

## Qualitative Estimation of Cellulase and Lignin Modifying Enzymes in Five Wild Fungal Species Collected From Northern West India

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**Abstract:** In the present paper five wild *Pleurotus* species namely *P. floridanus* Singer, *P. pulmonarius* (Fr.) Quél., *P. sapidus* Quél., *P. cystidiosus* O.K.Mill. and *P. sajor-caju* (Fr.) Sing., collected from different localities of North West India have been studied for the qualitative estimation of cellulose and lignin degradation enzymes through colorimetric assays. On the basis of the results obtained, the production of these enzymes started from the fifth day of mycelial growth on petriplate. Whereas, the lignin modifying enzymes were detected on the third day of mycelial growth. The degradation of carboxymethyl cellulose (CMC) was observed with a yellow opaque layer formation around the colony in the concentric manner in case of cellulase enzymes detection. The enzymes responsible for the modification of lignin were observed with the formation of brown oxidation zones around the colonies. The findings clearly indicate that likewise other wood degrading fungi. These wild despite of occurring on different hosts contain the enzymes responsible for cellulose and lignin degradation.

**Key words:** *Pleurotus* species • Cellulase • Lignin modifying enzymes

### INTRODUCTION

Genus *Pleurotus* (Fr.) P. Kumm, belongs to the family Pleurotaceae of order Agaricales. All the species of this genus are wood inhabiting. Qualitative estimation of lignin and cellulose has been used for the study of systematics and biodiversity in fungi [1]. The study of the enzymes responsible for the degradation of cellulose and lignin is essential for understanding the ecological and also in the biotechnology potential of enzymes involved in this process [2]. Lignocellulose is a heteropolymer consisting mainly of three components, cellulose, hemicellulose and lignin [3-5]. Lignocellulose degrading enzymes plays a vital role in the basic research on classification of enzymes [6,7]. Qualitative assays are powerful tools used in screening fungi for lignocellulose degrading enzyme production [8-10]. The importance of the qualitative tests in the detection of these enzymes further provide the basis of other classification of enzyme occurring in fungi and they are also useful in screening large numbers of fungal isolates for several classes of enzyme, where definitive quantitative data are not required [4]. Therefore, the present study was mainly

aimed to investigate the five wild species of fungi collected from the different localities are studied for their capability to degrade two common polysaccharide viz., lignin and cellulose, which is quite useful as the basic need for enzyme profiling.

### MATERIALS AND METHODS

**Qualitative Detection of Cellulase Enzymes:** Activity of cellulase enzymes was detected by dye staining of carboxymethyl cellulose (CMC). The Composition of Cellulose basic medium (CBM) ( $\text{g L}^{-1}$ ) is given below:

|   |       |
|---|-------|
| $\text{C}_4\text{H}_{12}\text{N}_2\text{O}_6$ | 5     |
| $\text{KH}_2\text{PO}_4$                      | 1     |
| $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$     | 0.5   |
| Yeast Extract                                 | 0.1   |
| $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$     | 0.001 |

CBM medium was supplemented with 2% w/v low viscosity CMC and 1.6% w/v agar and then autoclaved. Subsequently it was inoculated with test fungus after pouring. The test fungus was incubated at 25°C in

darkness. When the colony diameter reached 30 mm, the agar plates were stained firstly by flooding with 2% w/v aqueous Congo red which was then left to stand undisturbed for fifteen minutes. The stain was poured off and then plates were flooded with 1M NaCl so as to destain for another fifteen minutes. The activity then was observed as yellow opaque area against a red color of undegraded CMC.

#### Qualitative Detection of Lignin Modifying Enzymes:

Activity of lignin modifying enzymes was detected by dye staining of lignin modifying basal medium (LME). The Composition of lignin basic medium (LBM) (g L<sup>-1</sup>) is given below:

|  |       |
|--|-------|
| KH <sub>2</sub> PO <sub>4</sub>                              | 1     |
| Yeast Extract  | 0.01  |
| C <sub>4</sub> H <sub>12</sub> N <sub>2</sub> O <sub>6</sub> | 0.5   |
| CuSO <sub>4</sub> .5H <sub>2</sub> O                         | 0.001 |
| MgSO <sub>4</sub> .7H <sub>2</sub> O                         | 0.5   |
| Fe(SO <sub>4</sub> ) <sub>3</sub>                            | 0.001 |
| CaCl <sub>2</sub> .2H <sub>2</sub> O                         | 0.01  |
| MnSO <sub>4</sub> .H <sub>2</sub> O                          | 0.001 |

LBM was supplemented with 1.6% w/v agar and autoclaved. To this added 1 ml of separately sterilized 20% aqueous glucose solution and 1 ml of 1% w/v aqueous tannic acid solution to each 100 ml growth medium prepared. Then this medium was transferred aseptically into the petriplates and inoculated with test fungus after solidification of medium. The petriplates were incubated at 25 °C in darkness and examined plates daily for ten days. LME production was recorded as the appearance of brown oxidation zones around the colony.

### RESULTS AND DISCUSSION

Five wild species of fungi were collected from different hosts and from different localities and altitude

(Table 1 and Figure 1). The species of genus *Pleurotus* are all wood inhabiting as these contain cellulase as well as other wood inhabiting mushrooms. The five wild *Pleurotus* species were checked for cellulase activity qualitatively by dye diffusion method. In this carboxymethyl cellulose (CMC) is used as a substrate which is also a substrate for endoglucanase and so can be used as a test for endoglucanase and glucosidase activity. This assay is a good indicator of cellulolytic ability since endoglucanase is generally produced in larger quantity by fungi than cellobiohydrolase [11-13]. In addition many fungi that successfully degrade cellulose on wood produce no detectable cellobiohydrolase [14]. In this assay after growth of the fungus on CMC, a dye is used to differentiate between intact CMC and degraded substrate. CMC degradation around the colonies appears as a yellow-opaque area against a red colour for undegraded CMC. All the five species of *Pleurotus* exhibited good amount of cellulase activity. The activity is checked on the earlier stages of culture growth, when it was just five days old. All the species showed considerable activity against the lignocelluloses substrates. The prominent zone of yellow opaque area in the cellulase activity was observed on the fifth day of the mycelial growth on petriplate. The zone was in concentric manner around the mycelial colony. The formation of such zone was constant from fifth day till the maturation of mycelium. This assay is a well established procedure and has been used in many areas like systematics and biodiversity [15-20]. Lignin modifying enzyme assay can be useful in determining the ability of a fungus to utilize a lignin substrate. The method indicates degradation of phenolic components in lignin. Degradation of the more recalcitrant non-phenolic lignin components. These procedures offer a powerful research tool for obtaining data on possible methods of lignocellulose substrate utilization, or the production of commercially important enzymes (Figure 2 and Figure 3).

Table 1: Fungal species with their host and altitude range

| Fungal Species               | Host                      | Location                     | Altitude(m) | Type of Forest |
|------------------------------|---------------------------|------------------------------|-------------|----------------|
| <i>Pleurotus floridanus</i>  | <i>Ficus benghalensis</i> | Patiala (Punjab)             | 250         | Plains         |
| <i>Pleurotus pulmonarius</i> | <i>Albizia chinensis</i>  | Palampur (Himachal Pradesh)  | 1200        | Mixed          |
| <i>Pleurotus sapidus</i>     | <i>Grevillea robusta</i>  | Palampur (Himachal Pradesh.) | 950         | Plains         |
| <i>Pleurotus cystidiosus</i> | <i>Mangifera indica</i>   | Patiala (Punjab)             | 250         | Plains         |
| <i>Pleurotus sajor- caju</i> | <i>Albizia chinensis</i>  | Palampur (Himachal Pradesh.) | 1200        | Plains         |

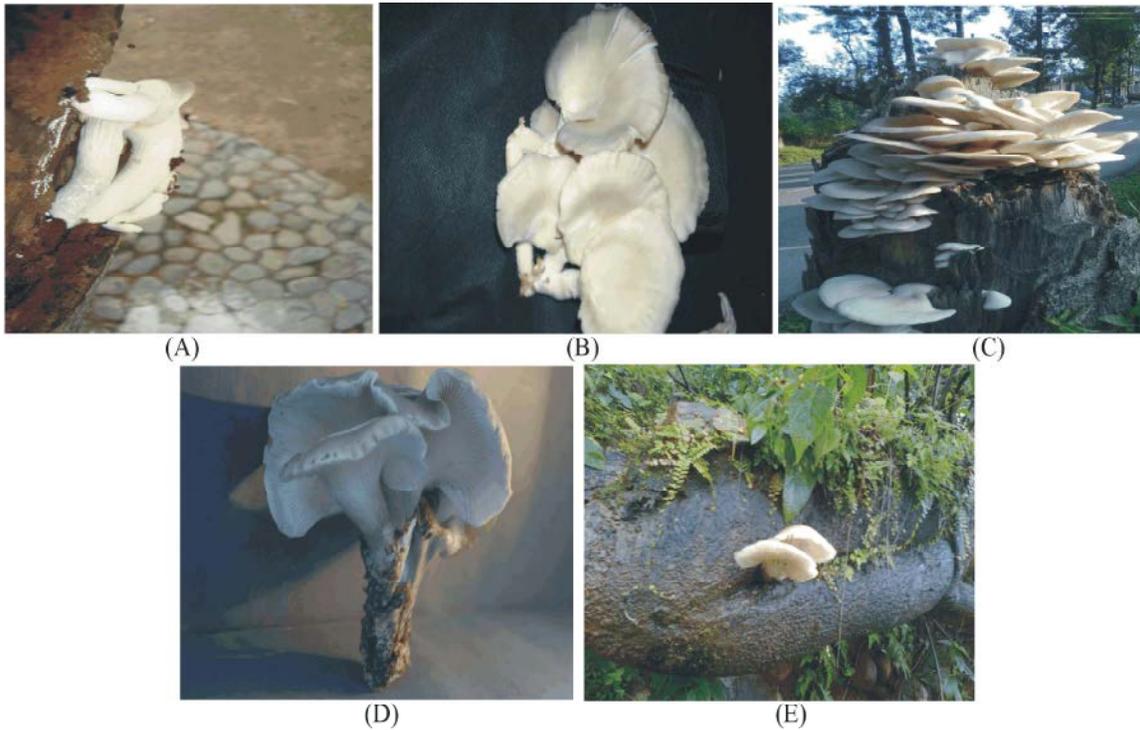


Fig. 1: A. *Pleurotus floridanus* B. *Pleurotus pulmonarius* C. *Pleurotus sapidus* D. *Pleurotus cystidiosus* and E. *Pleurotus sajor – caju*.

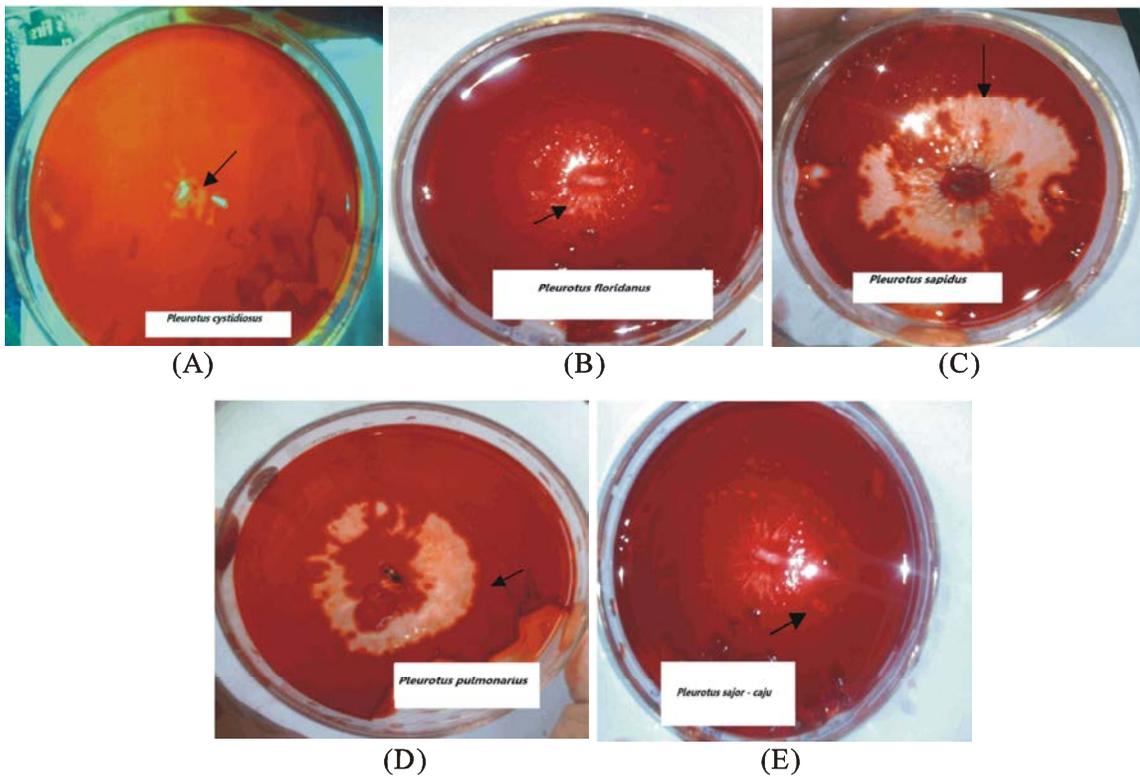


Fig. 2: A Cellulase activity of *Pleurotus cystidiosus* B. Cellulase activity of *Pleurotus floridanus* C Cellulase activity of *Pleurotus sapidus* D. Cellulase activity of *Pleurotus pulmonarius* E. Cellulase activity of *Pleurotus sajor – caju*.

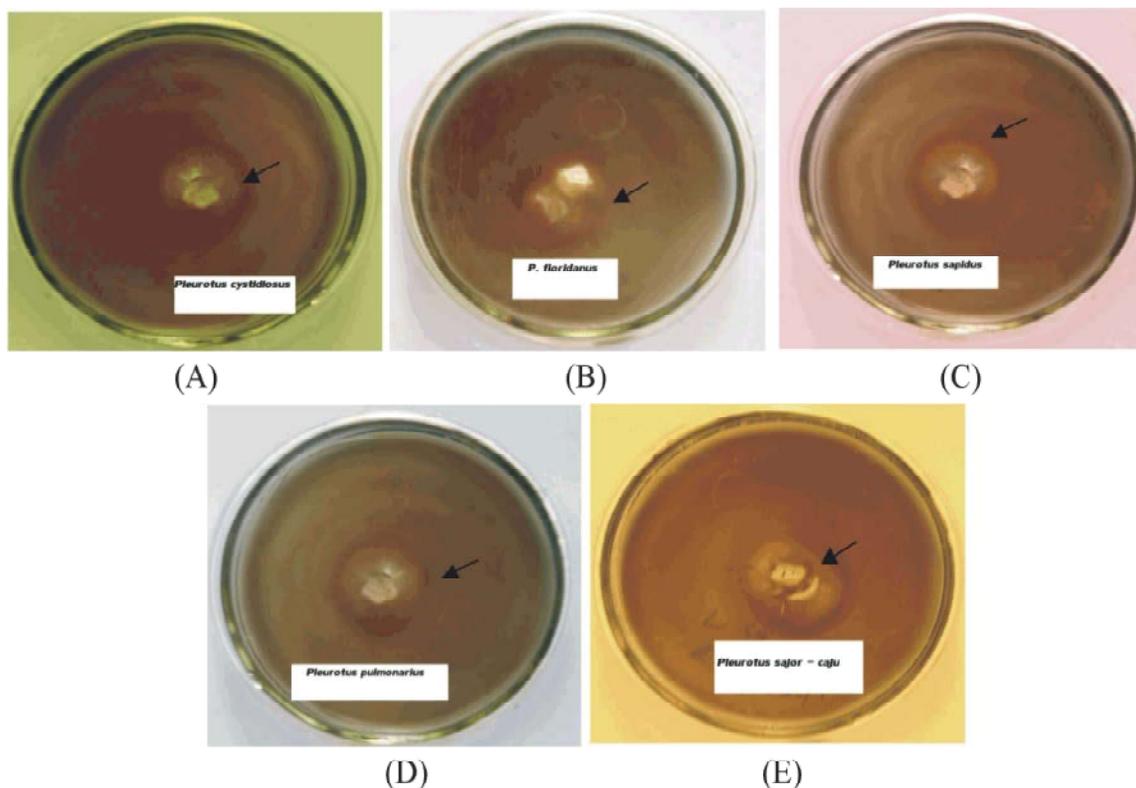


Fig. 3: A. Lignin modifying activity A. *Pleurotus cystidiosus* B. *Pleurotus floridanus* C *Pleurotus sapidus* D. *Pleurotus pulmonarius* E. *Pleurotus sajor – caju*.

### CONCLUSIONS

Qualitative estimation of the lignocellulosic enzymes provides the basic information for the presence of enzymes in the wood inhabiting mushrooms. This is quite useful in the fields of systematics and biodiversity. Further their role in the ecological aspects can be best understood with these basic tools.

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### REFERENCES

- Hyde, K.D., 1997. Biodiversity of Tropical Microfungi. Hong Kong University Press, Hong Kong.
- Reddy, C.A., 1995. The potential for white-rot fungi in the treatment of pollutants. Current Opinion in Biotechnology, 6: 320-328.
- Fengel, D. and G. Wegener, 1989. Wood. de Gruyter, New York, USA.
- Eaton, R.A. and M.D.C. Hale, 1993. Wood, Decay, Pests and Prevention. Chapman and Hall, London, UK.
- Egger, K.N., 1986. Substrate hydrolysis patterns of post-fire ascomycetes (Pezizales). Mycologia, 78: 771-780.
- Eggert, C., U. Temp, J.F.D. Dean and K.E.L. Eriksson, 1996. A fungal metabolite mediates degradation of non-phenolic lignin structures and synthetic lignins by laccase. FEBS Letters, 3(1): 144-148.
- Reddy, C.A. and T.M. D'Souza, 1994. Physiology and molecular biology of the lignin peroxidases of *Phanerochaete chrysosporium*. FEMS Microbiology Reviews, 13: 137-152.
- Thurston, C.F., 1994. The structure and function of fungal laccases. Microbiology, 140: 19-26.
- Reddy, C.A., 1995. The potential for white-rot fungi in the treatment of pollutants. Current Opinion in Biotechnology, 6: 320-328.
- Pointing, S.B., L.L.P. Vrijmoed and E.B.G. Jones, 1998. A qualitative assessment of lignocellulose degrading enzyme activity in marine fungi. Botanica Marina, 41: 293-298.

11. Cai, Y.J., I.A. Buswell and S.T. Chang, 1994. Cellulases and hemicellulases of *Volvariella volvacea* and the effect of tween 80 on enzyme production. *Mycological Research*, 98: 1019-1024.
12. Buswell, I.A., Y.J. Cai, S.T. Chang, I.F. Peberdy, S.Y., Fu and H.S. Yu, 1996. Lignocellulolytic enzyme profiles of edible mushroom fungi. *World Journal of Microbiology and Biotechnology*, 12: 537-542.
13. Rautella, G.S. and E.B. Cowling, 1966. Simple cultural test for relative cellulolytic activity of fungi. *Applied Microbiology*, 14: 892-898.
14. Cai, Y.J., I.A. Buswell and S.T. Chang, 1994. Cellulases and hemicellulases of *Volvariella volvacea* and the effect of tween 80 on enzyme production. *Mycological Research*, 98: 1019-1024.
15. Thorn, G., 1993. The use of cellulose azure agar as a crude assay of both cellulolytic and ligninolytic abilities of wood inhabiting fungi. *Proceedings of the Japanese Academy of Science*, 69: 29-34.
16. Rohrmann, S. and H.P. Molitoris, 1992. Screening for wood-degrading enzymes in marine fungi. *Canadian Journal of Botany*, 70: 2116-2123.
17. Paterson, R.R.M. and P.D. Bridge, 1994. *Biochemical techniques for filamentous fungi*. CAB International, London, UK.
18. Pointing, S.B., L.L.P. Vrijmoed and E.B.G. Jones, 1998. A qualitative assessment of lignocellulose degrading enzyme activity in marine fungi. *Botanica Marina*, 41: 293-298.
19. Sass, I.E., 1958. *Botanical Microtechnique*. Iowa State College Press, Iowa, USA.
20. Gessner, R.V., 1980. Degradative enzymes produced by salt marsh fungi. *Botanica Marina*, 23: 133-139.