

Manglicolous Marine Fungi of Kerala (South India)

R. Gayatri Nambiar and K. Raveendran

Department of P.G. Studies and Research in Botany,
Sir Syed College, Taliparamba, Kannur, Kerala-670142, India

Abstract: Twenty six manglicolous marine fungi comprising 20 Ascomycetes, 1 Basidiomycete and 5 Mitosporic fungi were isolated from the mangrove forests of Kerala, South India. Average isolates per wood sample and percentage colonization were 1.54 and 81.25 respectively. Based on the percent frequency of occurrence, *Lulworthia grandispora* (13.19%), *Dactylospora haliotrepha* (12.09%), *Savoryella lignicola* (10.99%) and *Cirrenalia pygmaea* (10.99 %) were the most frequent species.

Key words: Mangrove forest • Detritus • Decomposition • Marine fungi

INTRODUCTION

Tropical coastlines are covered by large expanses of mangrove forests which play significant roles in supporting planktonic and benthic communities and nutrient cycles, much of it through decomposition of detritus [1, 2]. Dead leaves, woody debris, animal remains and other materials constitute the main sources of organic matter, mostly produced *insitu* and some imported through inflowing freshwater streams. In terms of above-ground production and biomass, wood dominates detritus supply in mangroves [3]. Although wood tends to decompose slowly, it is an important component of food webs and nutrient cycles of terrestrial and fresh water ecosystem [4]. Degradation of woody substrates in aquatic ecosystem includes physical fragmentation and biological decomposition by bacteria and fungi [5, 6].

Although wood is reported to be high in mangrove ecosystem, little is known about its production [7, 8]. The exact proportion of wood maintained as biomass and the proportion of wood that enter the detritus path way in each year is unknown [9]. Measurements of standing timber stock on the forest floor are also not known [10]. This dead wood is likely to serve as an important energy source for primary consumers. Therefore detailed studies of the organisms involved in wood decay are necessary to gain a better understanding pathway of energy flow in mangrove ecosystem [11].

Mangrove trees are able to grow at salinities ranging from full sea water to fresh water, thus a different fungal flora can be expected with in this salinity gradient [12]. 54

mangrove trees with 60 associates were listed by Tomlinson [13]. 200 higher marine fungi were encountered from fifty five mangroves and their associates [14]. A total of 165 marine fungi encompassing 111 ascomycetes, 1 basidiomycete and 53 mitosporic fungi were reported from Indian mangroves [15]. Chinnaraj [16], Ravikumar and Vittal [17], Borse *et al* [18], Sarma and Vittal [19, 20], Sarma *et al*, [21], Maria and Sridhar [22- 24], Pawar and Borse [25], Raveendran and Manimohan [26], Gayatri and Raveendran [27- 29], Sridhar [30] etc have published reports on manglicolous marine fungi from Indian Peninsula. During marine mycological survey of Kerala coast (Arabian Sea), data on the frequency of occurrence of marine fungi were recorded and the results were presented in this paper.

MATERIALS AND METHODS

Collection and Treatment of Wood Samples: Wood samples were collected from the mangrove forests of Kavvai-Kunhimangalam backwater, the longest backwater stretch of northern Kerala, falls with in the geographic coordinates 11°59' 52"- 12°4' 36"N latitude and 75°06' 48" - 75°15' 40"E longitude. Sample collection was carried out during May 2007 -June 2008. Collected samples include decaying intertidal woods, drift woods and mangrove woods. They were washed well, placed in sterile polythene bags and were brought to the laboratory. After the preliminary screening for marine fungi under stereomicroscope, the wood samples were incubated at room temperature. Periodical isolation of marine fungi from

Table 1: List of manglicolous marine fungi from Kerala

Name of fungi	No: of isolates	Frequency of occurrence
Ascomycetes		
<i>Aigialus grandis</i> Kohlm. et Schatz	3	3.30
<i>Aniptodera chesapeakeensis</i> Shearer et Miller	4	4.39
<i>Aniptodera haispora</i> Vrijmoed, Hyde et Jones	1	1.10
<i>Aniptodera mangrovei</i> Hyde, Farrant et Jones	6	6.59
<i>Aniptodera salsuginosa</i> Nakagiri et Ito	3	3.30
<i>Ascocratera manglicola</i> Kohlm.	1	1.10
<i>Biatriospora marina</i> Hyde et Borse	2	2.20
<i>Dactylospora haliotrepha</i> Kohlm. et Kohlm) Hafellner	11	12.09
<i>Halorosellinia oceanica</i> (Schatz) Whalley, Jones, Hyde et Laessoe	6	6.59
<i>Halosarpheia marina</i> (Cribb et Cribb) Kohlm	5	5.49
<i>Halosarpheia minuta</i> Leong	2	2.20
<i>Leptosphaeria australiensis</i> (Cribb et Cribb) Hughes	8	8.79
<i>Lignincola laevis</i> Hohnk.	9	9.89
<i>Lignincola tropica</i> Kohlm.	6	6.59
<i>Lulworthia grandispora</i> Meyers	12	13.19
<i>Marinosphaera mangrovei</i> Hyde	5	5.49
<i>Panorbis viscosus</i> (Schmidt) Campb. anderson et Shearer	4	4.39
<i>Savoryella paucispora</i> (Cribb et Cribb) Koch	6	6.59
<i>Savoryella lignicola</i> Jones et Eaton	10	10.99
<i>Verruculina enalia</i> (Kohlm) Kohlm et Kohlm	7	7.69
Basidiomycete		
<i>Halocyphina villosa</i> Kohlm et Kohlm	5	5.49
Mitosporic fungi		
<i>Cirrenalia pygmaea</i> Kohlm.	10	10.99
<i>Periconia prolifica</i> Anastasiou	6	6.59
<i>Phoma</i> sp	2	2.20
<i>Trichocladium alopallonellum</i> (Meyers et Moore) Kohlm et V.kohlm	4	4.39
<i>Zalerion varium</i> Anastasiou	2	2.20

these wood samples were carried out for six months. Identifications of marine fungi were done using taxonomic keys [26, 31-33]. The marine fungi thus identified were tabulated and recorded (Table 1).

Presentation of Data

- Percent frequency of occurrence (FO) = Number of isolates of a particular species divided by total number of wood samples supporting marine fungi X 100.

On the basis of percentage occurrence, the marine fungi were classified as very frequent (occurring in > 10 % samples), frequent (in 5- 10 %), infrequent (in 3- 5 % samples) and rare (in < 3 %).

- Mean number of fungi per sample = Total number of fungal isolates divided by total number of wood samples supporting marine fungi.
- Percentage colonization = Total number of wood samples supporting marine fungi divided by total number of wood samples examined x 100.

RESULTS AND DISCUSSION

Altogether 26 manglicolous marine fungi comprising 20 Ascomycetes, 1 Basidiomycete and 5 Mitosporic fungi were encountered. Average isolates per wood sample and percentage colonization were 1.54 and 81.25 respectively.

Based on percent frequency of occurrence, *Lulworthia grandispora* (13.19%), *Dactyl ospora*

haliotrepha (12.09%), *Savoryella lignicola* (10.99%) and *Cirrenalia pygmaea* (10.99 %) were the most frequent species. Among the eleven frequent species isolated, *Lignincola laevis* (9.89%) showed maximum value. Five species, namely *Aniptodera chesapeakensis*, *Panorbis viscosus*, *Trichocladium alopallonellum*, *Aigialus grandis* and *Aniptodera salsuginosa* were occasionally isolated. While *Biatrispora marina*, *Halosarpheia minuta*, *Phoma* sp, *Zalerion varium*, *Ascocratera manglicola* and *Aniptodera haispora* were found sporadic.

Among the fungal species in a community, core-group fungi (frequency > 10%) exert major influence on turnover of litter in mangrove ecosystem [34]. Only 4 core group fungi were obtained in the present study. Ravikumar and Vittal [17] also obtained 4 core group fungi from Pichavaram mangrove forest of south east India. However, Maria and Sridhar [22] encountered 13 core-group fungi from west coast of India. The percentage colonization in the present study is higher than those seen in the Philippines (80%) [10] and Malaysia (80.4%) [35] but lower than Singapore (85%) [36]. The mean number per samples obtained in this study was higher than Mauritius (1.1) [37] and Seychelles (1.1) [38] but similar to Malaysia (1.5) [36]. Borse *et al.*, [18] reported *Julella avicenniae* and *Aigialus parvus* as the most frequent fungi in Gujarat, *Verruculina enalia* from east coast of India [39], *Leptosphaeria australiensis* and *Halocyphina villosa* in Malaysia [40]. However, *Dactylospora haliotrepha* was recorded most frequently from North Sumatra [41], *Savoryella lignicola* in Thailand [42] and *Lulworthia grandispora* in Seychelles [43].

The study supports that the occurrence of manglicolous marine fungi of Kerala is similar to other parts of Indian Ocean. However, the species composition is relatively poor. This could be attributed to limited number of samples examined. In addition, the availability of substrata, host specificity, tissue or organ preference, temperature, salinity, succession and seasonality may also influence marine fungal communities [44].

ACKNOWLEDGEMENT

The authors are thankful to the Principal and Management of Sir Syed College, Taliparamba for providing facilities.

REFERENCES

1. Robertson, A.I., D.M. Alongi and K.G. Boto, 1992. Food chains and carbon fluxes. In: *Tropical mangrove ecosystems* (Eds. A.I. Robertson and D.M. Alongi) American Geophysical Union, Washington, pp: 293-329.
2. Alongi, D.M., F. Tirendi, P. Dixon, A. Trott, Lo and G.J. Brunskill, 1999. Mineralization of organic matter in intertidal sediments of a tropical semi-enclosed delta. *Estuar. Coast. Shelf Sci.*, 48: 451-467.
3. Gong, W.K., 1984. Mangrove primary productivity. In: *Productivity of the mangrove ecosystem: management and implications* (Eds. J.E. Ong and W.K. Gong) University of Sains, Penang, Malaysia, pp: 10-19.
4. Maria, G.L., K.R. Sridhar and F. Barlocher, 2006. Decomposition of dead twigs of *Avicennia officinalis* and *Rhizophora mucronata* in a mangrove in south western India. *Bot. Mari.*, 49: 450-455.
5. Barlocher, F., 1992. *The Ecology of Aquatic Hyphomycetes*. Springer Verlag, Berlin.
6. Wong, M.K.M., T.K. Goh, I.J. Hodgkiss, K.D. Hyde, V.M. Ranghoo, C.K.M. Tsui, W.H. Ho, W.S.W. Wong, and T.K. Yuen, 1948. Role of fungi in fresh water ecosystems. *Biodiversity and Conservation*, 7: 1187-1206.
7. Bunt, J.S., K.G. Boto and G. Boto, 1979. A survey method for estimating potential levels of mangrove forest primary production. *Marine Biology.*, 52: 123-128.
8. Boto, K.G., J.S. Bunt and J.T. Wellington, 1984. Variations in mangrove forest productivity in Northern Australia and Papua New Guinea. *Estuar. Coast Shelf Sci.*, 19: 321-329.
9. Robertson, A.I., 1987. The determination of tropic relationships in mangrove- dominated systems: Areas of darkness In: *Mangrove ecosystems of Asia and the Pacific: Status, Exploitation and management* (Eds. C.D. Field and A.J. Dartnall). Australian Institute of Marine Science, Australia.
10. Jones, E.B.G., F.R. Uyenco and M.P. Folloso, 1988. Fungi on drift wood collected in the intertidal zone from the Philippines. *Asian. Marine Biology*, 5: 103-106.
11. Hyde, K.D., 1990. A comparison of the intertidal mycota of five mangrove tree species. *Asian Marine Biology*, 7: 93-107.

12. Kohlmeyer, J., 1969. Ecological notes on fungi in mangrove forest. Trans Br. Mycol. Soc., 53: 237-250.
13. Tomlinson, P.B., 1986. The botany of mangroves, Cambridge Univ. Press Cambridge, pp: 413.
14. Jones, E.B.G. and S.A. Alias, 1997. Biodiversity of mangrove fungi. In: Biodiversity of Tropical Marine fungi (Eds. K.D. Hyde), Hong Kong University Press, Hong Kong, pp: 71-92.
15. Sridhar, K.R., 2009a. Mangrove fungi of the Indian Peninsula. In: Frontiers in Fungal Ecology, Diversity and Metabolites (Eds. K.R. Sridhar) IK International publishing House Pvt Ltd, New Delhi, pp: 28-50.
16. Chinnaraj, S., 1993. Higher marine fungi from mangroves of Andaman and Nicobar islands. Sydowia, 45: 109-115.
17. Ravikumar, D.R. and B.P.R. Vittal, 1996. Fungal diversity on decomposing mangrove plant *Rhizophora* in Pichavaram estuary, east coast of India. Indian. J. Mar. Sci., 25: 142-144.
18. Borse, B.D., D.J. Kelkar and A.C. Patil, 2000. Frequency of occurrence of marine fungi from Piroton Island (Gujarat), India. Geobios, 27: 145-148.
19. Sarma, V.V. and B.P.R. Vittal, 2000. Biodiversity of marine mangrove on different substrata of *Rhizophora apiculata* and *Avicennia sp* from Godavari and Krishna deltas, east coast of India. Fungal Diversity, 5: 23-41.
20. Sarma, V.V. and B.P.R. Vittal, 2001. Biodiversity of manglicolous fungi on selected plants in the Godavari and Krishna deltas, east coast of India. Fungal Diversity, 6: 115-130.
21. Sarma, V.V., K.D. Hyde and B.P.R. Vittal, 2000. Frequency of occurrence of mangrove fungi from east coast of India. Hydrobiologia, 455: 41-53.
22. Maria, G.L. and K.R. Sridhar, 2002. Richness and diversity of filamentous fungi on woody litter of mangroves along the west coast of India. Curr. Sci., 83: 1573-1580.
23. Maria, G.L. and K.R. Sridhar, 2003. Diversity of filamentous fungi on woody litter of five mangrove plant species from the southwest coast of India. Fungal Diversity, 14: 109-126.
24. Maria, G.L. and K.R. Sridhar, 2004. Fungal colonization of immersed wood in mangroves of the south west coast of India. Can. J. Bot. 82: 1409-1418.
25. Pawar, N.S. and B.D. Borse, 2004. Marine fungi from Sundarbans (India) - VI. J. Adv. Sci., and Tech, 7: 17-28.
26. Raveendran, K. and P. Manimohan, 2007. Marine fungi of Kerala-A preliminary floristic and ecological study Malabar Natural History Society, Calicut, Kerala, India.
27. Gayatri, R. Nambiar and K. Raveendran, 2008. Biodiversity of marine mangrove fungi of Valappattanam and Pichavaram mangrove forests (South India). Ecochronicle, 3: 137-140.
28. Gayatri, R. Nambiar and K. Raveendran, 2009a. Frequency and abundance of marine mycoflora in mangrove ecosystem of North Malabar, Kerala (India). Academic J Plant Sci., 2: 65-68.
29. Gayatri, R. Nambiar and K. Raveendran, 2009b. Manglicolous marine fungi on *Avicennia* and *Rhizophora* along Kerala coast (India). Middle -East J. Scientific Res., 4: 48-51.
30. Sridhar, K.R., 2009b. Fungal diversity of Pichavaram mangroves, Southeast coast of India. Nature and Sci., 7: 67-75.
31. Kohlmeyer, J. and Volkman Kohlmeyer, 1979. Marine Mycology - The Higher Fungi. Academic Press, New York.
32. Kohlmeyer, J. and E. Kohlmeyer, 1991. Illustrated key to the filamentous higher marine fungi. Bot. Mar., 34: 1-61.
33. Hyde, K.D. and V.V. Sarma, 2000. Pictorial keys to higher marine fungi. Marine Mycology -A Practical Approach (Eds. K.D. Hyde and S.B. Pointing) Fungal Diversity Press, Hong Kong, pp: 205-270.
34. Ananda, K. and K.R. Sridhar, 2004. Diversity of filamentous fungi on decomposing leaf and woody litter of mangrove forests in the southwest coast of India. Curr. Sci., 87: 1431-1437.
35. Tan, T.K. and W.F. Leong, 1992. Lignicolous fungi of tropical mangrove wood. Mycol. Res., 96: 413-414.
36. Tan, T.K., W.F. Leong, and E.B.G. Jones, 1989. Succession of fungi on wood of *Avicennia alba* and *Avicennia lanata* in Singapore. Can. J. Bot., 67: 2686-2691.
37. Vrijimoed, L.L., K.D. Hyde, and E.B.G. Jones, 1994. Observation on mangrove fungi from Macau and Hong Kong, with the description of two new ascomycetes *Diaporthe salsuginosa* and *Aniptodera haispora*. Mycol. Res., 98: 699-704.
38. Hyde, K.D. and E.B.G. Jones, 1989. Ecological observation of marine fungi from the Seychelles. Bot. J. Linn. Soc., 100: 237-254.

39. Ravikumar, D.R., 1991. Studies on fungi from mangroves of the east coast of India. PhD. Thesis. University of Madras, India.
40. Alias, S.A., A.S. Kuthubutheen, and E.B.G. Jones, 1995. Frequency of occurrence of fungi on wood in Malaysian mangroves. *Hydrobiologia*, 295: 97-106.
41. Hyde, K.D., 1988. Observations on the vertical distribution of marine fungi on *Rhizophora* sp. at Kampong Danau Mangrove, Brunei. *Asian Marine Biology*, 5: 77-81.
42. Hyde, K.D., A. Chalermpongse and T. Boonthavikoon, 1990. Ecology of intertidal fungi at Ranong mangrove, Thailand. *Trans. Mycol. Soc., Japn*, 31: 17-27.
43. Hyde, K.D. and E.B.G. Jones, 1988. Marine mangrove fungi. *Mar. Ecol.*, 9: 15-33.
44. Sarma, V.V. and K.D. Hyde, 2001. A review on frequently occurring fungi in mangroves. *Fungal Diversity*, 8: 1-34.