (262.73-490.62)	283.80±5.78* (231.90-356.56)

SE = Standard error. \* =  $P < 0.05$ . \*\* =  $P < 0.01$ . \*\*\* =  $P < 0.001$

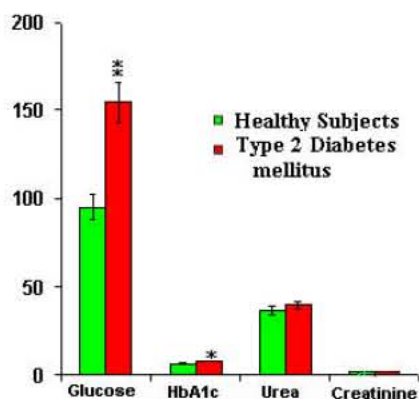


Fig. 1: Plasma glucose (mg/dl), glycated haemoglobin (HbA<sub>1c</sub>), plasma urea (mg/dl) and creatinine (mg/dl) in healthy subjects (control) (N=11) and type 2 Diabetes mellitus patients (\* =  $P < 0.05$ . \*\* =  $P < 0.01$ )

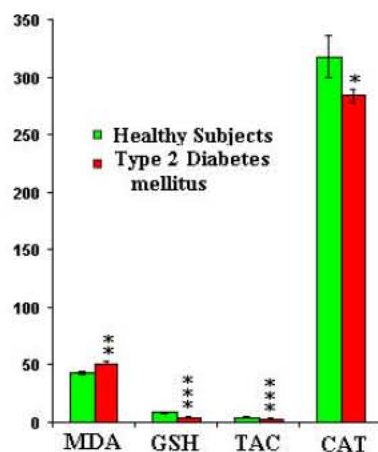


Fig. 2: Plasma lipid peroxides; MDA (nMol/ml), glutathione reduced; GSH (mMol/l), catalase activity; CAT (U/l) and total antioxidant capacity; TAC (mMol/l) in healthy subjects (control) (N=11) and type 2 Diabetes mellitus patients (\* =  $P < 0.05$ . \*\* =  $P < 0.01$ . \*\*\* =  $P < 0.001$ )

( $P < 0.01$ ) as compared with the control group, while, the activity of plasma CAT was decreased in T2DM patients ( $P < 0.05$ ). GSH ( $P < 0.001$ ) and the total antioxidant capacity (TAC) ( $P < 0.05$ ) decreased in T2DM group as compared to the control group (Table 3, Fig. 2).

### DISCUSSION

Fasting plasma glucose level was a high ( $P < 0.01$ ) in T2DM group, even they took hypoglycaemic agents as

compared with healthy subjects. Although this condition is consequently accompanied with a marked elevation of HbA<sub>1c</sub> ( $P < 0.05$ ), indicating that glycaemic in our diabetic population was moderately well-balanced, which explains the low rate of non-enzymatic glycation of haemoglobin A<sub>1c</sub> observed essentially in diabetes.

Oxidative stress status of patients with T2DM was evaluated by measuring plasma lipid peroxidation end product MDA, an important index marker of the extent of lipid peroxidation and evaluation of oxidative stress, as well as glutathione reduced (GSH) concentration, total anti-oxidant capacity (TAC) and the catalase enzyme activity. In this study, oxidative stress status was disturbed in diabetic patients with T2DM as compared to healthy subjects.

Plasma MDA concentration was higher in T2DM patients as compared with healthy control subjects, indicating higher lipid peroxidation [28]. The elevation of lipid peroxidation is related to the duration of diabetes [29,30]. This was consistent with the previous studies [31,32] especially in patients with vascular complications [33,34]. Thus, moderate, as in the current case, or poor diabetes control may enhance lipid peroxidation and diminishes the body's anti-oxidant capacity, hence a negative correlation between hyperglycaemic and oxidative stress was observed and agree with [35]. This increase of lipid peroxidation is frequently observed in diabetes due to mobilization of lipids for a further use as an energy source rather than glucose.

Plasma GSH, decreased in T2DM patients as compared to healthy control subjects which indicates the extent degree of oxidative stress in diabetes [36,37], it accompanies the decrease in the activity of catalase, which plays a co-enzyme role for scavenging H<sub>2</sub>O<sub>2</sub>. On the other hand, the decrease of plasma glutathione reduce may be due also, to a decrease in ascorbic acid concentration which plays a synergic role with GSH in the regeneration of vitamin E, during the elimination of free radicals [38]. The depletion of GSH is due in great part to a deficiency of NADPH<sub>2</sub> used in some oxido-reduction reactions like polyol pathway over and above its reduction formation through pentose phosphate and malic acid pathways owing to chronic hyperglycaemic as showed by [39] and [40], leading to impairment of GSH regeneration and depletion of an important free radical scavenger [41], this condition disturbs the antioxidant defenses and accelerate the oxidative damage [42] and hence reconfirming the negative effect of hyperglycaemic on GSH levels.

The TAC, represents the extrinsic (micronutrients) trace elements, vitamins (A, E,  $\beta$ -caroten and ascorbic acid) and intrinsic factors including group of organic anti-oxidants such as enzyme catalase glutathion peroxidase, superoxide dismutase and non-enzymatic anti-oxidants (GSH) and others like flavonoids, bilirubin and uric acid [43]. In this study, a very drastic decrease in plasma TAC in T2DM patients vs the control group was observed and agree with [37, 44], but less than that reported by [32]. In this respect, [45] reported that this diminution is due to poor control diabetes, which concurs truly with the disturbance in HbA<sub>1c</sub> [46].

Treatment of T2DM patients with hypoglycaemic agents is often followed by a strict regimen, which in turn must be well adapted to each diabetic patient, to avoid depletion in extrinsic antioxidants, that were not be influenced by auto-glycation as it's the case with antioxidant proteins.

Catalase (CAT) is a hemeprotein catalysing the reduction of hydrogen peroxides and protects against highly reactive hydroxyl radicals; decreased CAT activity during diabetes disease could result from inactivation by glycation of enzyme [47]. The present decrease in CAT activity in T2DM group was 10% ( $P < 0.05$ ), however, [46] observed a diminution by 30 % in type 2 diabetics. Negative correlations between serum glucose level and HbA<sub>1c</sub> and CAT were noticed in this study. The diminution in the catalase activity is also observed in diabetics with poor glucose control and vascular complications. Previous studies have shown that plant hypoglycaemic extracts like *Syzigium cumini* seeds increases catalase activity by diminishing blood glucose level [48] and vitamin C, as a potential antioxidant, also increases catalase activity [49], this may be due to the alleviate action of hypoglycaemic agents on enzyme glycation.

In conclusion, results from this study suggest that hypoglycaemic treatment has no favorable effect on antioxidant system in T2DM patients compared with healthy subjects. This condition suggests that under hypoglycaemic treatment a supplementation with micronutrients is necessary to improve the intrinsic antioxidant system.

#### ACKNOWLEDGMENT

We are grateful to Biochemistry Laboratory team of CHU Ibn Badis Constantine Algeria and Researchers of Veterinary Division of NRC, Dokki, Cairo, Egypt for their helpful works.

## REFERENCES

1. Song, S.H. and C.A. Hardisty, 2008. Early-onset type 2 diabetes mellitus: an increasing phenomenon of elevated Cardiovascular Risk. *Expert Rev. Cardiovascular Therap.*, 6: 315-322.
2. Yokoyama, H., M. Okudaira, T. Otani, H. Takaike, J. Miura, A. Saeki, Y. Uchigata and Y. Omori, 1997. Existence of early onset NIDDM Japanese demonstrating severe diabetic complications. *Diabetes Care*, 20: 844-847.
3. Halliwell, B., 2001. Role of free radicals in the neurodegenerative diseases: therapeutic implications for antioxidant treatment. [Review]. *Drugs Aging*, 18: 685-716.
4. Choi, S., I.F.F. Benzie, S. Ma, J.J. Strain and B.M. Hannigan, 2008. Acute hyperglycemia and oxidative stress: Direct cause and effect? *Free Radical Biol. Med.*, 44: 1217-1231.
5. Lyons, T.J., 1991. Oxidized low density lipoproteins: a role in the pathogenesis of atherosclerosis in diabetes? *Diabetic Med.*, 8: 411-419.
6. Giugliano, D., A. Ceriello and G. Paolisso, 1995. Diabetes mellitus, hypertension and cardiovascular diseases: which role for oxidative stress? *Metabolism*, 44: 363-368.
7. Quilliot, D., B. Dousset, B. Guerci, F. Dubois, P. Drouin and O. Ziegler, 2001. Evidence that diabetes mellitus favors impaired metabolism of zinc, copper and selenium in chronic pancreatitis. *Pancreas*, 22: 299-306.
8. Lebovitz, H.E., 2001. Oral therapies for diabetic hyperglycemia. *Endocrinology Metabolism Clinics of North America*, 30: 909-933.
9. Scheen, J., 2001. Pharm-clinics medication of the month, Repaglinide (NovoNorm). *Revue Médicale de Liège*, 65: 456-459.
10. Pospisilova, Y., 2001. New in oral antidiabetic therapy. *Drugs*, 61: 1625-1660.
11. Phillips, L.S., G. Grunberger, E. Miller, R. Patwardhan, E.B. Rappaport and A. Salzman, 2001. Once- and twice-daily dosing with rosiglitazone improves glycemic control in patients with type-2 diabetes. *Diabetes Care*, 24: 308-315.
12. Wagstaff, A.J. and K.L. Goa, 2002. Rosiglitazone: A review of its use in the management of type-2 diabetes mellitus. *Drugs*, 62: 1805-1837.
13. Okada, T., T. Nakao, H. Matsumoto, T. Shino, Y. Nagaoka, R. Tomaru and T. Wada, 2007. Association between markers of glycemic control, cardiovascular complications and survival in type 2 diabetic patients with end-stage renal disease. *Internal Medicine*. 46: 807-814.
14. DeFronzo, R.A., 1999. Pharmacologic therapy for type-2 diabetes mellitus. *Ann. Internal Med.*, 131: 281-303.
15. Setter, S.M., J.L. Iltz, J. Thams and R.K. Campbell, 2003. Metformin hydrochloride in the treatment of type-2 diabetes mellitus: a clinical review with a focus on dual therapy. *Clinical Therapeutics*, 25: 2991-3026.
16. Jennings, P.E., S. Chirico, A.F. Jones, J. Lunec and A.H. Barnett, 1987. Vitamin C metabolites and microangiopathy in diabetes mellitus. *Diabetes Res.*, 6: 151-154.
17. Cunningham, J.J., S.L. Ellis, K.L. McVeigh, R.E. Levine and J. Calles-Escandon, 1991. Reduced mononuclear leucocyte ascorbic acid content in adults with insulin-dependent diabetes mellitus consuming adequate dietary vitamin C. *Metabolism*, 40: 146-149.
18. Karpen, C.W., S. Cataland, T.M. O'Dorisio and R.V. Panganamala, 1985. Production of 12-hydroxyeicosatetraenoic acid and vitamin E status in platelets from type 1 human diabetic subjects. *Diabetes*, 34: 526-531.
19. Saxena, R., S.V. Madhu, R. Shukla, K.M. Prabhu and J.K. Gambhir, 2005. Postprandial hypertriglyceridemia and oxidative stress in patients of type 2 diabetes mellitus with macrovascular complications. *Clinica Chimica Acta*, 359: 101-108.
20. Trinder, P., 1969. Determination of glucose in blood using oxidase with an alternative oxygen acceptor. *Ann. Clin. Biochem.*, 6: 24-27.
21. Wahlefeld, A.W., G. Holz, Dans H.U. Bergmeyer and H.U. Bergmeyer, 1974. Methoden der enzymatischen Analyse. 3rd Edn. tome II, Verlag chemie, Weinheim, pp: 1834-1838.
22. Gutmann, I., H.U. Bergmeyer and Dans H.U. Bergmeyer, 1974. Methoden der enzymatischen Analyse. 3rd Edn. tome II, Verlag chemie, Weinheim, pp: 1842.
23. Satoh, K., 1978. Serum lipid peroxide in cerebrovascular disorders, determined by a new colorimetric method. *Clinica Chimica Acta*, 90: 37-43.
24. Beutler, E., O. Duron and B.M. Kelly, 1963. Improved method for the determination of blood glutathione. *J. Lab. Clin. Med.*, 61: 882-888.
25. Koracevic, D., G. Koracevic, V. Djordjevic, S. Andrejevic and V. Cosic, 2001. Method for the measurement of antioxidant activity in human fluids. *J. Clin. Pathol.*, 54: 356-361.
26. Aebi, H., 1984. Catalase *in vitro*. *Methods in Enzymology*, 105: 121-126.
27. Snedecor, G.W. and W.G. Cochran, 1982. *Statistical Methods*. 8<sup>th</sup> Edn., Iowa State University Press, U.S.A.

28. Esterbauer, H., R.J. Schaur and H. Zollner, 1991. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radical Biol. Med.*, 11: 81-128.
29. Hsu, W.T., L.Y. Tsai, S.K. Lin, J.K. Hsiao and B.H.C. Hen, 2006. Effects of diabetes duration and glycemic control on free radicals in children with type 1 diabetes mellitus. *Ann. Clin. Lab. Sci.*, 36: 174-178.
30. Firoozrai, M., M. Nourbakhsh and M. Razzaghy-Azar, 2007. Erythrocyte susceptibility to oxidative stress and antioxidant status in patients with type 1 diabetes. *Diabetes Res. Clin. Practice*, 77: 427-432.
31. Gallou, G., A. Ruelland, B. Legras, D. Maugendere, H. Allanic and L. Cloarec, 1993. Plasma malondialdehyde in type I and type II diabetic patients. *Clinica Chimica Acta*, 214: 227-234.
32. Thompson, K.H. and D.V. Godin, 1995. Micronutrients and antioxidants in progress in diabetes. *Nutr. Res.*, 15: 1377-1410.
33. Turk, H.M., A. Sevinc, C. Camci *et al.* 2002. Plasma lipid peroxidation products and antioxidant enzyme activities in patients with type 2 diabetes mellitus. *Acta Diabetol.*, 39: 117-122.
34. Kesavulu, M.M., B.K. Rao, R. Giri, J. Vijaya, G. Subramanyam and C. Apparao, 2001. Lipid peroxidation and antioxidant enzyme status in type 2 diabetics with coronary heart disease. *Diabetes Res. Clin. Practice*, 53: 33-39.
35. Ceriello, A., 2005. Postprandial hyperglycemia and diabetes complications: is it time to treat? *Diabetes*, 54: 1-7.
36. Kassab, A., S. Laradi, S. Ferchichi, A. Omezzine, B. Charfeddine, H. Ammar, L. Chaieb and A. Miled, 2003. Oxidative stress parameters in type 2 diabetes mellitus. *Immuno-analyse and Biologie spécialisée*, 18: 79-85.
37. Feillet-Coudray, C., E. Rock, C. Coudray, K. Grzelkowska, V. Azais-Braesco, D. Dardevet and A. Mazur, 1999. Lipid peroxidation and antioxidant status in experimental diabetes. *Clinica Chimica Acta*, 284: 31-43.
38. Murray, R.K., D.K. Grammer, P.A. Mayes and V.W. Rodwell, 1993. *Harper's Biochemistry*. 23<sup>rd</sup> Edn. A LANGE Medical Book.
39. Varvarovska, J., J. Racek, R. Stetina, J. Sykora, R. Pomahacova, Z. Rusavy *et al.*, 2004. Aspects of oxidative stress in children with type 1 diabetes mellitus. *Biomed. Pharmacother.*, 58: 539-545.
40. Bonnefont-Rousselot, D., J.P. Bastard, M.C. Jaudon and J. Delattre, 2000. Consequences of the diabetic status on the oxidant/antioxidant balance. *Diabetes and Metabolism*, 26: 163-176.
41. Travis, S.F., A.D. Morrison, R.S. Jr. Clements, A.I. Winegrad and F.A. Oski, 1971. Metabolic alterations in the human erythrocyte produced by increases in glucose concentration: the role of the polyol pathway. *J. Clin. Invest.*, 50: 2104-2112.
42. Dominguez, C, E. Ruiz, M. Gussinye and A. Carrascosa, 1998. Oxidative stress at onset and in early stages of type 1 diabetes in children and adolescents. *Diabetes Care*, 21: 1736-1742.
43. Irshad, M. and P.S. Chaudhuri, 2002. Oxidant-antioxidant system: role and significance in human body. *Indian J. Exp. Biol.*, 40: 1233-1239.
44. Tanaka, Y., P.O.T. Tran, J. Harmon and R.P. Robertson, 2002. A role for glutathione peroxidase in protecting pancreatic  $\beta$  cells against oxidative stress in a model of glucose toxicity. *Proceed. Nat. Acad. Sci.*, 99: 12363-12368.
45. Firoozrai, M., M. Nourbakhsh and M. Razzaghy-Azar, 2007. Erythrocyte susceptibility to oxidative stress and antioxidant status in patients with type 1 diabetes. *Diabetes Res. Clin. Practice*, 77: 427-432.
46. Komosińska-Vashev, K, K. Olczyk, P. Olczyk and K. Winsz-Szczotka, 2005. Effects of metabolic control and vascular complications on indices of oxidative stress in type 2 diabetic patients. *Diabetes Res. Clin. Practice*, 68: 207-216.
47. Yan, H. and J.J. Harding, 1997. Glycation-induced inactivation and loss of antigenicity of catalase and superoxide dismutase. *Biochem. J.*, 328: 599-605.
48. Prince, P.S.M., V.P. Menon and L. Pari, 1998. Hypoglycaemic activity of *Syzygium cumini* seeds: Effect on lipid peroxidation in alloxan diabetic rats. *J. Ethnopharmacol.*, 61: 1-7.
49. Santos, L.F.L., R.L.M. Freitas, S.M.L. Xavier, G.B. Saldanha and R.M. Freitas 2008. Neuroprotective actions of vitamin C related to decreased lipid peroxidation and increased catalase activity in adult rats after pilocarpine-induced seizures. *Pharmacol. Biochem. Behav.*, 89: 1-5.