International Journal of Microbiological Research 11 (3): 111-121, 2020 ISSN 2079-2093 © IDOSI Publications, 2020 DOI: 10.5829/idosi.ijmr.2020.111.121

Immunity to Fungal Infections: Review Article

Yitbarek Habtamu

National Institute for Control and Eradication of Tsetse Fly and Trypanosomosis, Addis Ababa, Ethiopia, P.O. Box: 19917

Abstract: Most fungi to which we are regularly exposed through the air or because they are part of our normal microbiota do not usually cause disease in immunocompetent individuals. However, some fungi rarely become pathogenic if host defenses breached or compromised with symptoms ranging from mild superficial infections to severe systemic diseases that are associated with a high degree of morbidity and mortality. Protective immunity against fungal pathogens achieved by the integration of two distinct arms of the immune system, the innate and adaptive responses. Innate and adaptive immune responses intimately linked and controlled by sets of molecules and receptors that act to generate the most effective form of immunity for protection against fungal pathogens. The way to respond primarily determined by interactions between pathogens and cells of the immune system of the host, but the actions of T cells will feedback into this dynamic equilibrium to regulate the balance between tolerogenic and inflammatory responses. However, the host responses against different fungal infections are as diverse and distinct as the different fungal diseases themselves. Clinical and basic research during the last decade has brought exciting new insights into the pathogenesis of fungi and revealed important molecular and cellular players in host-fungal interactions and host defense. However, commensal and normally non-pathogenic environmental fungi can cause life-threatening infections in immunocompromised individuals. Therefore, the objective of this review is to insight the mammalian immune responses against fungi infections.

Key words: Fungi · Infection · Immune Responses

spherical, such as Candida species, or multicellular and blood vessels by a growing fungus chokes off the filamentous, like Aspergillus. Some fungi live blood supply to the host tissue, damaging or killing it [2]. commensally on the topologically external surfaces of The exceptions are the dermatophytes, filamentous fungi the body, while others live most of their lives in the soil that infect the skin, hair and nails [2]. These organisms, as a mass (mycelium) of thread-like processes (hyphae). which include species of Epidermophyton, Microsporum Dimorphic fungi adopt a yeast-like form at one stage in and Trichophyton, cannot penetrate the living, cellular their life cycle and a hyphal form at another stage. tissue of a healthy host and so are restricted to parts of Fungal cells have a cell wall like bacteria but also cell the body that lack living cells, such as the keratinized membrane-like mammalian cells [1]. However, the fungal outer layer of skin. Important fungal pathogens are cell wall lacks the peptidoglycans, teichoic acids and *Blastomyces dermatitidis, Histoplasma capsulatum,* lipopolysaccharide components of the bacterial wall *Candida species, Aspergillus species, Cryptococcus* and the main component of the fungal cell membrane is *neoformans and Pneumocystis carinii* [1]. Blastomycosis ergosterol rather than the cholesterol found in mammalian occurs when conidia of the yeast-like fungus cell membranes. *B. dermatitidis* inhaled through the aerosol.

infection and clinical fungal-related disease has risen pulmonary infection that spreads through the blood to dramatically in the last two decades, which would suggest the skin, bones and male urogenital tract, but not an increasing pool of susceptible, immunocompromised the gut. In contrast, *H. capsulatum* is an intracellularly

INTRODUCTION individuals [2]. However, immunocompromised Fungi tend to be either unicellular (yeast-like) sometimes go on to become persistent. Invasion of The correlation between the incidence of fungal This organism replicates extracellularly to cause a individuals can suffer from acute infections that

Corresponding Author: Yitbarek Habtamu, National Institute for Control and Eradication of Tsetse fly and Trypanosomosis, Addis Ababa, Ethiopia. P.O. Box: 19917.

Inhaled microconidia develop into Histoplasma that take second-tier, adaptive response. up residence preferentially in local respiratory Decades ago, fungal immunology research largely macrophages [2]. focused on defining the molecular interactions between

tuberculosis can spread to the secondary lymphoid their cognate pattern recognition receptors (PRRs) and organs, mucosae, gut and adrenal glands. Candida was beginning to understand how immune cells could species such as *C. albicans* and *C. tropicalis* lurk in the interact with fungi as some of the molecules involved in normal flora at the mucosae (but not in the skin) and fungal recognition were discovered. Dendritic cells (DCs) cause disease only if these mucosal barriers are broken found to act at different levels in the immune response or compromised [1]. A deficiency of neutrophils in the against fungi [4]. They are not only able to mount an host (neutropenia) leaves the host especially immediate innate immune response by producing vulnerable to candidiasis. Such Candida infections are inflammatory mediators; they could also influence usually superficial in nature (such as vaginitis and subsequent adaptive immune responses, including cystitis) but can progress to infections of the eye, skin tolerance to commensal organisms. In fact, fungi are and brain [3]. associated with an extensive variety of diseases in

variety of diseases and can induce allergic responses. limiting pulmonary manifestations in immunocompetent Conidia of three species, *A. fumigatus*, *A. flavus* and individuals to inflammatory diseases and severe life-*A. niger*, are particularly pathogenic for humans, causing threating infections in immunocompromised patients [7]. invasive pulmonary infections that can be fatal if allowed Therefore, the objective of this review is to insight the to entrench [2, 3]. Aspergillus species also produce mammalian immune responses against fungi infections. mycotoxins that damage hepatocytes, macrophages and CTLs. *C. neoformans* is a yeast-like fungus often present **Evasion Strategies:** Invasive fungal infections caused by in pigeon droppings. When a host inhales Candida species or Aspergillus fumigatus and other unencapsulated spores of Cryptococcus, the parasite filamentous moulds are devastating in immune enters the lung and synthesizes a protective capsule that compromised patients [7]. Many fungi adopt different inhibits phagocytosis. If the infection becomes forms at different stages in their life cycles, making established, the result may be cryptococcosis, a immune defense necessarily more complex. The structure syndrome of pulmonary infection accompanied by of the fungal cell wall and membrane means that

(innate or adaptive) vary depending on the fungal species offset the effector actions of neutrophils, macrophages, encountered [5] the target organism and the site of CTLs and NK cells [8]. If fungal pathogens overcome the infection. Survival within phagocytes from where fungi initial epithelial barrier and (start to) invade the host can later disperse throughout their host is one particular tissue, they may get in contact with immune cells in the elegant strategy [1, 6]. To maintain a stable host-fungi tissue and/or depending on the route and degree of interaction, the immune response segregated into an invasion in the circulation [8].

replicating yeast-like fungus that causes histoplasmosis. innate first-line defense, which latterly strengthened by a

A progressive pulmonary disease resembling pathogen-associated molecular patterns (PAMPs) and Inhalation of spores of Aspergillus species causes a mammals, ranging from cutaneous lesions and acute self-

meningitis [3, 4]. **Fungi** generally can avoid complement-mediated lysis. The mechanistic aspects of immune responses In addition, many fungi have developed strategies to

Table: Fungal mechanisms to evade the host immune

I able. Fullgal mechanisms to evade the host immune	
Immune system element thwarted	Fungal mechanisms
PPR recognition	Have no LPS or peptide glycan in cell wall
Specificity of T and B cells	Have a multi stage life cycle
Complement	Block access to the cell membrane via cell wall
Phagocytosis	Block phagocytosis via polysaccharide capsule
T and B cell function	Induce immune deviation to Th ₂
Block NF-KB activation	Increase NO production to decrease lymphocyte proliferation
	Block phagocytosis
	Inhibit neutrophils migration
	Decrease IL-12 and B7 expression by monocytes
	Activate regulatory T- cell via polysaccharide capsule component
	Produce melanin to decrease Th1 and Th2 responses
	Block TNF production

barrier of skin or mucosa greeted by the innate immune fungal species [24]. B-1, 3-glucan, which is usually hidden system; second line of defense. Immunologists refer this by other carbohydrates, is one of the most potent fungal system asinnate, which is a defense mechanism that all PAMPs. The sequestration of β -1, 3-glucan by less mammalians naturally seem to have. The innate immune immune-stimulatory cell wall components evolved as a system has evolved to sense conserved microbial potent immune evasion strategy in many pathogenic structures, so-called pathogen- associated molecular fungi [25-29]. patterns (PAMPs) via germline-encoded PRRs. Ligand binding by PRRs induces the activation of signaling **C-Type Lectin Receptors (CLR):** The main PRRs involved cascades inside the cell that leads to gene expression in in fungal recognition belong to the family of myeloid the nucleus. The production of cytokines, chemokines, C-type lectin receptors (CLRs) that are highly expressed complement units and antimicrobial factors by innate by dendritic cells (DCs), neutrophils and macrophages. immune cells results in the activation and recruitment of CLRs are classified by the presence of one or several effector cells to the site of infection and in the elimination C-type lectin-like domains (CTLDs), many of which bind of pathogens, respectively [7]. Innate immune cells to carbohydrates such as those in the fungal cell wall [30]. including neutrophils, monocytes/macrophages and The family of CLRs encompasses soluble molecules such dendritic cells rapidly detect the presence of fungi and as the mannose-binding protein (MBP) or surfactant induce an antimicrobial response. Neutrophils, protein D that activate the complement cascade, macrophages and DCs are all critical to the antifungal endocytic receptors that internalize their ligands such as response [9-11]. The release of inflammatory cytokines, as the mannose receptor (MR) and signaling receptors that well as reactive oxygen intermediates and antimicrobial act as bona fide PRRs to initiate innate and adaptive peptides, can then clear the fungi in target organs immunity, while others have immunomodulatory activities [6]. Fungal recognition is mediated by a variety of [30-32]. The prototypic signaling CLR is dectin-1. It binds surface-bound and soluble pattern recognition receptors to β -(1, 3)-glucans in the fungal cell wall. Dectin-1 (PRRs) recognizing fungal cell wall components and contains an ITAM-like motif, also called hemITAM, in its nucleic acids including Toll like receptors (TLRs), C-type cytoplasmic domain [33-36]. Selective signaling via dectinlectins (Dectin-1, Dectin-2, Mincle, dendritic cell-specific 1 results in the induction of high levels of TNF, IL-6 and intercellular adhesion molecule-3-grabbing non-integrin, IL-23, but little IL-12 and is thus qualitatively distinct from mannose-binding lectin 2 (MBL2) and long pentraxin 3 TLR- mediated signaling [37]. Dectin-1 signaling can also (PTX3)) [12-22]. engage phospholipase C γ 2 (PLC γ 2)-dependent nuclear

of defence in antifungal immunity. Upon recognition signaling leads to the induction of IL-1 β via the activation of surface-expressed PAMPs such as β -glucan and of caspase-1, which mediates cleavage of pro- IL- 1 β into α -mannan, CLRs initiate inflammatory innate responses mature IL-1 β [39, 40]. Dectin-1 signaling was implicated that in turn polarize a Th17 adaptive immune response, in the response to many pathogenic fungi and its hostwhich is generally beneficial to fungal control and protective role was demonstrated in mouse models clearance [23]. The primary fungal PAMPs recognized by with *C. albicans* [41], *A. fumigatus* [42-44] and the innate immune system of the host are components of Pneumocystis [45-47]. Dectin-2 and mincle are two the cell wall. The fungal cell wall is composed of skeletal additional Syk-coupled CLRs that recognize fungal and matrix components, which resemble mesh and mortar α -mannan structures [48]. Unlike dectin-1, they lack the in concrete buildings [23]. The skeletal structures at the hemiTAM motif in their cytoplasmic tail. Instead, they base of the cell wall consist of chitin, which is a β -(1, 4)- assemble with FcR γ , an ITAM-containing adaptor for linked polymer of N-acetylglucosamine and β -1, 3- and signaling [49]. In addition, dectin-2 and mincle have β -1, 6-glucans that are stabilized by intermolecular recently been shown to associate with MCL (also called hydrogen bonds. The fungal cell wall matrix is composed dectin-3) for enhanced ligand binding [50-53]. of mannoproteins, namely, heavily glycosylated proteins with mannose-containing polysaccharides (mannans). **Mannose-Binding Lectin:** The mannose receptor (MR) These mannoproteins attached to β -1, 3-glucans or chitin is a PRR primarily present on the surface of macrophages by a linker structure. Important differences in cell wall and dendritic cells and thus there are 2 MLB isomers in

Innate Immunity: Any invader that breaches the physical composition and conformation exist between different

PRRs in Fungal Recognition: PRRs are the first line the production of IL-2 and IL-10 [38]. Moreover, dectin-1 factor of activated T cell (NFAT) activation, resulting in

MBL2. Mannose-binding lectin (MBL) is a soluble lectin NOD-like receptor NLRP3 and ASC to provide a scaffold belonging to the collectin family and consists of a CRD for caspase-1 activation. The importance of the NLRP3 that is attached to a collagen region via α -helical coil inflammasome in antifungal immunity was demonstrated domain [4]. It is produced by the liver and secreted into in NLRP3-deficient mice, which display an increased the blood, where, after binding to microbial carbohydrate susceptibility to systemic and superficial candidiasis [64]. surfaces, it can activate the lectin pathway of the Activation of the NLRP3 inflammasome by C. albicans complement cascade, enhancing the phagocytosis of depends on yeast-to-hyphal transition, which may reflect microorganisms and modulating inflammatory responses the dependence on β -glucan exposure at the fungal cell [54-57]. surface [65-67] and the secretion of secreted aspartyl

Responses Intensive research in this area in the last few by C. albicans- derived aspartyl proteases [69-70]. IL-1 β years has identified several fungal PAMPs recognized by is closely related to IL-1 α , which signals through the same TLRs; however, the primary structures of the fungal receptor. IL-1 α also depends on processing for becoming ligands have not yet been fully resolved. Some studies bioactive, but it is not a substrate of caspase- 1 [63]. have suggested that mannosylated structures derived However, the NLRP3 inflammasome and caspase-1 is from Candida, Cryptococcus and Scedosporium can implicated indirectly in the secretion of IL-1 α [71] by directly interact with specific TLRs, including TLR1, catalysing the processing of IL-1 β , which was proposed TLR2, TLR4 and TLR6, triggering inflammatory responses to bind to intracellular IL-1a and to serve as a shuttle for [58]. Besides CLRs, certain TLRs are also implicated in IL-1 α release [72]. Cell surface-bound IL-1 α can be fungal recognition, including TLR2, TLR4, TLR7 and cleaved by calpain I and II at the cell membrane and TLR9. The family of TLRs is the best-characterized family secreted pro-IL-1 α can be processed by extracellular of PRRs. TLRs is membrane-bound receptors composed proteases [73]. IL-1 α , which is constitutively expressed in of leucine-rich repeats for ligand recognition and a some cells such as keratinocytes, is also released upon conserved Toll/IL-1R-domain in the cytoplasmic domain. cytolytic cell death. In addition that caspase-1 has a The latter mediates signaling via MyD88 (and/or TRIF in critical function in the processing of IL-1 β ; caspase- 1 some cases) to couple to NF-êB activation (or IRF activation can also induce pyroptosis, an inflammatory activation in case of TRIF-mediated signaling) and the form of programmed cell death [74]. Pyroptosis results in induction of pro-inflammatory target genes. TLR2 and DNA fragmentation and chromatin condensation. TLR4 are expressed at the cell surface, where they However, in contrast to apoptosis, which is a non-lytic recognize fungal phospholipomannan and O-mannan mechanism, pyroptosis involves cell swelling, porestructures, respectively [59, 60]. The endosomal TLRs mediated lysis and release of intracellular components, TLR7 and TLR9 have also been implicated in fungal which usually are not exposed to the extracellular recognition, namely, in sensing of nucleic acids and compartment, including cytoplasmic cytokines such as induction of IL-12 and/or type I interferons in response to IL-1 α and IL-1 β ATP, HMGB1 and nucleic acids. Due to C. neoformans, A. fumigatus and C. albicans [61]. their inflammatory properties when released in the Moreover, TLRs contribute to antifungal immunity by extracellular environment, these molecules were also modulating the response induced by other PRRs [62]. called damage-associated molecular patterns (DAMPs)

Inflammasome Activation by Fungal Pathogens: Besides identified that promote inflammatory responses. other cytokines and chemokines, many fungal pathogens induce the production of IL-1 β . Biosynthesis of bioactive **Roles of Defensins in Fungal Innate Immunity:** They are IL-1 β requires two independent signals. The first cationic, microbicide peptides active against many Gramregulates transcription and translation of pro-IL-1 β and negative and Gram-positive bacteria, fungi and enveloped the second induces the proteolytic cleavage of pro-IL-1 β viruses composed of three pairs of intermolecular into the active IL-1 β [63]. Fungi trigger both steps of IL-1 β disulphide bonds which are classified in to alpha, beta synthesis. Importantly they induce proteolytic cleavage and theta [76]. Defensin exist in the mammalian white by caspase-1 via the assembly of Inflammasome with blood cells such as macrophages, granulocytes, NK cells distinct subunit composition. *C. albicans* stimulates the and only beta defensins are located in the epithelial cell

the mouse; MBL1 and MBL2, while humans only have assembly of a canonical inflammasome composed of the **Toll-Like Receptors (TLRs):** Ligands and Cellular be processed by neutrophil-derived proteinase-3 [68] and proteinase (SAP) 2 and SAP6 [68]. In addition, IL-1 β can [75] that for some of them, cellular receptors were

fungi microbes that are negative due to that facilitate adherence to host cells and may also lipopolysaccharide and acid encapsulated by the cell promote phagocytosis. Pro-inflammatory cytokines membrane that the peptides have higher affinity to the induced by products of complement activation also binding site compared to Ca2+ and Mg2+ ions. Therefore, contribute to anti-fungal defense [8]. Complement the peptides exchange place with those ions, thus activation plays a central role during bloodstream affecting the stability of the membrane by passing infections [87]. Fungal pathogens rapidly activate the across the membrane due to changes in the electric complement via multiple pathways including the potential and aggregate in to dimers [78]. Finally pore alternative complement pathway [88]. Rapid activation of complex will be created as a result of breaking the the C3 convertase leads to C. albicans opsonization by hydrogen bonds between the amino acids in the terminal C3b binding to β -(1, 6)-glucan [89], which facilitates end of the strands connecting defensins monomers phagocytosis by neutrophils in a CR3-dependent manner causing membrane depolarization and cell lysis [79]. [90]. Importantly, complement activation also results in Defensins not only have the ability to strengthen the the induction of anaphylatoxins C3a and C5a with innate immune system but can also enhance the important immune stimulatory activities, in particular on adaptive immune system by chemo taxis of monocytes, neutrophils and monocytes, which are abundant in the T-lymphocytes, dendritic cells and mast cells to the circulation and they can act as chemo attractants for these infection site that improves the capacity of macrophage cells in tissues [90]. phagocytosis [80].

defends the mucosae, antibodies are thought to cytokine production [1]. Cell-mediated innate immunity is contribute in only a limited way to defense against fungi. the primary means by which fungi infections are Antibody-mediated opsonisation may promote controlled. Neutrophils and macrophages both carry out phagocytosis and thus contribute to the presentation vigorous phagocytosis and produce powerful anti-fungal of fungal antigens that activates Th1 cells [81]. defensins. These defensins induce an osmotic imbalance Antibody-mediated immunity is generally thought to in pathogens such as Candida and Cryptococcus that kills play a minor role for natural protection from fungal them. Neutrophils and macrophages also secrete copious infections. Although antibodies are generated in response quantities of IL-1, IL-12 and TNF. IL-12 stimulation to commensal or environmental fungi and can be detected activates NK cells that contribute to fungal cell killing via in the serum, their specificities are not usually protective cytokine secretion (rather than natural cytotoxicity) [2]. to the host [81]. More recently, it has become clear IFN produced first by activated NK cells and later by that depending on the specificity and isotype, certain activated Th1 cells also hyper activates macrophages, antibodies can modulate the course of fungal which can initiate granuloma formation. Interestingly, a infections and thereby benefit or harm the host [82]. fungus present in its unicellular yeast-like form tends to Antibody- mediated immunity is now viewed as a provoke a protective Th1 response, whereas its hyphal promising therapeutic approach against fungal infections form tends to induce a non-protective Th2 response. and several protective antibodies to fungi have now There is some evidence that either distinct subsets of been developed [82, 83]. Anti-cell wall mannoprotein DCs, or distinct receptors on DCs, respond to the two antibodies can block adhesion of *C. albicans* [84, 85]. different fungal morphologies [1]. These DCs then In contrast, antibodies directed against *C. neoformans* proceed with phagocytosis and antigen processing and glucuronoxylomannan (GXM) protect via enhancing presentation and influencing Th1/Th2 differentiation in cellular immunity including phagocytosis and antibody- the direction best suited to eliminate the particular form of mediated cellular cytotoxicity [82, 83, 86]. the fungus present. The Th1 response induced by

complement cascade, they are generally resistant to mediated by cells producing copious quantities of IL-2 complement-mediated lysis. However, they are subject to and IFN [2]. Th2 responses are comparatively rare during phagocytosis when opsonised by complement products. infections with yeast-like fungi.

[77]. They interact with the membrane of invading Fungi also express analogues of complement receptors

Adaptive Antifungal Mechanisms can induce several cellular responses, including **Humoral Defense:** Other than the secretory IgA that phagocytosis, respiratory burst and chemokine and **Complement:** While fungal cells can activate the mucosal fungal flora that have a yeast-like form, is **Cell Mediated Immunity:** After ligand binding, dectin-1 exposure to airborne fungal spores, or invasion by skin or cells carry antigen receptors that recognize antigenic canprotect mice against cryptococcosis due to the peptides presented in the context of MHC class II induction of robust Th1 and Th17 immunity [101]. molecules on antigen-presenting cells. Each T cell carries In recent researches there are more than two subunit antigen receptors of a different specificity and thereby the vaccines containing recombinant C. Albicans derived overall T cell population displays thereby a nearly proteins found to confer immunogenicity in phase I unlimited diversity of different antigenic specificities [91]. clinical trials as the most promising candidates for a The T cell repertoire is generated through somatic human vaccine [102]. Finally, due to the similarity between recombination of germline-encoded gene segments. fungal and mammalian cells significantly complicating Antigen recognition by CD4+ T cells is limited to peptide drug development and therapeutic approaches to combat antigens that are presented in the context of MHC fungal disease. However, advances in our understanding class II molecules [91]. To become fully functional, naïve of the interplay between fungi and the host have led to T cells require stimulation by cognate antigen-MHC-II the exploration and design of innovative complexes that are presented by antigen-presenting immunotherapeutic approaches. Initial therapeutic cells to induce their activation and clonal expansion, strategies were only focused on the use of recombinant which precedes their differentiation into effector T cells cytokines and Monoclonal antibodies have been recently [86, 90]. Only a few fungal T cell epitopes were evolved [97]. identified to date. These include the C. albicans-derived pALS3236-253 and pADH126-140 epitopes, which are **CONCLUSION** functionally conserved in diverse non-albicans species of Candida [91-95]. Different aspects of the innate and adaptive immune

in clinical practice due to several challenges. Among proven to be a promising area of scientific research. several challenges in the development of working fungal The pool of immunocompromised individuals is rapidly vaccines, most of immune-compromised individuals expanding, indicating that there would be an urgent (vulnerable groups) are unable to mount effective and need to develop novel and more potent antifungal drugs. strong immune response [96]. Additionally, the complexity This option would be especially relevant for of the eukaryotic fungal cells with a double layer of immunocompromised hosts, although the safety and protection with inner layer of plasma membrane and outer efficacy of novel antifungal remain questionable. A better layer of cell wall similar as mammalian cells caused major understanding of the role of the host-pathogen setback in the development of any vaccine or therapeutic interaction will lead to further development of potential drugs [97]. However, there are some exciting studies in novel antifungal therapies. recent years using fungal components of the cell wall and plasma membrane make theoretically possible to develop **REFERENCES** a universal vaccine [98]. For instance, a glycoconjugate vaccine composed of β -glucan, laminarin and the 1. Simon Altmeier and Salomé Leibund Gut-Landmann, diphtheria toxoid can mount a strong immune response 2017. Immunity to Fungal Infections. Section of and confer protection against infection with Candida or Immunology, Vetsuisse Faculty,University of Zurich, Aspergillus in mice [99]. Preclinical studies of vaccines Winterthurerstrasse 266a, 8057 Zurich, Switzerland. containing attenuated fungi have shown promising DOI 10.1007/978-3-319-50842-9. results. Forinstance, in2006, Pep1 was shown to be a cell 2. Brown, G.D., 2006. Dectin-1: a signaling non-TLR wall dominant antigen that was protective in mice pattern-recognition receptor. Nat. Rev. Immunol., challenged with C. posadasii by applying 6: 33-43. immunoproteomic and bioinformatic tools, researchers 3. Romani, L., 2011. Immunity to fungal infections. Nat. then identified five peptides from Pep1 that were predicted Rev. Immunol., 11: 275-288. to have high afnity to MHC-II and these peptides were 4. Fabián, S., D. Gordon and Brown, 2018. Antifungal able to induce IFN- γ by peptide-exposed lymphocytes Innate Immunity: A Perspective from the Last, [100]. In addition, subcutaneous vaccination with glucan 10 Years.

Antigenic Specificity of Antifungal CD4+ T Cells: T particles containing Cryptococcus alkaline extracts

Immunization and TreatmentAgainst Fungal Infections: Furthermore, novel analytic techniques capable of Currently, there are no proven fungal vaccines available detecting immune responses elicited by fungi have response to fungi have been intensely investigated.

-
-
-
-
-
- 2006. Invasive fungal infections: a review of Microbe., 17: 452-65. epidemiology and management options. J. Med. 19. NCT01926028 andNCT01067131.https://
- Immunology and Immunogenetics Insights, 8: 3-6 20. Nanjappa, S.G. and B.S. Klein, 2014. Vaccine
- 8. Butler, W.H., 1964. Fungi in human and animal Immunol., 28: 27-33. disease. Proc. R Soc. Med., 57: 416. 21. Lin, L., A.S. Ibrahim, X. Xu, J.M. Farber,
-
- reassessment. Med. Mycol., 46: 515-529. infection in mice. PLoS Pathog., 5: e1000703.
- 11. Chai, L.Y., M.G. Netea and A.G. Vonk, 2009. Kullberg 22. Brown, G.D., 2011. Innate antifungal immunity: immune response. Med. Mycol., 47: 227-236. 29: 1-21.
-
- a pivotal principle in host response to viral 4: 165rv113. infections. Clin Immunol., 143: 99-115. 24. Netea, M.G., G.D. Brown, B.J. Kullberg and N.A. Gow,
- N. Kutukculer and N. Karaca, 2015. DOCK8 Nat. Rev. Microbiol., 6: 67-78. Immunol., 35: 189-98. Microbiol., 14: 163-7.
- Gut- Landmann, 2012. A novel Th cell epitope of system. PLoS Pathog., 2: e35. Candida albicans mediates protection from fungal 27. Rappleye, C.A., L.G. Eissenberg and W.E. Goldman,
- adherent human neutrophils: triggering by receptor. P Natl Acad. Sci. USA, 104: 1366-70. colony- stimulating factors CSF-GM and CSF-G. 28. Kozel, T.R. and R.P. Mastroianni, 1976. Inhibition of
- S. Becattini, T. Rulicke, F. Sallusto and S. Leibund of phagocytosis. Infect. Immun., 14: 62-7. Gut-Landmann, 2015. Antigen-specific Th17 cells are 29. Gantner, B.N., R.M. Simmons and D.M. Underhill, 11: e1005164. 24: 1277-86.
- 5. Mihretu, A., 2019. Review on Immunity to Fungal 18. Wuthrich, M., T.T. Brandhorst, T.D. Sullivan, Infection. International Journal of Microbiological H. Filutowicz, A. Sterkel, D. Stewart, M. Li, Research, 10(3): 134-138, 2019 ISSN 2079-2093 © T. Lerksuthirat, V. LeBert and Z.T. Shen, 2015. IDOSI Publications, 2019 DOI: 10.5829/ Calnexin induces expansion of antigen-specific idosi.ijmr.2019.134.138. CD4+ T cells that confer immunity to fungal 6. Enoch, D.A., H.A. Ludlam and N.M. Brown, ascomycetes via conserved epitopes. Cell Host
- Microbiol., 55: 809-818. clinicaltrials.gov/ ct2/ show/NCT01926028and 7. Jiang, 2016. Immunity against Fungal Infections. https://clinicaltrials.gov/ct2/show/NCT01067131.
	- doi:10.4137/III.s38707. immunity against fungal infections. Curr Opin
- 9. Blanco, J.L., 2008. Garcia ME. Immune response to V. Avanesian, B. Baquir, Y. Fu, S.W. French, Jr. fungal infections. Vet. Immunol. Immunopathol., J.E. Edwards and B. Spellberg, 2009. Th1-Th17 cells 125: 47-70. mediate protective adaptive immunity against 10. Romani, L., 2008. Cell mediated immunity to fungi: a Staphylococcus aureus and Candida albicans
	- BJ. Fungal strategies for overcoming host innate the key role of phagocytes. Annu. Rev. Immunol.,
- 12. Tak, W., Mak and E. Mary, 2008. Saunders the 23. Brown, G.D., D.W. Denning, N.A. Gow, S.M. Levitz, Immune Response Basic and Clinical Principles. M.G. Netea and T.C. White, 2o12. Hidden killers: 13. Akram, A. and R.D. Inman, 2012. Immunodominance: human fungal infections. Sci. Transl. Med.,
- 14. Aydin, S.E., S.S. Kilic, C. Aytekin, A. Kumar, 2008. An integrated model of the recognition of O. Porras, L. Kainulainen, L. Kostyuchenko, F. Genel, Candida albicans by the innate immune system.
	- deficiency: clinical and immunological phenotype and 25. Erwig, L.P. and N.A.R. Gow, 2016. Interactions of treatment options - a review of 136 patients. J. Clin fungal pathogens with phagocytes. Nat. Rev.
- 15. Bar, E., A. Gladiator, S. Bastidas, B. Roschitzki, 26. Wheeler, R.T. and G.R. Fink, 2006. A drug-sensitive H. Acha-Orbea, A. Oxenius and S. Leibund genetic network masks fungi from the immune
- infection. J. Immunol., 188: 5636-43. 2017. Histoplasma capsulatum alpha-(1, 3)-glucan 16. Nathan, C.F., 1989. Respiratory burst in blocks innate immune recognition by the beta-glucan
- Blood, 73: 301-6. **phagocytosis** by cryptococcal polysaccharide: 17. Trautwein-Weidner, K., A. Gladiator, F.R. Kirchner, dissociation of the attachment and ingestion phases
	- primed by distinct and complementary dendritic cell 2005. Dectin-1 mediates macrophage recognition of subsets in oropharyngeal candidiasis. PLoS Pathog. Candida albicans yeast but not filaments. EMBO J.,
- cellular states. J. Immunol., 176: 3717-24. and Syk. Eur. J. Immunol., 44: 3729-40.
- 31. Dambuza, I.M. and G.D. Brown, 2015. C-type lectins 41. Sun, W.K., X. Lu, X. Li, Q.Y. Sun, X. Su, Y. Song,
- Immunol., 13: 817-22. 42. Taylor, P.R., S.V. Tsoni, J.A. Willment,
-
- O. Schulz, E. Schweighoffer, D.L. Williams, S. Gordon, 43. Hohl, T.M., H.L. Van Epps, A. Rivera, L.A. Morgan, type lectins. Immunity, 22: 507-17. PLoS Pathog., 1: e30.
- 35. Strasser, D., K. Neumann, H. Bergmann, 44. Steele, C., R.R. Rapaka, A. Metz, S.M. Pop, protein kinase C-sigma to elicit Card9 adaptor- PLoS Pathog., 1: e42. mediated innate immunity. Immunity, 36: 32-42. 45. Werner, J.L., A.E. Metz, D. Horn, T.R. Schoeb,
- 442: 651-6. 182: 4938-46.
- 37. Hara, H., C. Ishihara, A. Takeuchi, T. Imanishi, 46. Saijo, S., N. Fujikado, T. Furuta, S.H. Chung, ITAM-associated and Toll-like receptors. Nat. albicans. Nat. Immunol., 8: 39-46. Immunol., 8: 619-29. 47. Cunha, C., M. Di Ianni, S. Bozza, G. Giovannini,
-
- 39. Slack, E.C., M.J. Robinson, P. Hernanz-Falcon, antifungal immunity. Blood, 116: 5394-402. G.D. Brown, D.L. Williams, E. Schweighoffer, 48. Sainz, J., C.B. Lupianez, J. Segura-Catena, L. Vazquez, Eur. J. Immunol., 37: 1600-12. Aspergillosis Infection. Plos One, 7(2): e32273.
- 30. Gersuk, G.M., D.M. Underhill, L. Zhu and K.A. Marr, 40. Cohen-Kedar, S., L. Baram, H. Elad, E. Brazowski, 2006. Dectin-1 and TLRs permit macrophages to H. Guzner-Gur and I. Dotan, 2014. Human intestinal distinguish between different Aspergillus fumigatus epithelial cells respond to beta-glucans via Dectin-1
- in immunity: recent developments. Curr. Opin. H.M. Sun and Y. Shi, 2012. Dectin-1 is inducible and Immunol., 32: 21-7. plays a crucial role in Aspergillus-induced innate 32. Hardison, S.E. and G.D. Brown, 2012. C-type lectin immune responses in human bronchial epithelial cells. receptors orchestrate antifungal immunity. Nat. Eur. J. Clin. Microbiol. Infect. Dis., 31: 2755-64.
- 33. Osorio, F. and C. Reis e Sousa, 2011. Myeloid C-type K.M. Dennehy, M. Rosas H. Findon, K. Haynes, lectin receptors in pathogen recognition and host C. Steele, M. Botto, S. Gordon and G.D. Brown, 2007. defense. Immunity, 34: 651-64. Dectin-1 is required for beta-glucan recognition and 34. Rogers, N.C., E.C. Slack, A.D. Edwards, M.A. Nolte, control of fungal infection. Nat. Immunol., 8: 31-8.
	- V.L. Tybulewicz, G.D. Brown and C. Reis e Sousa, P.L. Chen, M. Feldmesser and E.G. Pamer, 2005. 2005. Syk-dependent cytokine induction by Dectin-1 Aspergillus fumigatus triggers inflammatory reveals a novel pattern recognition pathway for C responses by stage-specific beta-glucan display.
	- M.J. Marakalala, R. Guler, A. Rojowska, K.P. Hopfner, D.L. Williams, S. Gordon, J.K. Kolls and G.D. Brown, F. Brombacher, H. Urlaub and G. Baier, 2012. 2005. The beta-glucan receptor dectin-1 recognizes Syk kinase-coupled C-type lectin receptors engage specific morphologies of Aspergillus fumigatus.
- 36. Gross, O., A. Gewies, K. Finger, M. Schafer, M.M. Hewitt, L.M. Schwiebert, I. Faro-Trindade, T. Sparwasser, C. Peschel, I. Forster and J. Ruland, G.D. Brown and C. Steele, 2009. Requisite role for 2006. Card 9 controls a non-TLR signaling the dectin-1 beta-glucan receptor in pulmonary pathway for innate anti-fungal immunity. Nature., defense against Aspergillus fumigatus. J. Immunol.,
	- L. Xue, S.W. Morris, M. Inui, T. Takai, A. Shibuya H. Kotaki, K. Seki, K. Sudo, S. Akira, Y. Adachi and and S. Saijo, 2007. The adaptor protein CARD9 is N. Ohno, 2007. Dectin-1 is required for host defense essential for the activation of myeloid cells through against Pneumocystis carinii but not against Candida
- 38. Leibund Gut-Landmann, S., O. Gross, M.J. Robinson, S. Zagarella, T. Zelante, C. D'Angelo, A. Pierini, F. Osorio, E.C. Slack, S.V. Tsoni, E. Schweighoffer, L. Pitzurra and F. Falzetti, 2010. Dectin-1 Y238X V. Tybulewicz, G.D. Brown, J. Ruland and C. Reis e polymorphism associates with susceptibility to Sousa, 2007. Syk- and CARD9- dependent coupling invasive aspergillosis in hematopoietic of innate immunity to the induction of T helper cells transplantation through impairment of both that produce interleukin 17. Nat. Immunol., 8: 630-638. recipient- and donor-dependent mechanisms of
	- V.L. Tybulewicz and C. Reis e Sousa, 2007. R. Rios, S. Oyonarte, K. Hemminki, A. Forsti and Syk-dependent ERK activation regulates IL-2 and M. Jurado, 2012. Dectin-1 and DC-SIGN IL-10 production by DC stimulated with zymosan. Polymorphisms Associated with Invasive Pulmonary
- for high mannose. Glycobiology, 16: 422-30. receptors. J. Infect Dis., 188: 165-72.
- 50. Sato, K., X.L. Yang, T. Yudate, J.S. Chung, J. Wu, 61. Netea, M.G., C.A. Van Der Graaf, A.G. Vonk, Fc receptor gamma chain to induce innate immune J. Infect. Dis., 185: 1483-9. responses. J. Biol. Chem., 281: 38854-66. 62. Biondo, C., A. Malara, A. Costa, G. Signorino,
-
- 52. Zhao, X.Q., L.L. Zhu, Q. Chang, C. Jiang, Y. You, Eur. J. Immunol., 42: 2632-43. T. Luo, X.M. Jia and X. Lin, 2014. C-type lectin 63. Bochud, P.Y., J.W. Chien, K.A. Marr and
- 53. Zhu, L.L., X.Q. Zhao, C. Jiang, Y. You, X.P. Chen, J. Med., 359: 1766-77. Y.Y. Jiang, X.M. Jia and X. Lin, 2013. C-type lectin 64. Dinarello, C.A., 1998.Interleukin-1 beta, interleukin-18 pattern-recognition receptor for host defense against Y Acad Sci., 856: 1-11.
- Lett., 136: 1-12. **Example 2.1** for the NLRP3 inflammasome. MBio. 6.
- 55. Gringhuis, S.I., Den J. Dunnen, M. Litjens, B. Van Het 66. Cheng, S.C., F.L. Van De Veerdonk, M. Lenardon,
- Purification of the human alveolar macrophage Candida albicans. J. Leukoc Biol., 90: 357-66.
- a 175-kDa membrane protein. Proc. Natl. Acad. Sci. U kinase. PLoS One., 5:e10008. S A, 83: 2501-5. 68. Tomalka, J., S. Ganesan, E. Azodi, K. Patel,
-
- 59. Bianchi, M., A. Hakkim, V. Brinkmann, U. Siler, 7: e1002379. R.A. Seger, A. Zychlinsky and J. Reichenbach, 2009. 69. Ochs, H.D. and C.I. Smith, 1996. X-linked CGD controls aspergillosis. Blood, 114: 2619-22. analysis. Medicine, 75(6): 287-99.
- 49. McGreal, E.P., M. Rosas, G.D. Brown, S. Zamze, 60. Jouault, T., S. Ibata-Ombetta, O. Takeuchi, S.Y. Wong, S. Gordon, L. Martinez-Pomares and P.A. Trinel, P. Sacchetti, P. Lefebvre, S. Akira and P.R. Taylor, 2006. The carbohydrate-recognition D. Poulain, 2003. Candida albicans domain of Dectin-2 is a C-type lectin with specificity phospholipomannan is sensed through toll-like
	- K. Luby-Phelps, R.P. Kimberly, D. Underhill, Jr. I. Verschueren, J.W. Van der Meer and B.J. Kullberg, P.D. Cruz and K. Ariizumi, 2006. Dectin-2 is a pattern 2002. The role of toll-like receptor (TLR) 2 and TLR4 recognition receptor for fungi that couples with the in the host defense against disseminated candidiasis.
- 51. Miyake, Y., O.H. Masatsugu and S. Yamasaki, 2015. F. Cardile, A. Midiri, R. Galbo, S. Papasergi, C-type lectin receptor MCL facilitates mincle M. Domina and M. Pugliese, 2012. Recognition of expression and signaling through complex formation. fungal RNA by TLR7 has a nonredundant role in J. Immunol., 194: 5366-74. host defense against experimental candidiasis.
	- receptor dectin-3 mediates trehalose 6, 6'-dimycolate W.M. Leisenring, A. Upton, M. Janer, (TDM)-induced Mincle expression through S.D. Rodrigues, S. Li, J.A. Hansen and L.P. Zhao, CARD9/Bcl10/MALT1-dependent nuclear factor 2008. Toll-like receptor 4 polymorphisms and (NF)-kappaB activation. J. Biol. Chem, 289: 30052-62. aspergillosis in stem-cell transplantation. N Engl.
	- receptors Dectin-3 and Dectin-2 form a heterodimeric and the interleukin-1 beta converting enzyme. Ann. N
- fungal infection. Immunity, 39: 324-34. 65. Bruno, V.M., A. C. Shetty, J. Yano, PL. Jr. Fidel, 54. Redelinghuys, P. and G.D. Brown, 2011. Inhibitory M.C. Noverr and B.M. Peters, 2015. Transcriptomic C-type lectin receptors in myeloid cells. Immunol. analysis of vulvovaginal candidiasis identifies a role
- Hof, Y. Van Kooyk and T.B. Geijtenbeek, 2007. C-type M. Stoffels, T. Plantinga, S. Smeekens, L. Rizzetto, lectin DC-SIGN modulates Toll-like receptor signaling L. Mukaremera, K. Preechasuth and D. Cavalieri, 2011. via Raf-1 kinase-dependent acetylation of The dectin-1/inflammasome pathway is responsible transcription factor NF-kappaB. Immunity, 26: 605-16. for the induction of protective T-helper 17 responses 56. Stephenson, J.D. and V.L. Shepherd, 1987. that discriminate between yeasts and hyphae of
- mannose receptor. Biochem Biophys Res. Commun., 67. Said-Sadier, N., E. Padilla, G. Langsley and 148: 883-9. D.M. Ojcius, 2010. Aspergillus fumigatus stimulates 57. Wileman, T.E., M.R. Lennartz and P.D. Stahl, 1986. the NLRP3 inflammasome through a pathway Identification of the macrophage mannose receptor as requiring ROS production and the Syk tyrosine
- 58. Jordens, R., A. Thompson, R. Amons and F. Koning, P. Majmudar, B.A. Hall, K.A. Fitzgerald and 1999. Human dendritic cells shed a functional, A.G. Hise, 2011. A novel role for the NLRC4 soluble form of the mannose receptor. Int. Immunol., inflammasome in mucosal defenses against the 11: 1775-80. fungal pathogen Candida albicans. PLoS Pathog,
	- Restoration of NET formation by gene therapy in agammaglobulinemia. A clinical and molecular
- patients with multiple myeloma in the era of high-dose therapy and novel agents. Clin Infect Dis: Off Publ Infect Dis Soc Am., 49(8): 1211-25. Doi: 10.1086/605664.
- 71. Henriet, S.S., P.E. Verweij and A. Warris, 2012. Aspergillus nidulans and chronic granulomatous disease: a unique host-pathogen interaction. J. Infect Dis., 206(7): 1128-37. doi:10.1093/infdis/ jis473.
- 72. Qu, Y., L. Franchi, G. Nunez and G.R. Dubyak, 2007. Nonclassical IL-1 beta secretion stimulated by P2X7 receptors is dependent on inflammasome activation and correlated with exosome release in murine macrophages. J. Immunol., 179: 1913-25.
- 73. Kuida, K., Lippke, J.A., G. Ku, M.W. Harding, D.J. Livingston, M.S. Su and R.A. Flavell, 1995. Altered cytokine export and apoptosis in mice deficient in interleukin-1 beta converting enzyme. Science, 267: 2000-3.
- 74. Heutinck, K.M., I.J. Ten Berge, C.E. Hack, J. Hamann and A.T. Rowshani, 2010. Serine proteases of the human immune system in health and disease. Mol. Immunol., 47: 1943-55.
- 75. Wallach, D., T.B. Kang, C.P. Dillon and D.R. Green, 2016. Programmed necrosis in inflammation: Toward identification of the effector molecules. Science, 352: aaf2154.
- 76. White, S.H., W.C. Wimley and M.E. Selsted, 1995. "Structure, function and membrane integration of defensins". Curr. Opin. Struct. Biol., 5(4): 521-7. doi:10.1016/0959-440X (95)80038-7. PMID 8528769.
- 77. Hellgren, O. and B.C. Sheldon, 2011. "Locus-specific protocol for nine different innate immune genes $(antimicrobial peptides: \beta-defensins) across passengerine$ bird species reveals within-species coding variation and a case of trans-species polymorphisms". Molecular Ecology Resources 11(4): 686-692. doi:10.1111/j.1755-0998.2011.02995.x.]
- 78. Van Dijk, A., E.J. Veldhuizen and H.P. Haagsman, 2008. "Avian defensins". Vet. Immunol. Immunopathol., 124(1-2): 1-18. doi:10.1016/j. vetimm.2007.12.006. PMID 18313763.].
- 79. Sugiarto, H. and P.L. Yu, 2004. "Avian antimicrobial peptides: the defense role of beta-defensins". Biochem. Biophys. Res. Commun., 323(3): 721-7. doi:10.1016/j.bbrc.2004.08.162. PMID 15381059.
- 80. Hoover, D.M., O. Chertov and J. Lubkowski, 2001. "The structure of human beta-defensin-1: new insights into structural properties of betadefensins". J. Biol. Chem., 276(42): 39021-6. doi:10.1074/jbc.M103830200. PMID 11486002.].
- 70. Nucci, M. and E. Anaissie, 2009. Infections in 81. Casadevall, A., W. Cleare, M. Feldmesser, A. Glatman-Freedman, D.L. Goldman, T.R. Kozel, N. Lendvai, J. Mukherjee, L.A. Pirofski and J. Rivera, 1998. Characterization of a murine monoclonal antibody to Cryptococcus neoformans polysaccharide that is a candidate for human therapeutic studies. Antimicrob Agents Chemother., 42: 1437-46.
	- 82. Casadevall, A., J. Mukherjee, S.J. Devi, R. Schneerson, J.B. Robbins and M.D. Scharff, 1992. Antibodies elicited by a Cryptococcus neoformanstetanus toxoid conjugate vaccine have the same specificity as those elicited in infection. J. Infect. Dis., 165: 1086-93.
	- 83. Feldmesser, M. and A. Casadevall, 1997. Effect of serum IgG1 to Cryptococcus neoformans glucuronoxylomannan on murine pulmonary infection. J. Immunol., 158: 790-9.
	- 84. Granja, L.F., L. Pinto, C.A. Almeida, D.S. Alviano, M.H. Da Silva, R. Ejzemberg and C.S. Alviano, 2010. Spores of mucor ramosissimus, mucor plumbeus and mucor circinelloides and their ability to activate human complement system *in vitro*. Med. Mycol., 48: 278-84.
	- 85. Kozel, T.R., 1996. Activation of the complement system by pathogenic fungi. Clin. Microbiol. Rev., 9: 34-46.
	- 86. Kozel, T.R., L.C. Weinhold and D.M. Lupan, 1996. Distinct characteristics of initiation of the classical and alternative complement pathways by Candida albicans. Infect. Immun., 64: 3360-8.
	- 87. Thong, Y.H. and A. Ferrante, 1978. Alternative pathway of complement activation by Candida albicans. Aust NZ J. Med., 8: 620-2.
	- 88. Hunniger, K., K. Bieber, R. Martin, T. Lehnert, M.T. Figge, J. Loffler, R.F. Guo, N.C. Riedemann and O. Kurzai, 2015. A second stimulus required for enhanced antifungal activity of human neutrophils in blood is provided by anaphylatoxin C5a. J. Immunol., 194: 1199-210.
	- 89. Radovanovic, I., A. Mullick and P. Gros, 2011. Genetic control of susceptibility to infection with Candida albicans in mice. PLoS One., 6: e18957.
	- 90. Mullick, A., M. Elias, S. Picard, L. Bourget, O. Jovcevski, S. Gauthier, A. Tuite, P. Harakidas, C. Bihun, B. Massie and P. Gros, 2004. Dysregulated inflammatory response to Candida albicans in a C5-deficient mouse strain. Infect Immun., 72: 5868-76.
	- 91. Mullick, A., Z. Leon, G. Min-Oo, J. Berghout, R. Lo, E. Daniels and P. Gros, 2006. Cardiac failure in C5-deficient A/J mice after Candida albicans infection. Infect Immun., 74: 4439-51.
- 92. Yeaman, M.R., S.G. Filler, S. Chaili K. Barr, H. Wang, 98. Levitz, S.M., H. Huang, G.R. Ostroff and C.A. Specht, C.S. Schmidt and Jr. J.E. Edwards, 2014. Mechanisms vaccines. Semin Immunopathol., 37: 199-207. of NDV-3 vaccine efficacy in MRSA skin versus 99. Torosantucci, A., C. Bromuro, P. Chiani, F. De
- Jr. J.P. Hennessey, 2012. NDV-3, a recombinant 100. Tarcha, E.J., V. Basrur, C.Y. Hung, M.J. Gardner and
- 94. Drummond, R.A., C. Wallace, D.M. Reid, S.S. Way, 74: 516-527. D.H. Kaplan and G.D. Brown, 2012. Cutting edge: 101. Specht, C.A., C.K. Lee, H. Huang, D.J. Tipper,
- I. Leiner, D.B. Sant'Angelo and E.G. Pamer, 2006. MBio, 6: e01905-01915. Innate immune activation and CD4+ T cell priming 102. De Bernardis, F., M. Amacker, S. Arancia, S. Sandini,
- T. Warner and M. Evans, 2011. Vaccine-induced 30: 4490-4498. protection against 3 systemic mycoses endemic to North America requires Th17 cells in mice. J. Clin. Invest., 121: 554-68.
- 97. Armstrong-James, D., G.D. Brown, M.G. Netea, T. Zelante, M.S. Gresnigt, F.L. Van De Veerdonk and S.M. Levitz, 2017. Immunotherapeutic approaches to treatment of fungal diseases. Lancet Infect Dis., 17: e393-e402.
- D. Kupferwasser, Jr. J.P. Hennessey, Y. Fu, 2015. Exploiting fungal cell wall components in
- invasive infection. Proc. Natl. Acad. Sci. USA., Bernardis, F. Berti, C. Galli, F. Norelli, C. Bellucci, 111: E5555-63. L. Polonelli, P. Costantino, R. Rappuoli and 93. Schmidt, C.S., C.J. White, A.S. Ibrahim, S.G. Filler, A. Cassone, 2005. A novel glyco-conjugate vaccine Y. Fu, M.R. Yeaman, Jr. J.E. Edwards and against fungal pathogens. J. Exp. Med., 202: 597-606.
	- alum-adjuvanted vaccine for Candida and G.T. Cole, 2006. A recombinant aspartylprotease of Staphylococcus aureus, is safe and immunogenic in Coccidioidesposadasiiinducesprotectionagainstpu healthy adults. Vaccine, 30: 7594-600. lmonarycoccidioidomycosisinmice. Infect. Immun.,
- failure of antigen-specific CD4+ T cell recruitment Z.T. Shen, J.K. Lodge, J. Leszyk, G.R. Ostroff and to the kidney during systemic candidiasis. J. S.M. Levitz, 2015. Protection against experimental Immunol., 193: 5381-5. cryptococcosis following vaccination with glucan 95. Rivera, A., G. Ro, H.L. Van Epps, T. Simpson, particles containing cryptococcus alkaline extracts.
- during respiratory fungal infection. Immunity, C. Gremion, R. Zurbriggen, C. Moser and 25: 665-75. A. Cassone, 2012. A virosomal vaccine against 96. Wuthrich, M., B. Gern, C.Y. Hung, K. Ersland, candidal vaginitis: immunogenicity, efficacy N. Rocco, J. Pick-Jacobs, K. Galles, H. Filutowicz, and safety profile in animal models. Vaccine,